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Title: Effect of molybdenum treatment on molybdenum concentration and nitrate reduction in maize seedlings

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Abstract: Since 1940 molybdenum has been known as an essential trace element in plant nutrition and physiology. It has a central role in nitrogen metabolism, and its deficiency leads to nitrate accumulation in plants. In this study, we cultivated maize seedlings (*Zea mays* L. cv. Norma SC) in nutrient solution and soil (rhizoboxes) to investigate the effect of molybdenum treatment on the absorption of molybdenum, sulphur and iron. These elements have been previously shown to play important roles in nitrate reduction, because they are necessary for the function of the nitrate reductase enzyme. We also investigated the relationship between molybdenum treatments and different nitrogen forms in maize. Molybdenum treatments were 0, 0.96, 9.6 and 96 $\mu\text{g kg}^{-1}$ in the nutrition solution experiments, and 0, 30, 90, 270 mg kg^{-1} in the rhizobox experiments.

On the basis of our results, the increased Mo level produced higher plant available Mo concentration in nutrient solution and in soil, which resulted increased concentration of Mo in shoots and roots of maize seedlings.

In addition it was observed that maize seedlings accumulated more molybdenum in their roots than in their shoots at all treatments. In contrast, molybdenum treatments did not affect significantly either iron or sulphur concentrations in the plant, even if these elements (Mo, S and Fe) play alike important roles in nitrogen metabolism. Furthermore, the physiological molybdenum level ($1 \times \text{Mo} = 0.01 \mu\text{M}$) reduced $\text{NO}_3\text{-N}$ and enhanced the $\text{NH}_4\text{-N}$ concentrations in seedlings, suggesting that nitrate reduction was more intense under a well-balanced molybdenum supply.

COVER LETTER

Motivation for the work and summarise the findings:

Molybdenum has a central role in nitrogen metabolism, and its deficiency leads to nitrate accumulation in plants, so in this study, we cultivated maize seedlings in nutrient solution and soil (rhizoboxes) to investigate the effect of molybdenum treatment on the absorption of molybdenum, sulphur and iron. These elements have been previously shown to play important roles in nitrate reduction, because they are necessary for the function of the nitrate reductase enzyme, moreover we also investigated the relationship between molybdenum treatments and different nitrogen forms in these plants. Our findings indicate that there is a strong correlation between molybdenum levels and nitrate reduction in maize seedlings, and nitrate content of these plants can be effectively reduced by supplying their physiological molybdenum demand.

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Yours sincerely,

Prof. Dr. Béla Kovács

1 **Effect of molybdenum treatment on molybdenum concentration and**
2 **nitrate reduction in maize seedlings**

3 Highlights

- 4
- 5 • Application of molybdenum (Mo) improved the absorption of Mo in
6 shoots and roots.
 - 7 • Maize seedlings accumulated more molybdenum in their roots than in
8 their shoots.
 - 9 • The absence of Mo supply resulted the accumulation of nitrate in maize
10 seedlings.
 - 11 • Physiological Mo level (0.01 μM) reduced the $\text{NO}_3\text{-N}$ and enhanced the
12 $\text{NH}_4\text{-N}$ contents.

Effect of molybdenum treatment on maize seedlings

Effect of molybdenum treatment on molybdenum concentration and nitrate reduction in maize seedlings

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Abstract

Since 1940 molybdenum has been known as an essential trace element in plant nutrition and physiology. It has a central role in nitrogen metabolism, and its deficiency leads to nitrate accumulation in plants. In this study, we cultivated maize seedlings (*Zea mays* L. cv. Norma SC) in nutrient solution and soil (rhizoboxes) to investigate the effect of molybdenum treatment on the absorption of molybdenum, sulphur and iron. These elements have been previously shown to play important roles in nitrate reduction, because they are necessary for the function of the nitrate reductase enzyme. We also investigated the relationship between molybdenum treatments and different nitrogen forms in maize. Molybdenum treatments were 0, 0.96, 9.6 and 96 $\mu\text{g kg}^{-1}$ in the nutrition solution experiments, and 0, 30, 90, 270 mg kg^{-1} in the rhizobox experiments.

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40 On the basis of our results, the increased Mo level produced higher plant
41 available Mo concentration in nutrient solution and in soil, which resulted
42 increased concentration of Mo in shoots and roots of maize seedlings.

43 In addition it was observed that maize seedlings accumulated more
44 molybdenum in their roots than in their shoots at all treatments. In contrast,
45 molybdenum treatments did not affect significantly either iron or sulphur
46 concentrations in the plant, even if these elements (Mo, S and Fe) play alike
47 important roles in nitrogen metabolism. Furthermore, the physiological
48 molybdenum level (1x Mo = 0.01 μ M) reduced NO₃-N and enhanced the NH₄-
49 N concentrations in seedlings, suggesting that nitrate reduction was more
50 intense under a well-balanced molybdenum supply.

51

52 **Keywords:** molybdenum, nitrate accumulation, nitrate reduction, nitrogen
53 metabolism

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1. Introduction

56

57 Molybdenum, a rare transition element, has for a long time been recognized
58 as an essential micronutrient for higher plants (Bortels, 1930; Arnon and Stout,
59 1939). Though required only in small amounts, it has a large role within the
60 plant system. As with other metals required for plant growth, molybdenum has
61 been used by specific plant enzymes in the process of reduction and oxidative
62 reactions (Mendel and Hänsch, 2002).

63 Molybdenum itself is not biologically active. It is, however, an integral part
64 of an organic pterin complex called the molybdenum co-factor (Moco). Moco
65 binds to the molybdoenzymes (enzymes which require molybdenum) found in
66 most higher plants (Zimmer and Mendel, 1999; Kaiser et al., 2005; Mendel and
67 Kruse, 2012; Bittner, 2014).

68 Molybdenum has been found as a cofactor in nitrate reductase, nitrogenase,
69 xanthine oxidase and sulfite oxidase. In these enzymes molybdenum has both
70 structural and catalytic functions as well as direct involvement in redox
71 reactions. It has been found to play a vital role in the nitrogen metabolism of
72 plants, including the processes of nitrogen fixation, nitrate reduction, and the
73 transportation of nitrogen compounds (Srivastava, 1997; Mendel and Schwarz,
74 1999).

75 An essential aspect of molybdenum's crucial role as a plant nutrient is the
76 part it plays in NO₃ reduction as a co-factor to nitrate reductase (NR) (Hamlin,
77 2007). Nitrate reductase is a homodimeric protein, as are other molybdenum
78 enzymes in plants. Each identical subunit is able to operate in an independent
79 way in nitrate reduction (Marschner, 1995), and each is made up of three
80 functional domains: the N-terminal domain associated with a molybdenum

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81 cofactor (Moco), the central heme domain (cytochrome b557), and the C-
82 terminal FAD domain (Mendel and Schwarz, 1999). It acts as a catalyst in the
83 first step of the NO_3^- reduction pathway, yielding NO_2^- , which in turn is further
84 reduced to NH_4^+ (Campbell, 2001; Morozkina and Zvyagilskaya, 2007).

85 The induction of nitrate reductase in plants requires both nitrate and
86 molybdenum: if either nutrient is deficient, the enzyme is either non-existent or
87 less active. In deficient plants, the induction of enzyme activity by
88 molybdenum has been found to be much faster than the induction of nitrate
89 reductase activity by nitrate (Hamlin, 2007).

90 In fact, many studies have shown that application of Mo improves the
91 absorption of Mo, the transformation of NO_3^- -N to NH_4^+ -N as well as free
92 nitrogen to albuminous nitrogen in seeds, and it increases the nitrate reductase
93 (Li-Ping et al., 2007).

94 Liu and Yang (1999) investigated the relationship between molybdenum and
95 the nitrogen metabolism of three soybean varieties in each stage of growth.
96 Five levels of molybdenum were studied. An increase in both nitrate reductase
97 activity and total N content were found in leaves and a reduction of NO_3^- -N
98 content was found with molybdenum application. In addition to this, according
99 to Vieira et al. (1998) experiment, molybdenum foliar spray (40 g ha^{-1} of Mo)
100 at 25 days after plant emergence significantly aided nitrate reductase activities,
101 producing an increase of the total nitrogen accumulated in the plant shoots of
102 common beans.

103 The nitrogen metabolism has been found to be affected by Mo-treatment in
104 several studies: an increased nitrate reductase (NR), and a decreased NO_3^-
105 content of the leaves was observed by Salcheva et al. (1979), an increase of
106 Moco leaves and dry seeds was recorded by Vunkova-Radeva et al. (1988).
107 This suggests that molybdenum directly affects the NR molecule because it
108 contains a Moco pterine domain. This domain is common for all Mo-enzymes
109 with the exception of nitrogenase (Campbell, 1988; Pelsy and Caboche, 1992).
110 Since NR is the key enzyme in inorganic nitrogen assimilation, it may be
111 assumed that the cryoprotective effect of molybdenum on NR activity is
112 reflected in the nitrate assimilatory pathway.

113 On the other hand, Calonago et al. (2010) discovered that the absence of Mo
114 foliar supply made for the accumulation of nitrate in common bean leaves: this
115 as a result of the increased nitrogen availability in the soil, which indicated the
116 inefficiency of nitrogen assimilation of plants in the absence of Mo. Srivastava
117 (1997) came to a similar conclusion, stating that in molybdenum-deficient
118 plants, nitrate-reductase activity is often reduced, which results in the buildup
119 of a high concentration of NO_3^- .

120 Furthermore, a higher concentration of total nitrogen was recorded in Mo-
121 deficient winter wheat, where Mo was seen to be the essential element for

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122 nitrate reduction (Yu et al., 2010). Mo deficiency, therefore, resulted in an
123 imbalanced nitrogen metabolism, evidenced by a much higher concentration of
124 total nitrogen and nitrate (Hu et al., 2002; Yu et al., 2006). Thus, nitrogen
125 metabolism was seen to be affected by the Mo status of a plant.

126 Nitrate accumulation in crop plants due to molybdenum deficiency might
127 have serious consequences for human health. Excess nitrate consumption can
128 increase the risk of cancer in adults and causes serious health damage
129 especially in children. It can cause methaemoglobinaemia, a type of rare but
130 potentially fatal haemoglobinopathy (Sanchez-Echaniz et al., 2001). In nitrate-
131 induced methaemoglobinaemia, dietary nitrate is reduced to nitrite in the
132 stomach, and the absorbed nitrite then converts haemoglobin to
133 methaemoglobin in red blood cells by oxidising the heme Fe^{2+} ion to Fe^{3+}
134 (Bradberry, 2012; Wright et al., 1999). This oxidation prevents
135 methaemoglobin from binding oxygen and compromises oxygen delivery to
136 peripheral tissues. Methaemoglobinaemia underlines the importance of optimal
137 nitrate reduction in crop plants, which can be achieved by providing optimal
138 molybdenum nutrition.

139 The present investigation deals with the treatment of maize seedlings with
140 molybdenum and the effect of this treatment on element contents
141 (molybdenum, iron, sulphur) and on endogenous concentrations of nitrate-,
142 nitrite- and ammonium-nitrogen in shoots and roots. The main aim of the
143 present study was to prove under laboratory circumstances that have a close
144 relation between molybdenum supply and nitrate reduction: nitrate content of
145 plants can be reduced by supporting their physiological Mo demand. To ensure
146 adequate supply of Mo, nitrate content in the leaf and root vegetables can be
147 reduced, to produce and consume healthier raw materials and foods, which are
148 essential for human health aspects.

149

150

2. Materials and methods

151

2.1 General plant propagation

152 A maize (*Zea mays* L. cv Norma SC) as a monocotyledon was chosen for
153 our research to study the contents of various elements (Mo, S, Fe) and nitrogen
154 species in roots and shoots separately (*Figure 1-2*). Disinfected maize seeds
155 were geotropically germinated between wet fluted filter papers at 22°C.
156 Seedlings with 2.5-3.0 cm coleoptiles were placed into aerated nutrient
157 solutions or rhizoboxes depending on experimental settings. Maize plants were
158 grown in a climate room under strictly regulated environmental conditions.
159 Relative humidity was maintained between 65-75%, light/dark cycle was 16/8
160 hrs with a respective 25/20°C temperature periodicity, and light intensity was
161 kept at a constant $220 \mu\text{E m}^{-2} \text{s}^{-1}$ during daytime.

162

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163

164 *2.2 Plant growth in nutrient solution*

165 The nutrient solution used for plant growth had the following composition:
166 2.0 mM Ca(NO₃)₂, 0.7 mM K₂SO₄, 0.5 mM MgSO₄, 0.1 mM KH₂PO₄, 0.1 mM
167 KCl, 0.1 μM H₃BO₃, 0.5 μM MnSO₄, 0.5 μM ZnSO₄, 0.2 μM CuSO₄. Iron was
168 supplied in the form of 10⁻⁴M Fe-EDTA (Cakmak and Marschner, 1990).

169 Molybdenum was supplemented to the nutrient solution as (NH₄)₆Mo₇O₂₄ at
170 four different concentrations as follows: Ø Mo, 1x Mo (0.96 μg kg⁻¹=0.01 μM),
171 10x Mo, 100x Mo, respectively. Nutrient solution was changed every other
172 day, and evaporated water was replenished regularly. Since nitrate reduction in
173 roots is dependent on respiration, we kept the temperature of the nutrient
174 solution at a constant 20°C. The experiment ended 9 days after planting, when
175 seedlings had approximately 12-12 cm long shoots and roots, respectively.
176 Experiments were carried out in triplicates.

177

178 *2.3 Plant growth in soil*

179 Experiments in soil were carried out in rhizoboxes, which allowed us to
180 easily monitor many aspects of root development, including overall growth,
181 circadian rhythm of the growth as well as symptoms of phytotoxicity that might
182 have been caused by increased concentrations of molybdenum. The
183 experiments used calcareous chernozem soil obtained from the Látókép
184 Experimental Station of our university. The parameters of this soil (Table 1)
185 were essentially the same as previously described by Nagy et al. (2010). No
186 additional PNK fertilization was carried out on this soil. Molybdenum was
187 supplemented to the soil as an aqueous solution prepared with distilled water at
188 four different concentrations: 0 (control), 30, 90, 270 mg kg⁻¹.

189 In order to ensure steady water uptake by plants, wet fluted filter papers
190 were placed at the bottom of rhizoboxes before the soil was added. After
191 planting the seedlings in the soil, the transparent side walls of rhizoboxes were
192 covered with black foil. The plants were geotropically stimulated to force root
193 growth along the transparent wall of the box, thus allowing convenient
194 monitoring of the roots. The mass of rhizoboxes and the length of the roots
195 were measured daily. Evaporated water was also replenished daily.

196

197 *2.4 Sample preparation and analytical methods*

198 At the end of the experiments, shoots were separated from roots. Plant parts
199 were dried at 85°C until constant weight was achieved, then cooled to room
200 temperature and weighed by an analytical scale (OHAUS). Dried samples (1±
201 0.01 g) were homogenized and decomposed by HNO₃-H₂O₂ treatment as
202 previously described (Kovács et al., 1996). Briefly, samples were kept in 10 ml
203 concentrated HNO₃ overnight, then heated to 60°C for 45 min in a LABOR

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204 MIM OE 718/A block digestion apparatus. Following the first digestion step, 3
205 ml 30% H₂O₂ was added to the samples, and digestion was continued at 120°C
206 for another 90 min. After cooling the samples to room temperature, volume was
207 adjusted to 50 ml with deionized water. Samples were then mixed by shaking
208 and filtered through FILTRAK 388 filters.

209 Element analysis was carried out by inductively coupled plasma optical
210 emission spectroscopy (ICP-OES) (Perkin Elmer OPTIMA 3300 DV) and
211 inductively coupled plasma mass spectrometry (ICP-MS) (Thermo Elemental
212 X7). In addition to molybdenum content, the concentrations of iron and sulfur
213 were also determined since nitrate reductase also requires these elements for its
214 function. Instrument settings and parameters were the same as described
215 previously (Puskás-Preszner and Kovács, 2009).

216 To analyse the amounts of different nitrogen forms in maize seedlings, 0.1 g
217 of dried and homogenized samples were weighed into centrifuge tubes. Then,
218 10 ml of 1 M KCl was added, and the test tubes were placed into an ultrasonic
219 shaker for 60 min to release N-forms. NO₃-N, NO₂-N and NH₄-N
220 concentrations of plant samples were determined by a FIAstar 5000
221 Analyzer.

222

223 *2.5 Statistical analysis*

224 Experimental data were analyzed by a bivariate general linear model
225 (GLM), which is a combination of variance and linear regression analysis.
226 Independent variables were molybdenum treatments and plant parts, while
227 dependent variables were concentrations of elements in seedlings. R-squared
228 values were calculated to assess how well independent variables predicted
229 dependent variables. Statistical analysis was done by SPSS 22.0.

230

231

3. Results

232

233 *3.1 Plant growth in nutrient solution*

234 Molybdenum treatments in nutrient solutions resulted in only a slight
235 decrease in the dry weights of shoots and roots compared to the control
236 experiment (data not shown). Therefore, molybdenum did not appear to have
237 any toxic effects even at its highest level used in our experiments.

238 When seedlings were cultivated in molybdenum-free nutrient solutions,
239 molybdenum concentration of the plants was relatively low (Table 2), most
240 likely corresponding to the original molybdenum concentration of the seeds.
241 Molybdenum concentrations increased gradually with increasing molybdenum
242 level in the nutrient solution. At 100x Mo treatment molybdenum concentration
243 increased approximately 8-fold and 11-fold in shoots and roots, respectively,
244 compared to baseline (Ø Mo) levels. Molybdenum supplementation of the

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245 nutrient solution did not cause a significant increase in sulphur concentration of
246 the seedlings. Iron concentration of shoots did not change significantly with
247 increasing molybdenum level, but iron concentration of roots doubled at 1x and
248 10x molybdenum treatments. The highest (100x Mo) treatment reduced iron
249 concentration in roots to baseline levels (Table 2). For all three elements,
250 concentrations were consistently higher in roots than in shoots. Molybdenum
251 supplementation resulted in comparable changes of these elements in shoots
252 and roots, except for iron that appeared to be preferentially accumulated in
253 roots. These data suggest that the three elements were present at optimal
254 concentrations in leaves, and transport of these elements from roots was not
255 activated at higher molybdenum levels.

256 Since molybdenum is important for nitrogen assimilation, we hypothesized
257 that molybdenum levels significantly affected nitrate, nitrite and ammonium
258 concentrations in maize plants. This effect is expected to be more prominent in
259 seedlings which require increased protein synthesis and enzyme activities to
260 support intensive vegetative growth. In order to test this hypothesis, we set out
261 to investigate nitrate, nitrite and ammonium concentration in maize seedlings
262 under different levels of molybdenum supplementation. Nitrate reduction can
263 take place in two plant parts. In leaves, the needed reducing power is supplied
264 by the light reaction of photosynthesis, while in roots, the active hydrogen ions
265 are derived from respiration. The observed higher molybdenum and iron
266 concentrations of roots (compare Table 2) suggests that nitrogen assimilation
267 might be more intense in roots than in shoots.

268 Maize seedlings grown on molybdenum deficient nutrient solution have only
269 endogenous molybdenum reserves, which would allow a baseline nitrogen
270 reductase activity. Accordingly, $\text{NO}_3\text{-N}$ concentration in shoots was relatively
271 high, and $\text{NH}_4\text{-N}$ concentration was relatively low under this condition (Table
272 3). When physiological concentration (1x Mo = 0.01 μM) of molybdenum was
273 provided, $\text{NO}_3\text{-N}$ concentration of shoots decreased, while $\text{NH}_4\text{-N}$
274 concentration was much higher compared to the baseline (\emptyset Mo) condition.
275 The 10x Mo treatment resulted in significant increase in the $\text{NO}_3\text{-N}$
276 concentration, but the $\text{NH}_4\text{-N}$ concentration also remained high. This seemingly
277 counterintuitive observation can be explained by the enhanced uptake of
278 nitrate. If $\text{NO}_3\text{-N}$ is the only nitrogen source in the medium, a high affinity
279 nitrate transporter system is activated, and this process is dependent on
280 molybdenum. The resulting high concentration of nitrate in the plant in turn
281 induces nitrate reductase, which leads to elevated $\text{NH}_4\text{-N}$ concentrations.
282 However, increasing the molybdenum level even further (100x Mo) decreased
283 $\text{NH}_4\text{-N}$ concentration of shoots to baseline level.

284 In roots, $\text{NO}_3\text{-N}$ and $\text{NH}_4\text{-N}$ concentrations were higher than in shoots even
285 in molybdenum-free nutrient solution (Table 3). This observation could

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286 indicate that in addition to the high affinity nitrate uptake mechanism,
287 considerable amounts of nitrate were taken up by alternative mechanisms, such
288 as ATP-dependent symport through the plasma membrane of root hair cells.
289 The high $\text{NH}_4\text{-N}$ concentrations in roots could be the consequence of higher
290 molybdenum concentrations in roots compared to shoots. In fact, the rate of
291 nitrogen assimilation (estimated from $\text{NH}_4\text{-N}/\text{NO}_3\text{-N}$ ratios) was approximately
292 2-3 times higher in roots than in shoots regardless of the molybdenum
293 concentration in the nutrient solution, and this difference correlated well with
294 the difference in the molybdenum concentration of the plant parts (Table 2).
295 Therefore, roots appear to be more active in nitrogen assimilation than shoots
296 in maize seedlings.

297 Statistical analysis using the general linear model (Table 4 and 5) indicated
298 that molybdenum treatment and plant part had 92-99% effect on the Mo, S and
299 Fe concentrations of maize seedlings. Molybdenum treatment had a greater
300 effect on molybdenum concentration than plant part had. For sulphur and iron,
301 the effect of plant part was one order of magnitude higher than that of
302 molybdenum treatment. Molybdenum treatment had no statistically significant
303 effect on $\text{NO}_3\text{-N}$ and $\text{NO}_2\text{-N}$ concentrations of the seedlings, which could be
304 the consequence of the relatively large standard deviation within the groups.
305 The concentrations of these two nitrogen forms were primarily determined by
306 the plant part. On the other hand, the concentration of $\text{NH}_4\text{-N}$ was influenced
307 by plant part and molybdenum treatment together, which accounted for 95.7%
308 of this nitrogen form, although the effect of plant part was somewhat more
309 dominant. We have to note, that during the statistical analysis of the $\text{NO}_2\text{-N}$
310 concentration, the variance analysis model did not show significance ($R^2 =$
311 0.411).

312

313 *3.2 Plant growth in soil*

314 Dry weights of the shoot and root were 0.303 g and 0.244 g, respectively,
315 when the maize seedling was grown in rhizobox without molybdenum
316 treatment. Supplementation of the soil with 30 mg kg^{-1} molybdenum caused a
317 significant increase in the dry weights of the shoot (175%) and root (202%).
318 However, further increase in the soil's molybdenum level decreased the dry
319 weights of both plant parts. The 90 and 270 mg kg^{-1} molybdenum treatments
320 also decreased the dry weight ratio of shoots and roots compared to the control
321 experiment, suggesting that these treatments inhibited the growth of the shoots
322 more than the growth of the roots.

323 Maize seedling grown in rhizobox took up only a small amount of
324 molybdenum from control soil. Molybdenum concentrations of the root and
325 shoot were very low under these conditions, but increased gradually when the
326 soil was supplemented with increasing levels of molybdenum (Figure 3).

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327 Although we used only one seedling per molybdenum treatment in the
328 experiment, the observed trend was very similar to that of the nutrient solution
329 experiments (Table 2), supporting the validity of our results.

330 Molybdenum treatment also affected the concentrations of different nitrogen
331 forms in maize seedlings grown in rhizoboxes. In shoots, $\text{NO}_3\text{-N}$ concentration
332 varied and was highest when the soil had the lowest and highest molybdenum
333 content. $\text{NH}_4\text{-N}$ concentration rose with increasing molybdenum level of the
334 soil, indicating definite connection between molybdenum treatment and
335 nitrogen assimilation in maize shoots. In roots, molybdenum treatments
336 increased $\text{NH}_4\text{-N}$ and reduced $\text{NO}_3\text{-N}$ concentration, which suggested intensive
337 nitrate reductase activity in this plant part (Table 6).

338 We have to note that both shoots and roots demonstrated similar ammonia
339 concentrations in our rhizobox experiments, implying that nitrate reductase
340 activities were comparable in these plant parts (Table 6).

341

342

4. Discussion

343

344 In this study we have found that molybdenum nutrition significantly affected
345 molybdenum concentrations and the concentrations of different nitrogen forms
346 in maize seedlings when they were cultivated either in nutrient solution or soil.

347 Seedlings grown in molybdenum-free solution contained very small
348 amounts of molybdenum, most likely reflecting the molybdenum reserves
349 derived from the seeds. Molybdenum treatments significantly increased
350 molybdenum concentrations in both the shoots and roots of the seedlings, with
351 roots having consistently higher concentrations than shoots at all treatments
352 (including the molybdenum-free conditions). Molybdenum treatments did not
353 affect either iron or sulphur concentrations significantly, indicating that
354 molybdenum was not necessary for the absorption of these elements. Maize
355 seedlings grown in rhizoboxes showed similar absorption trends of
356 molybdenum than those grown in nutrient solution. Although molybdenum-free
357 conditions could not be established in these experiments, due to endogenous
358 amounts of this element in the soil, molybdenum uptake from the control soil
359 was relatively low. Similar to the nutrition solution experiments, seedlings
360 grown in rhizoboxes accumulated more molybdenum in their roots than in their
361 shoots.

362 Kádár (1995) obtained similar data in his microelement-load experiments in
363 calcareous chernozem soil obtained from Nagyhöröcsök. According to his
364 results, increased molybdenum load caused extremely high molybdenum
365 accumulation in maize, but there were only small differences (with the
366 exception of control soil) between molybdenum concentrations of shoots and
367 roots. The ratios of molybdenum in roots vs shoots at different soil

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368 molybdenum concentration were, 10, 1.30, 1.60 and 1.26, while in our
369 experiments under the same conditions, they were 1.17, 2.98, 2.25 and 2.07,
370 respectively (molybdenum treatments were the same in both sets of
371 experiments). This comparison suggests that molybdenum transport from roots
372 to shoots was somewhat less efficient in our experiments.

373 The consistently higher molybdenum concentration in roots could indicate
374 that this plant part was more active in nitrogen assimilation, due to the fact that
375 molybdenum is essential for the stability and activity of the nitrate reductase
376 enzyme. Although we did not determine the activity of this enzyme directly, we
377 assessed nitrogen assimilation in maize seedlings by measuring the
378 concentrations of different nitrogen forms after molybdenum treatments. As
379 expected, physiological molybdenum treatment (1x Mo = 0.01 μ M) reduced
380 $\text{NO}_3\text{-N}$ and increased the $\text{NH}_4\text{-N}$ concentrations in seedlings, suggesting that
381 nitrate reduction was more intense under a well-balanced molybdenum supply.
382 These data clearly suggest that there is a close relationship among molybdenum
383 levels, nitrate absorption, nitrate reduction and overall nitrogen assimilation.
384 Large proportion of the absorbed nitrate is reduced to ammonia which links to
385 glutamate and subsequently, through transamination reactions, is utilized for
386 the synthesis of other amino acids and proteins. Our measured ammonia
387 concentrations most likely do not reflect the actual nitrate reductase activity
388 due to fixation of ammonia by biochemical reactions mentioned above.
389 Ammonia is cytotoxic, therefore its concentration must be tightly controlled. It
390 acts as an uncoupling agent in thylakoid membranes and causes the
391 depolarization of these membranes without the synthesis of ATP. Since our
392 plants did not show toxic symptoms, we assume that sufficient amount of
393 glutamate was available to fix ammonia. In addition, the citric acid cycle that
394 provides the carbon skeleton of glutamate must be also active and efficient to
395 support the glutamate demand of the plant. Therefore, our data suggest that
396 molybdenum has an essential role in the intensive overall metabolism of maize
397 seedlings. We have to note, however, that molybdenum treatment increased
398 plant $\text{NH}_4\text{-N}$ concentrations only up to the 10x Mo treatment. The
399 concentration of this nitrogen form significantly dropped after 100x Mo
400 treatments, implying that this molybdenum concentration might have inhibited
401 nitrate reductase activity. These results differ from those of Kádár et al. (2000)
402 who found a definite increase in shoot $\text{NO}_3\text{-N}$ concentration after ammonium
403 paramolybdenate load. They hypothesized, that the $\text{NH}_4\text{-N}$ was nitrified by the
404 end of their experiment, which contributed to the elevated concentrations on
405 nitrate in shoots.

406 It was proved by our experiments, when Mo was added to nutrient solution
407 the free nitrate concentration was decreased. In natural conditions, when the
408 Mo is in low amount in soil, the nitrate accumulation can be effectively

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409 minimized in the plants. Molybdenum has a physiological importance as the
410 part of enzymes having outstanding role in nitrogen metabolism. Whereas the
411 nitrogen is in reduced form in the organic chains, the reduction of nitrate needs
412 lots of energy, which means high electric potential. Mo plays a role in the
413 electron transport chain due to its different oxidation states, therefore it takes
414 part in the nitrogen metabolism, moreover Mo can not be substituted by any
415 other element in this physiological process.

416 Overall, our results demonstrate a correlation between molybdenum
417 nutrition and nitrogen assimilation. In the absence of molybdenum, nitrate
418 reduction slows down, which results in nitrate accumulation in plants. On the
419 other hand, adequate molybdenum supply in soils can ensure a reduced nitrate
420 content of leafy and root vegetables and in general, all fresh-cut agricultural
421 products. However, it is important to note, that nitrate content of fresh
422 vegetables is influenced by other environmental factors such as light conditions
423 and the length of the daylight. Therefore, plants grown in greenhouses might
424 have nitrate concentrations a magnitude higher than those grown in fields.

425 When planning molybdenum enrichment of soils, the potentially hazardous
426 consequences of this treatment should also be taken into account. Although
427 excessive molybdenum accumulation does not lead to metabolic problems or
428 phytotoxic effects in plants, the same high molybdenum concentration can
429 cause molybdenosis in animals. The risk of this disease significantly increases
430 when molybdenum content is over 5 mg kg⁻¹, and this detrimental effect must
431 be considered when feeding farm animals with plants grown on molybdenum
432 enriched soils.

Contribution

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434
435 Conceived and designed the experiments: Béla Kovács, Anita Puskás-Preszner,
436 László Lévai and Éva Bódi. Performed the experiments: Béla Kovács, Anita
437 Puskás-Preszner, László Lévai and Éva Bódi. Analyzed the data: Béla Kovács,
438 László Huzsvai and Éva Bódi. Contributed reagents/materials/analysis tools:
439 László Lévai and Béla Kovács. Wrote the paper: Béla Kovács, László Huzsvai
440 and Éva Bódi.

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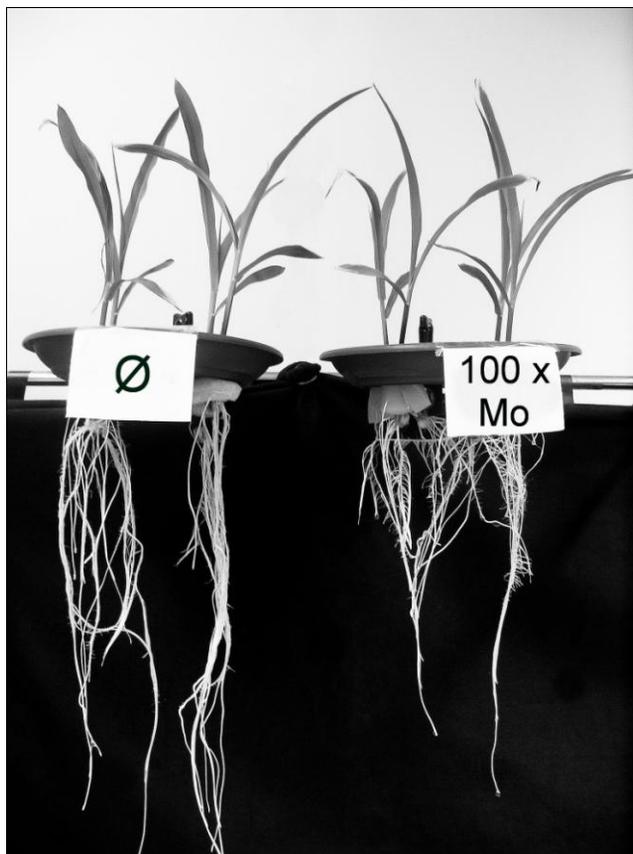
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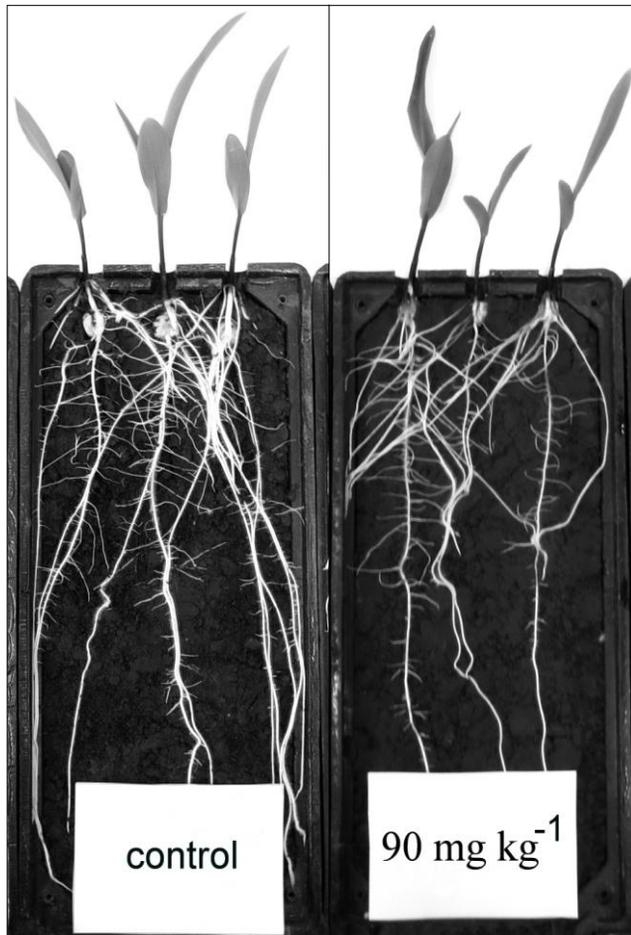
541

542 **Fig. 1** Maize seedlings grown in nutrient solution ($\emptyset=0 \text{ mg dm}^{-3} \text{ Mo}$, $100x \text{ Mo}=1 \text{ }\mu\text{M}$

543 $(\text{NH}_4)_6\text{Mo}_7\text{O}_{24}$)

544

Effect of molybdenum treatment on maize seedlings



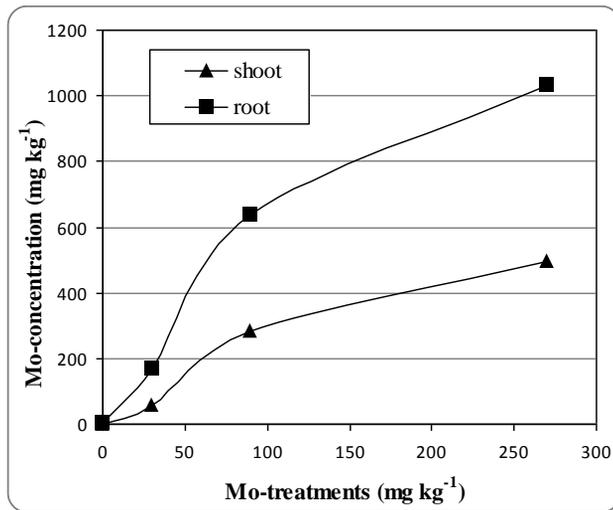
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546 **Fig. 2** Maize seedlings grown in rhizoboxes (control, 90 mg kg⁻¹ Mo)

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Effect of molybdenum treatment on maize seedlings

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551 **Fig. 3** Molybdenum concentration (mg kg⁻¹ dry weight) of shoots and roots of maize seedlings

552 grown in rhizoboxes

553

554

Effect of molybdenum treatment on maize seedlings

555 **Table 1** Parameters of soil applied in the experiments carried out in rhizoboxes

556		
557	Depth	0-0.3 m
558	pH (KCl)	5.71
559	pH (H ₂ O)	6.58
560	Soil texture category	loamy clay
561	Total water-soluble salt	0.015 %
562	CaCO ₃	0.202 %
563	Humus	3.54 %
564	KCl-soluble NO ₃ -N+NO ₂ -N	8.04
565	AL-soluble P ₂ O ₅	199 mg kg ⁻¹
566	AL-soluble K ₂ O	451 mg kg ⁻¹
567	AL-soluble Na	332 mg kg ⁻¹
568	KCl-soluble Mg	176 mg kg ⁻¹
569	KCl-soluble SO ₄ ²⁻ -S	6.04 mg kg ⁻¹
570	KCl-EDTA-soluble Cu	5.79 mg kg ⁻¹
571	KCl-EDTA-soluble Zn	7.9 mg kg ⁻¹
572	KCl-EDTA-soluble Mn	262 mg kg ⁻¹
573		

Effect of molybdenum treatment on maize seedlings

574 **Table 2** Mo, S and Fe concentration (mg kg^{-1} dry weight) of shoots and roots of maize
575 seedlings grown in nutrient solution in case of \emptyset Mo (molybdenum free solution), 1x Mo (0.01
576 μM), 10x Mo and 100x Mo treatments
577

Plant part	Mo-treatment	Mo	S	Fe
Shoot	\emptyset Mo	2.52 \pm 0.17	2049 \pm 148	60.9 \pm 6.4
	1x Mo	3.06 \pm 0.08	2152 \pm 210	57.0 \pm 10.3
	10x Mo	6.08 \pm 0.40	2359 \pm 84	58.3 \pm 1.6
	100x Mo	20.6 \pm 0.50	2367 \pm 142	72.2 \pm 25.5
Root	\emptyset Mo	5.78 \pm 0.12	4644 \pm 145	167 \pm 14
	1x Mo	7.89 \pm 0.46	4970 \pm 425	303 \pm 64.5
	10x Mo	17.9 \pm 0.1	5197 \pm 634	331 \pm 38
	100x Mo	66.5 \pm 1.4	5076 \pm 439	187 \pm 26

578

Effect of molybdenum treatment on maize seedlings

579 **Table 3** Ammonium- ($\text{NH}_4\text{-N}$), nitrite- ($\text{NO}_2\text{-N}$), nitrate-nitrogen ($\text{NO}_3\text{-N}$) concentration (mg
580 kg^{-1} dry weight) of shoots and roots of maize seedlings grown in nutrient solution in case of \emptyset
581 Mo (molybdenum free solution), 1x Mo (0.01 μM), 10x Mo and 100x Mo treatments
582

Plant part	Mo-treatment	$\text{NH}_4\text{-N}$	$\text{NO}_2\text{-N}$	$\text{NO}_3\text{-N}$
Shoot	\emptyset Mo	111 \pm 55	0.59 \pm 0.16	2153 \pm 553
	1x Mo	199 \pm 142	1.06 \pm 0.29	1730 \pm 78
	10x Mo	160 \pm 39	1.42 \pm 1.20	3939 \pm 1427
	100x Mo	124 \pm 2	0.73 \pm 0.23	1772 \pm 1086
Root	\emptyset Mo	703 \pm 84	2.28 \pm 0.52	4407 \pm 180
	1x Mo	914 \pm 23	2.27 \pm 0.89	4611 \pm 221
	10x Mo	534 \pm 6	2.01 \pm 0.38	4934 \pm 820
	100x Mo	1075 \pm 125	2.00 \pm 1.73	5055 \pm 1109 \pm

583

Effect of molybdenum treatment on maize seedlings

584 **Table 4** Variance analysis of maize seedlings grown in nutrient solution, dependent variables is
 585 the concentration of Mo, S and Fe (mg kg⁻¹ dry weight)
 586

Dependent Variable	Source	df	Mean Square
Mo	Corrected Model	7	1368.147***
	Intercept	1	6369.618***
	Plant parts	1	1624.619***
	Mo-treatments	3	2051.483***
	Plant parts * Mo-treatments	3	599.320***
	Error	16	0.318
	Total	24	
	Corrected Total	23	
S	Corrected Model	7	6538590.167***
	Intercept	1	311371288.167***
	Plant parts	1	45040120.167***
	Mo-treatments	3	224434.500
	Plant parts * Mo-treatments	3	18902.500
	Error	16	111250.667
	Total	24	
	Corrected Total	23	
Fe	Corrected Model	7	38061.528***
	Intercept	1	573566.002***
	Plant parts	1	205313.002***
	Mo-treatments	3	9069.116***
	Plant parts * Mo-treatments	3	11303.449***
	Error	16	910.100
	Total	24	
	Corrected Total	23	

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Effect of molybdenum treatment on maize seedlings

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Table 5 Variance analysis of maize seedlings grown in nutrient solution, dependent variables is the concentration of ammonium-nitrogen (NH₄-N), nitrite- (NO₂-N) and nitrate- (NO₃-N) of maize (mg kg⁻¹ dry weight)

Dependent Variable	Source	df	Mean Square
NH₄-N	Corrected Model	7	445086.614***
	Intercept	1	5476796.106***
	Plant parts	1	2596505.005***
	Mo-treatments	3	86111.718***
	Plant parts * Mo-treatments	3	86922.045***
	Error	16	6031.260
	Total	24	
	Corrected Total	23	
NO₂-N	Corrected Model	7	1.411
	Intercept	1	57.645***
	Plant parts	1	8.431**
	Mo-treatments	3	0.174
	Plant parts * Mo-treatments	3	0.308
	Error	16	0.885
	Total	24	
	Corrected Total	23	
NO₃-N	Corrected Model	7	6262698.586***
	Intercept	1	306727900.091***
	Plant parts	1	33228744.261***
	Mo-treatments	3	2037619.581
	Plant parts * Mo-treatments	3	1499095.699
	Error	16	689026.543
	Total	24	
	Corrected Total	23	

Effect of molybdenum treatment on maize seedlings

592 **Table 6** Ammonium- ($\text{NH}_4\text{-N}$), nitrite- ($\text{NO}_2\text{-N}$), nitrate-nitrogen ($\text{NO}_3\text{-N}$) concentration (mg
593 kg^{-1} dry weight) of shoots and roots of maize seedlings grown in rhizoboxes in case of different
594 molybdenum treatments (mg kg^{-1})
595

Plant part	Mo-treatment	$\text{NH}_4\text{-N}$	$\text{NO}_2\text{-N}$	$\text{NO}_3\text{-N}$
Shoot	control	237	0.424	41.5
	30	331	0.681	0.736
	90	308	0.102	8.44
	270	401	1.51	214
Root	control	280	0.040	35.5
	30	334	0.040	15.8
	90	333	0.459	15.8
	270	200	2.84	134

596

Contribution

Conceived and designed the experiments: Béla Kovács, Anita Puskás-Preszner, László Lévai and Éva Bódi. Performed the experiments: Béla Kovács, Anita Puskás-Preszner, László Lévai and Éva Bódi. Analyzed the data: Béla Kovács, László Huzsvai and Éva Bódi. Contributed reagents/materials/analysis tools: László Lévai and Béla Kovács. Wrote the paper: Béla Kovács, László Huzsvai and Éva Bódi.