


REVIEW ARTICLE

Gynecology

Trichomonas vaginalis infection is associated with increased risk of cervical carcinogenesis: A systematic review and meta-analysis of 470 000 patients

Balázs Hamar^{1,2}  | Brigitta Teutsch^{1,3} | Eszter Hoffmann^{1,2} | Péter Hegyi^{1,3,4} | Alex Váradi¹ | Péter Nyirády^{1,5} | Zsombor Hunka^{1,2} | Nándor Ács^{1,2} | Balázs Lintner^{1,2} | Réka Juhász Hermánne¹ | Zsolt Melczer^{1,2}

¹Center for Translational Medicine, Semmelweis University, Budapest, Hungary

²Department of Obstetrics and Gynecology, Semmelweis University, Budapest, Hungary

³Institute for Translational Medicine, Medical School, University of Pécs, Pécs, Hungary

⁴Institute of Pancreatic Diseases, Semmelweis University, Budapest, Hungary

⁵Department of Urology, Semmelweis University, Budapest, Hungary

Correspondence

Balázs Hamar, HU-1088, Budapest, Baross utca 27, Hungary.

Email: hamar.balazs@med.semmelweis-univ.hu

Abstract

Background: *Trichomonas vaginalis* infection is the most prevalent non-viral sexually transmitted infection (STI) in women and has been suggested as a risk factor for developing cervical cancer.

Objective: We aimed to investigate the associations between *T. vaginalis* infection and cervical carcinogenesis.

Search Strategy: A comprehensive systematic search was conducted in five databases on 21 October 2021.

Selection Criteria: Studies assessing the relationship between *T. vaginalis* infection, HPV co-infections, cervical dysplasia, and cervical cancer were found eligible.

Data Collection and Analysis: Summary estimates for pooled odds ratios (ORs) and their 95% confidence intervals (CIs) were calculated with a random-effects model. Statistical heterogeneity was measured with I^2 and Cochran's Q tests.

Main Results: The 29 articles included 473 740 women, of whom 8518 were *T. vaginalis*-positive. Our results showed that *T. vaginalis*-infected women had 1.79 times higher odds of being diagnosed with HPV co-infection (95% CI 1.27–2.53; I^2 95%). We also found that *T. vaginalis* infection was associated with high-grade squamous intraepithelial lesion diagnosis (OR 2.34, 95% CI 1.10–4.95; I^2 75%) and cervical cancer (OR 5.23, 95% CI 3.03–9.04; I^2 3%).

Conclusions: Our results showed an association between *T. vaginalis* and cervical carcinogenesis in sexually active women.

KEYWORDS

cervical intraepithelial neoplasia, cervical lesion, cervical precancer, protozoal infection

This is an open access article under the terms of the [Creative Commons Attribution](https://creativecommons.org/licenses/by/4.0/) License, which permits use, distribution and reproduction in any medium, provided the original work is properly cited.

© 2023 The Authors. *International Journal of Gynecology & Obstetrics* published by John Wiley & Sons Ltd on behalf of International Federation of Gynecology and Obstetrics.

1 | INTRODUCTION

Cervical cancer is the fourth most commonly diagnosed cancer and the fourth leading cause of cancer-related death in women.¹ Currently, there are effective ways of fighting cervical cancer through immunization, screening, and oncologic treatment.² HPV vaccination provides a high level of protection against oncogenic HPV strains and can reduce the burden of cervical cancer. Moreover, cervical smear and HPV tests improve the reliability of cervical cancer screening significantly.³ In the treatment line of cervical cancer, immunotherapy and target therapy can have an increasing role besides the classical chemotherapeutic regimens.⁴ Despite all of these, cervical cancer is still the most frequently diagnosed cancer in developing countries, and the leading cause of cancer-related mortality in emergent nations.^{1,2}

The main risk factor of cervical cancer is infection with high-risk HPV types, responsible for various cancer types. Most notably, HPV 16 and HPV 18 types are accountable for 70% of cervical cancers worldwide.⁵ After being incorporated into the host cell genome, the virus leads to overexpression of proto-oncogene proteins.^{6,7} A persistent HPV infection and the inability of the immune system to clear out the infection in the cervix are key elements of the carcinogenesis.⁵ The disruption of the vaginal microbiota performs an essential role in persistent HPV infection, as vaginal dysbiosis occurs, proinflammatory cytokines are increased and immunoclearance is reduced.⁸ It is well known that other risk factors, including smoking, promiscuity, using oral contraceptive drugs, immunosuppressed state, and sexually transmitted infections (STIs), can also contribute to developing cervical cancer.^{7,9-11}

Trichomonas vaginalis, a common cause of STI, causes around 170–190 million infections annually.¹² Infection of the genital tract with these anaerobic protozoa can lead to discomfort by causing odorous discharge, dysuria, itching, and vulvar irritation. However, up to 85% of trichomoniasis can be symptomless in women. Moreover, 5%–35% of women can also be reinfected.¹³ *Trichomonas vaginalis* can contribute to the development of cervical cancer by causing inflammation, abruption of the cervical epithelium, and influencing the immune system to eliminate HPV. Current evidence on the relationship between *T. vaginalis* infection, cervical dysplasia, and cervical cancer is conflicting. Although several articles report strong associations, other publications do not find *T. vaginalis* to be a risk factor for cervical carcinogenesis.¹⁴⁻¹⁷ Two meta-analyses have been conducted on this topic. The first article, published in 1994, included populations where *T. vaginalis* detection was based only on cytology, which can often underdetect *T. vaginalis*.¹⁸⁻²⁰ The other meta-analysis focused on cervical dysplasia without differentiating between the different states of cervical lesions and did not investigate the relationship between *T. vaginalis* and HPV.²¹

Hence, on the basis of the available literature, this study aimed to investigate the association between *T. vaginalis* and HPV, cervical dysplasia, and carcinogenesis. We hypothesized that *T. vaginalis* was a risk factor for developing cervical cancer.

2 | MATERIALS AND METHODS

We conducted our systematic review and meta-analysis according to the PRISMA 2020 and MOOSE guidelines (see [Figure 1](#); [Tables S1](#) and [S2](#)) while we followed the recommendations of the Cochrane Handbook.²²⁻²⁴ The pre-study protocol was registered in PROSPERO (CRD42021286097), and we fully adhered to it.

2.1 | Literature search and eligibility criteria

The systematic search was conducted using five major databases on October 20, 2021: MEDLINE (via PubMed), Embase, Cochrane Central Register of Controlled Trials (CENTRAL), Scopus, and Web of Science. We accepted only peer-reviewed articles; therefore, we did not search on ClinicalTrials.gov, nor did our preliminary search find any suitable studies. No filters or restrictions were applied during the search. We used two population, exposure, and outcome (PEO) frameworks to define the eligibility criteria for the articles.²⁵ All studies reporting sexually active (P_1) or HPV-positive (P_2) women who were screened for *T. vaginalis* infection (E) were deemed eligible. The outcomes of interest (O_1) were HPV positivity, cervical dysplasia, and cervical cancer. In HPV-positive women (P_2), the investigated outcomes (O_2) were cervical dysplasia and cervical cancer. The articles had to include a population of *T. vaginalis*-negative women forming the control group.

Articles were considered where *T. vaginalis* was detected with cytology, wet-mount, culture, or polymerase chain reaction (PCR) methods. Articles were excluded in which *T. vaginalis* was diagnosed on the basis of clinical features or medical history. Studies were suitable if HPV exposure was diagnosed with any nuclear amplification method. Articles where HPV was detected only by cytology were excluded because of the low sensitivity of the method.²⁶ Cytologic and histopathologic diagnoses were acceptable for cervical intraepithelial neoplasia (CIN) and cancer confirmation. We evaluated the following outcomes in the dysplasia group: atypical squamous cells of undetermined significance (ASCUS), atypical glandular cells, atypical squamous cells for which one could not rule out high-grade squamous intraepithelial lesions (ASC-H), low-grade squamous intraepithelial lesions (LSIL), and high-grade squamous intraepithelial lesions (HSIL). For cytologic samples, the Bethesda classification was required. Articles that proclaimed CIN1–3 diagnoses were divided into LSIL (CIN1) and HSIL (CIN2–3) groups for a more straightforward interpretation.

Observational studies, such as cross-sectional, case-control, and cohort analyses, were accepted. Abstracts were excluded in our review. Non-English language articles were translated for possible evaluation.

2.2 | Search strategy

During the systematic search, we used the following main concepts: “trichomonas”, “human papillomavirus”, “cervical intraepithelial

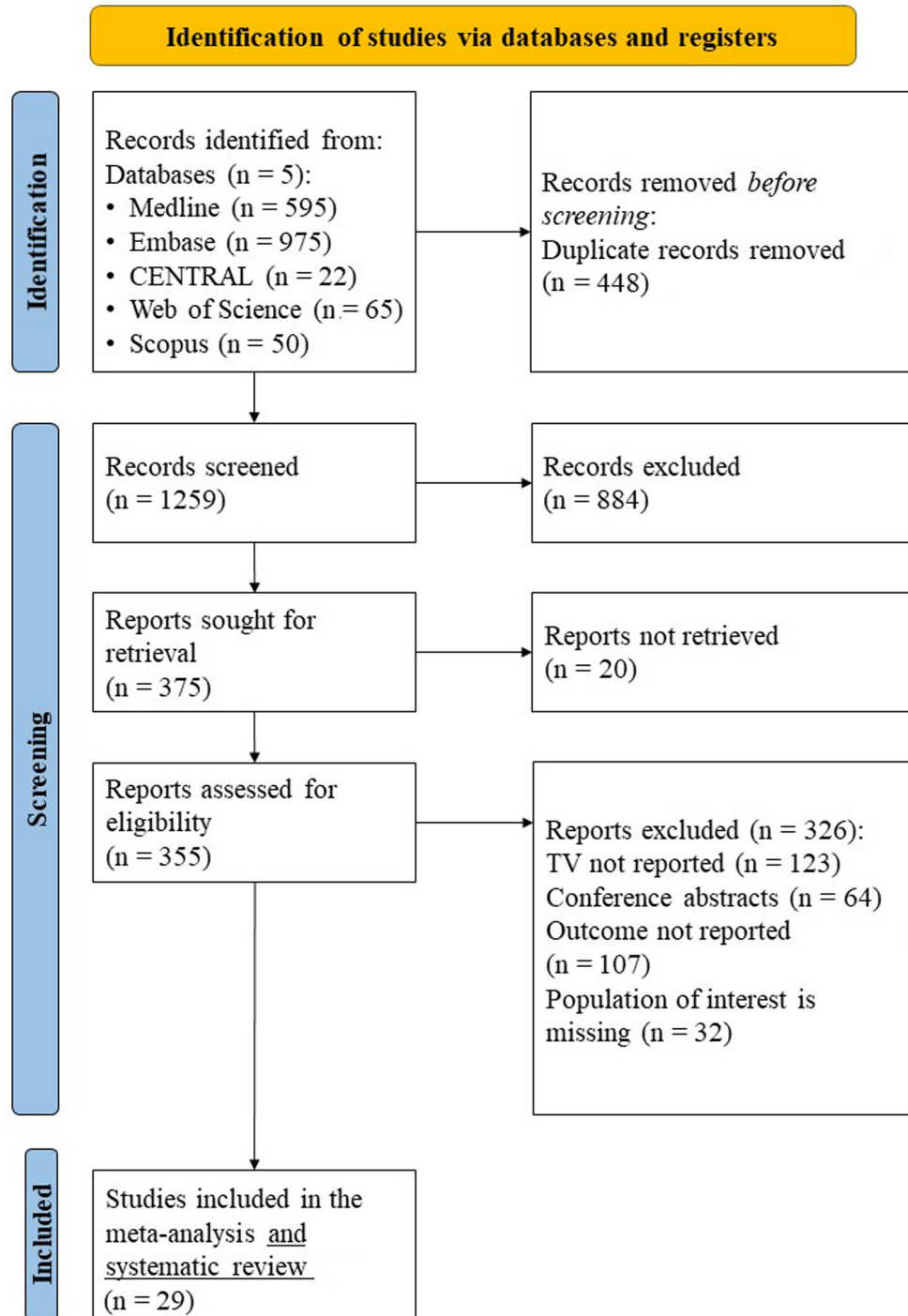


FIGURE 1 PRISMA 2020 flowchart representing the study selection process.

neoplasia”, and “cervical cancer”. The whole search key can be found in [Supporting Information](#).

2.3 | Study selection and data collection

A reference management program (ENDNOTE X9) was used to select the articles. First, a duplicate removal was performed, then, two independent reviewers (BH, EH) carried out a title and abstract

selection and then full-text selection. Cohen's κ coefficient measured the degree of agreement.²⁷ A third independent investigator (ZSH) agreed on debated articles. If we could not find an article, or data were missing, we contacted the authors.

Two independent reviewers (BH, EH) extracted variables from the eligible studies into a pre-defined Microsoft EXCEL spreadsheet (Windows 11 Pro). The following variables were collected from each article: first author, publication year, digital object identifier, study design, study type, demography (age, sample size), country,

centers, the detection method of *T. vaginalis*, HPV, and cytologic/histologic lesions. Where possible, data regarding the outcomes were extracted in two-by-two tables. Otherwise, we collected the unadjusted odds ratios (ORs). In order to handle confounding factors, when possible, we collected adjusted ORs, and the variables for these results were adjusted. In case of any disagreement, a consensus was reached involving a third investigator (ZSH).

2.4 | Risk of bias and quality assessment of the included articles

To critically assess the outcome data, we performed a risk of bias assessment with the help of the Quality in Prognostic Studies (QUIPS) tool.²⁸ The QUIPS tool includes six domains: study participation, study attrition, prognostic factor measurement, outcome measurement, study confounding, and statistical analysis reporting. In each domain, four classifications can be given: not applicable, low risk, moderate risk, and high risk of bias. To grade the level of evidence of our findings, we implemented the Grades of Recommendation, Assessment, Development, and Evaluation (GRADE) approach. The Summary of Findings table was prepared by using the GRADEPRO tool.²⁹ Both QUIPS and GRADE were performed by two independent reviewers (BH, EH), and in case of disagreement, a third investigator resolved the dispute (ZSH).

2.5 | Synthesis methods

During data synthesis, both qualitative and quantitative assessments were carried out. The R programming language was used for statistical data analysis (R Core Team, 2022; R version 4.2). The minimum number of studies for performing the quantitative synthesis was three. Forest plots were used to visualize individual studies and overall results. Subgroup analyses were performed based on the detection method of *T. vaginalis* and the country of origin of the article. Sensitivity analyses were carried out for four outcomes.

Where possible, we estimated pooled ORs with 95% confidence intervals (CIs) with a random-effects model, using the `tel-Haenszel` method with the `metabin` function from `META v5.50` package, and we applied the Paule-Mandel method to estimate the between-study variance.³⁰⁻³² Statistical significance was substantiated for a result when $P < 0.05$. I^2 and Cochran's Q tests were used to measure statistical heterogeneity, where $P < 0.1$ indicated significant heterogeneity.²⁸ A general interpretation of the heterogeneity values is as follows: 0%–40% possibly not important heterogeneity; 30%–60% moderate heterogeneity; 50%–90% substantial heterogeneity; and 75%–100% considerable heterogeneity. Beside I^2 , we also reported the prediction intervals (i.e. the expected range of effects of future studies) of the pooled estimates if the minimum study number was reached.³³

The inspection of funnel plots and an Egger's test were used to assess publication bias when a minimum of 10 articles were available for one outcome.

3 | RESULTS

3.1 | Search and selection

Our comprehensive search identified 1707 articles. After duplicate removal, 1259 publications were screened based on title and abstract. During the full-text selection, 355 articles were screened, resulting in 29 eligible studies for the quantitative and qualitative data syntheses. Cohen's κ was 0.9 for title and abstract selection; and 0.85 for full-text selection. Despite contacting authors for non-retrievable articles, we received only a few responses.

3.2 | Basic characteristics of included studies

The eligible articles were published between 2009 and 2021, with 11 publications from Asia, five from Europe, seven from South America, five from Africa, and one from North America. According to study type, we found 22 cross-sectional, five case-control, and one prospective cohort study.

As for demographics, the mean age of women was 37.57 years. In 15 articles, *T. vaginalis* was detected with PCR, in eight with wet-mount, in four with cytology and in two with cultures and wet-mount. All the studies assessed the exposure and the outcome at the same time.

Altogether 473 740 women were included in our meta-analysis. Of them, 8518 patients had *T. vaginalis* infection in the exposure group. Baseline characteristics of the eligible studies are detailed in [Table 1](#).

3.3 | Quantitative and qualitative analysis

The association between *T. vaginalis* and HPV infections

Twenty-four studies including 7291 women in the *T. vaginalis*-infected group and 452 161 in the control group reported an association between *T. vaginalis* and HPV infections.^{14,16,17,34-54} Our results showed that *T. vaginalis*-positive women were 1.79 times more likely to be diagnosed with an HPV co-infection (95% CI 1.27–2.53; I^2 95%; [Figure 2](#)) compared with *T. vaginalis*-negative women.

When a *T. vaginalis* infection was confirmed with the wet-mount method, the odds of detecting a co-infection with HPV were slightly higher, by the odds of 2.29 (95% CI 1.23–4.28; I^2 97%). The results from the subgroups based on region showed that *T. vaginalis*-positive women from Asia had the highest chance for HPV co-infection (OR 2.05, 95% CI 1.08–3.88; I^2 97%; see [Figure S1](#)). A sensitivity analysis (leave-one-out method) did not recognize any influential study (see [Figure S2](#)).

In one article,⁵⁴ a multivariate analysis showed 2.29 odds (95% CI 1.46–3.60) for the diagnosis of HPV in the case of *T. vaginalis*

TABLE 1 Basic characteristics of included studies.

Author, year	Study type	Region	Number of patients	Mean age, y	Diagnosis of <i>Trichomonas vaginalis</i>	Diagnosis of HPV	Diagnosis of cervical lesion	Cervical lesion-related outcomes
Verteramo et al., 2009 ³⁴	Cross-sectional	Europe	860	32.7	Culture and wet-mount	PCR	NA	NA
Noel et al., 2010 ³⁵	Case-control	Europe	507	<30–50 ^a	Cytology	HCII	NA	NA
Depuydt et al., 2010 ³⁶	Cross-sectional	Europe	62 944	42	PCR	PCR	NA	NA
Caiyan et al., 2012 ³⁷	Cross-sectional	Asia	6339	39.2	Wet-mount	HCII	Histology	LSIL, HSIL
Donders et al., 2013 ³⁸	Cross-sectional	Europe	63251	NA	PCR	PCR	Cytology	ASCUS, LSIL, HSIL
Mendoza et al., 2013 ³⁸	Cross-sectional	South America	181	30 ^b	Culture and wet-mount	PCR	NA	NA
Paesi et al., 2013 ³⁹	Cross-sectional	South America	208	13–69 ^a	Cytology	PCR	NA	NA
Lazenby et al., 2014 ¹⁴	Cross-sectional	Africa	324	38	PCR	HCII	Cytology/histology	LSIL, HSIL
Liu et al., 2015 ⁴⁰	Cross-sectional	Asia	429	39	Wet-mount	PCR	NA	NA
Casillas-Vega et al., 2016 ⁴¹	Cross-sectional	South America	662	31	PCR	PCR	NA	NA
Camporiondo et al., 2016 ⁴²	Cross-sectional	Europe	309	49 ^b	PCR	PCR	NA	NA
Dey et al., 2016 ⁵⁵	Cross-sectional	Asia	7962	NA	Cytology	NA	Cytology	ASCUS, LSIL, HSIL
de Abreau et al., 2016 ⁵⁷	Cross-sectional	South America	685	40.3	PCR	NA	Cytology/histology	HSIL
Kim et al., 2016 ⁴³	Case-control	Asia	1000	NA	PCR	PCR	Cytology	ASCUS, ASC-H LSIL, HSIL
Amorim et al., 2017 ⁵⁶	Case-control	South America	132	38.2	PCR	NA	Cytology/histology	LSIL, HSIL
Costa-Lira et al., 2017 ⁴⁴	Cross-sectional	South America	180	16–50 ^a	PCR	PCR	–	NA
Ghosh et al., 2017 ¹⁷	Case-control	Asia	483	30–60 ^a	Wet-mount	HCII	Histology	LSIL, HSIL, CA
Al-Awadhi et al., 2018 ¹⁵	Cross-sectional	Asia	8836	NA	Cytology	NA	Cytology	ASCUS, LSIL, HSIL
Lockhart et al., 2019 ⁴⁵	Prospective cohort	Africa	344	18–49 ^a	PCR	PCR	NA	NA
Ferre et al., 2019 ⁴⁶	Cross-sectional	Africa	320	25	PCR	PCR	NA	NA
Ly et al., 2019 ⁴⁷	Cross-sectional	Asia	826	38.5	Wet-mount	PCR	NA	NA
Cunha et al., 2020 ⁴⁸	Cross-sectional	South America	353	39.7	PCR	PCR	NA	NA
Wang et al., 2020 ⁴⁹	Cross-sectional	Asia	4449	43.6	Wet-mount	PCR	NA	NA
Yang et al., 2020 ⁵⁰	Cross-sectional	Asia	310 545	>30	Wet-mount	PCR	NA	NA
Zheng et al., 2020 ⁵¹	Case-control	Asia	532	42.2	Wet-mount	PCR	Histology	LSIL, HSIL, CA
Gupta et al., 2020 ⁵⁸	Case-control	Asia	168	21–65 ^a	Wet-mount	NA	Histology	CA
Taku et al., 2021 ⁵²	Cross-sectional	Africa	205	45 ^b	PCR	PCR	NA	NA
Jary et al., 2021 ⁵³	Cross-sectional	Africa	144	37	PCR	PCR	NA	NA
Belfort, 2021 ⁵⁴	Cross-sectional	South America	562	30–49 ^c	PCR	PCR	NA	NA

Abbreviations: ASC-H, atypical squamous cells for which one cannot rule out high-grade squamous intraepithelial lesions; ASCUS, atypical squamous cells of undetermined significance; CA, cervical cancer; HCII, Hybrid capture II; HSIL, high-grade squamous intraepithelial lesions; LSIL, low-grade squamous intraepithelial lesions; NA, not available; PCR, polymerase chain reaction.

^aMinimum-maximum age values.

^bMedian age value.

^cIn this age range, 48.40% of patients were included.

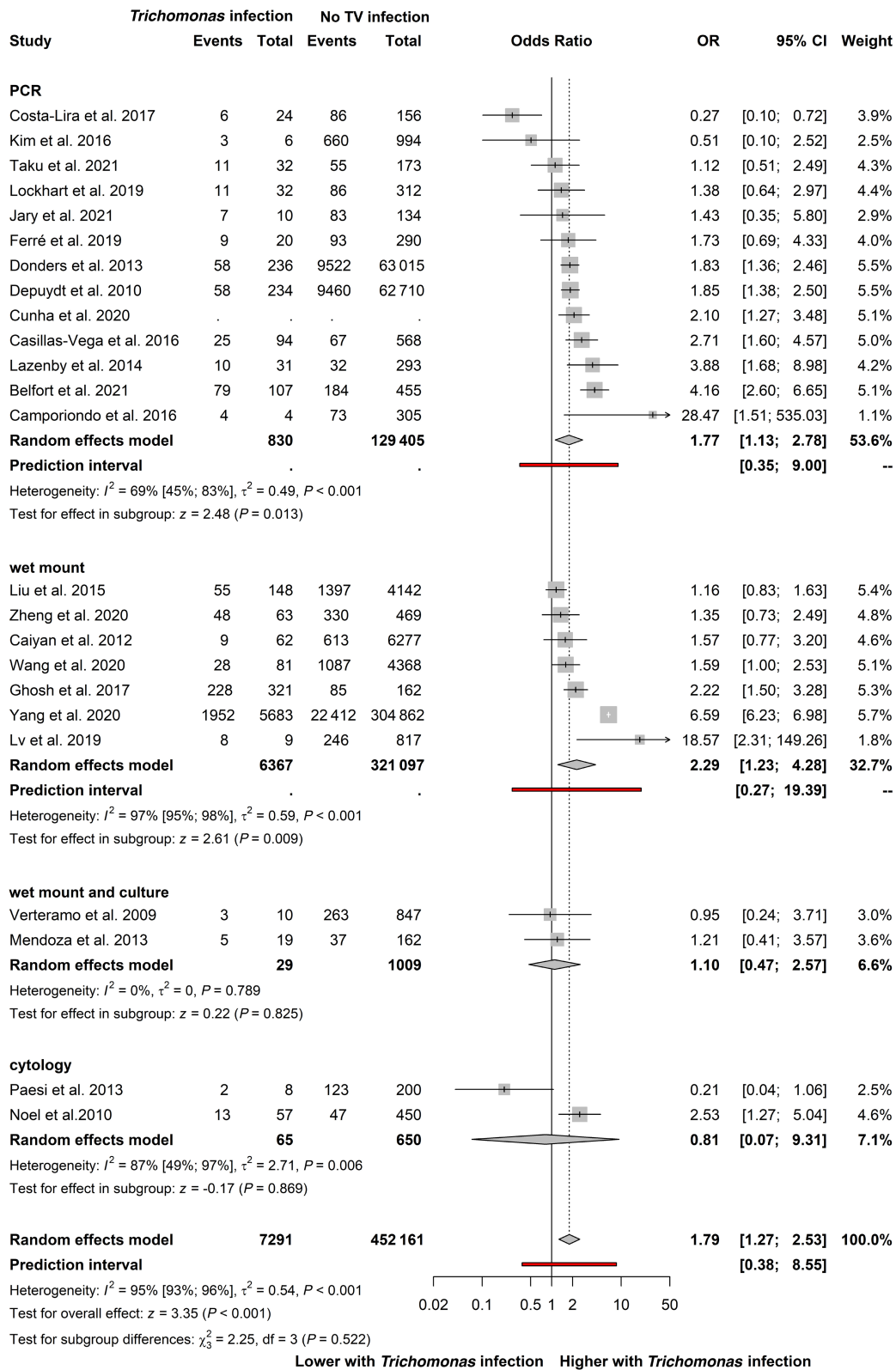


FIGURE 2 Forest plot of studies representing that *Trichomonas vaginalis* infection was associated with HPV co-infection. CI, confidence interval; OR, odds ratio; PCR, polymerase chain reaction; TV, *Trichomonas vaginalis*.

detection. A second study¹⁴ found even higher chances for HPV-co-infection (OR 4.10, 95% CI 1.70–9.80, see Table S3).

The association between *T. vaginalis* and cervical dysplasia

For the ASCUS outcome, five studies evaluated 1493 women in the exposure group and 75 135 women in the control group.^{15,16,43,54,55} Women who were *T. vaginalis*-positive had a 2.3 times higher chance for ASCUS diagnosis (95% CI 1.63–3.26; I^2 52%, see Figure S3) compared with women who did not have a *T. vaginalis* infection. A subgroup analysis based on the screening method showed that when *T. vaginalis* was detected with PCR, this association was even stronger (OR 2.91; 95% CI 1.95–4.35; I^2 0%). Articles from South America and Europe found almost threefold increased odds for the diagnosis of ASCUS (Belfort et al.⁵⁴: OR 2.99, 95% CI 1.06–8.43; Donders et al.¹⁶: OR 2.94, 95% CI 1.88–4.57, respectively; see Figure S4). When the leave-one-out analysis was carried out, the exclusion of Al-Awadhi et al.¹⁵ from ASCUS resulted in a higher association (OR 2.79, 95% CI 2.21–3.53; I^2 0%; see Figure S5).

Two studies investigated atypical glandular cells and *T. vaginalis* infection in sexually active women. Neither found an association between *T. vaginalis* positivity and atypical glandular cells (Donders et al.¹⁶: OR 1.33, 95% CI 0.08–21.40; Al-Awadhi et al.¹⁵: OR 1.55, 95% CI 0.46–5.41).

Altogether there were 10 eligible studies concerning LSIL, investigating 1740 women in the *T. vaginalis* group and 82 362 in the control group.^{14–17,37,43,51,54–56}

When examining the association between *T. vaginalis* and LSIL, we found that women who were infected with *T. vaginalis* had almost twofold odds of having LSIL (OR 1.92, 95% CI 0.78–4.77; I^2 91%; see Figure S6), compared with women who were not *T. vaginalis*-infected. However, the findings were statistically not significant. When *T. vaginalis* was detected with PCR, women had higher odds of having an LSIL diagnosis (OR 3.66, 95% CI 1.51–8.86; I^2 69%). Regarding the analysis of regional differences, we detected a ninefold chance for LSIL when *T. vaginalis* was present in women from South America (OR 9.36, 95% CI 2.34–37.36; I^2 63%; Figure S7). When the leave-one-out analysis was carried out, the exclusion of the study by Al-Awadhi et al.¹⁵ resulted in a higher association between *T. vaginalis* and LSIL detection (OR 2.79; 95% CI 1.61–4.82; I^2 65%). Furthermore, when the article by Amorim et al.⁵⁶ was excluded, we found an OR of 1.51 (95% CI 0.65–3.55; I^2 95%; see Figure S8).

Regarding a relationship between *T. vaginalis* and ASC-H, we could not find any association (OR 1.78, 95% CI 0.21–15.12; see Figure S9).^{16,43,54}

Eleven studies assessed 1796 women in the exposure group and 80 276 women in the control group for the association between *T. vaginalis* infection and HSIL.^{14–17,37,43,51,54–57} Patients diagnosed with *T. vaginalis* infection had 2.34 times higher odds of having an HSIL diagnosis (95% CI 1.10–4.95; I^2 75%; Figure 3) than women who were not diagnosed with *T. vaginalis*. According to the *T. vaginalis* detection

method, women diagnosed with PCR had higher odds of receiving an HSIL result (OR 3.81, 95% CI 1.23–11.78; I^2 81%). The subgroup analysis of the origins of the articles displayed sixfold odds in South America (OR 6.52, 95% CI 0.74–57.75; I^2 92%), although the findings were statistically not significant. One study from Europe found high odds for HSIL when *T. vaginalis* was present (Donders et al.¹⁶: OR 3.14, 95% CI 1.49–6.78; see Figure S10). When the leave-one-out analysis was carried out, the exclusion of the article by Al-Awadhi et al.¹⁵ from HSIL resulted in an OR of 2.87 (95% CI 1.43–5.75; I^2 67%). In addition, when the article by Amorim et al.⁵⁶ was excluded, the OR changed to 1.72 (95% CI 1.01–2.91; I^2 42%; see Figure S11).

One paper⁵⁴ performed a multivariate analysis (see Table S3).

The association between *T. vaginalis* and cervical cancer

Three articles were quantitatively analyzed, with 219 women in the *T. vaginalis*-positive group and 397 women in the control group.^{17,51,58} Women who were *T. vaginalis*-positive had 5.24 times higher odds of having cervical cancer (OR 5.23, 95% CI 3.03–9.04; I^2 3%; Figure 4) compared with *T. vaginalis*-negative women.

Association between *T. vaginalis*, cervical lesions, and cervical cancer in the HPV-positive population

We found four articles for the quantitative synthesis regarding the HPV-positive population when evaluating the association between *T. vaginalis* infection and cervical lesions.^{17,50,51,57}

For LSIL, three articles were analyzed, assessing 1932 women in the exposure group and 20 033 in the control group.^{17,50,51} *Trichomonas vaginalis*-positive women had 2.81 higher odds for LSIL diagnosis than women who were not *T. vaginalis* infected (95% CI 2.37–3.33; I^2 0%; see Figure 5).

In total, there were 1921 women in the exposure group and 20 750 women in the control group for HSIL.^{17,50,51,57} Patients who were diagnosed with *T. vaginalis* had more than twofold odds of having HSIL compared with women who were not diagnosed with *T. vaginalis* (OR 2.36, 95% CI 1.79–3.11; I^2 10%; see Figure 5).

Three studies examined cervical cancer, with 1811 women in the exposure group and 19 331 in the control group.^{17,50,51} We found that women who were *T. vaginalis*-positive had increased odds of being diagnosed with cervical cancer compared with women who were not *T. vaginalis*-positive (OR 3.09, 95% CI 1.66–5.77; I^2 45%; see Figure 5).

3.4 | Risk of bias assessment and quality of evidence

The results of the risk of bias assessment are presented for every outcome. For the *T. vaginalis*-HPV co-infection outcome, seven articles demonstrated “a moderate risk for study confounding bias”,

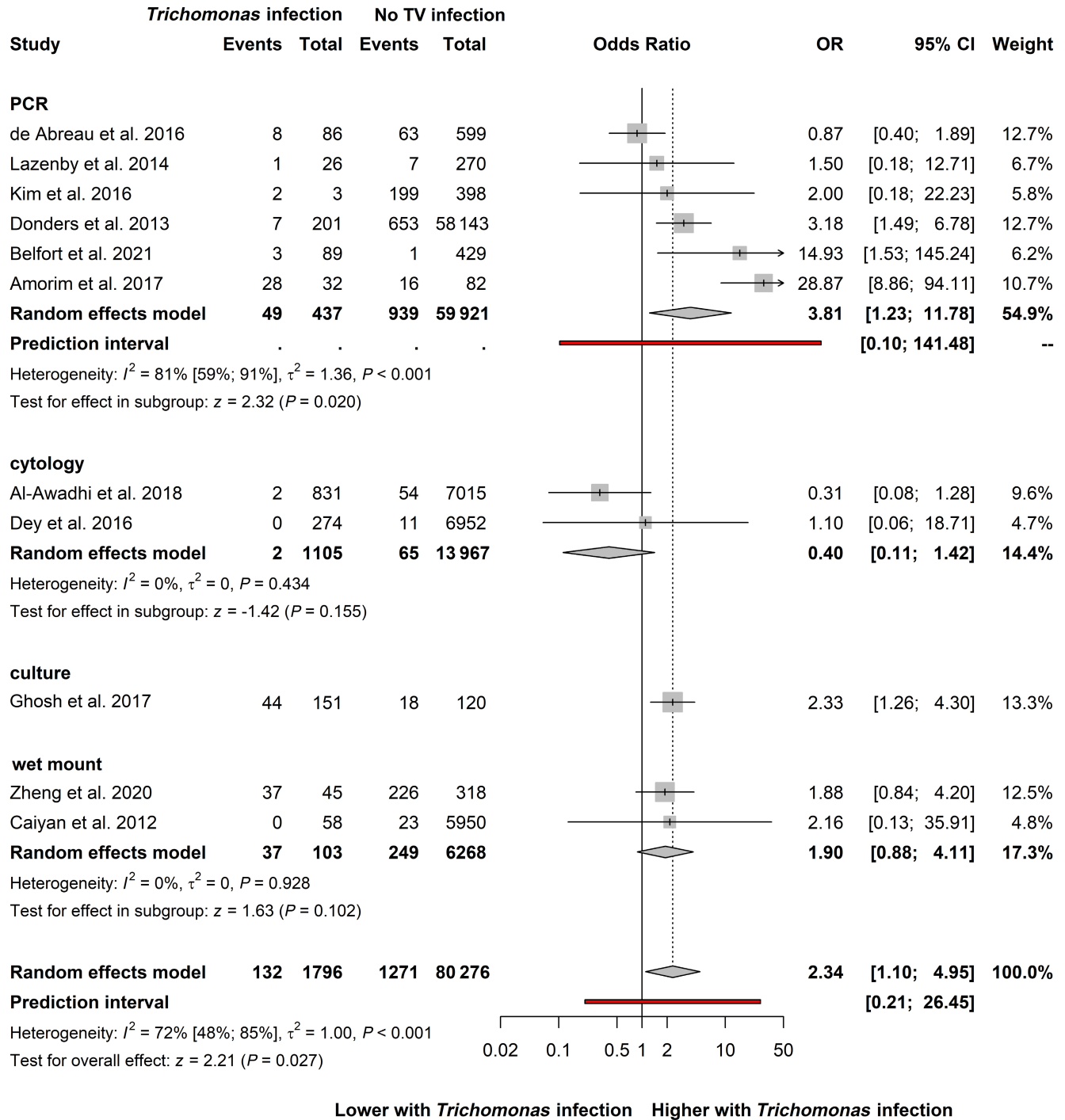


FIGURE 3 Forest plot of studies representing that *Trichomonas vaginalis* infection was associated with high-grade squamous intraepithelial lesions. CI, confidence interval; HSIL, high-grade squamous intraepithelial lesions; OR, odds ratio; PCR, polymerase chain reaction; TV, *Trichomonas vaginalis*.

and three articles demonstrated “a high risk of bias”. In the study participation domain we detected a high risk of bias in three articles. In the cervical dysplasia groups, the risk of bias was low. In the study confounding domain, we found one article to be at a high risk of bias in ASCUS and two articles of a high risk of bias in the LSIL and HSIL groups. In the cervical cancer, group, we found one article

in the confounding domain of the study at a high risk of bias. (see [Figures S12–S22](#)).

Our Summary of Findings included six outcomes for the first PEO and three for the second PEO (see [Tables S3 and S4](#)). The quality of evidence was “low” for six outcomes and “very low” for three outcomes.

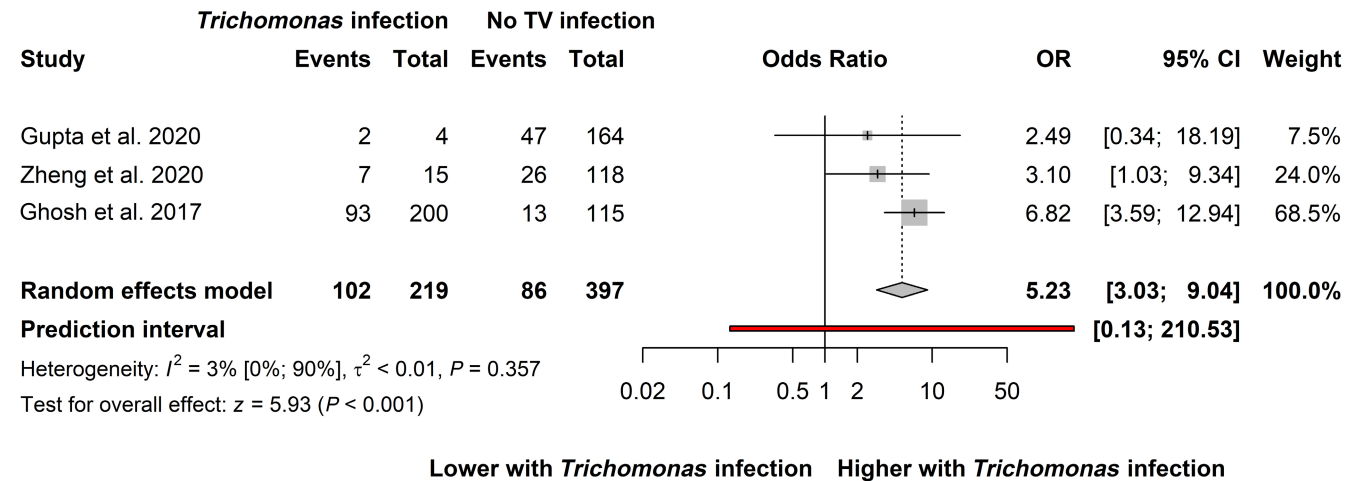


FIGURE 4 Forest plot of studies representing that *Trichomonas vaginalis* infection was associated with cervical cancer. CI, confidence interval; OR, odds ratio; TV, *Trichomonas vaginalis*.

3.5 | Publication bias and heterogeneity

We performed the Egger test and a funnel plot to assess publication bias in the *T. vaginalis*-HPV co-infection, *T. vaginalis*-LSIL, and *T. vaginalis*-HSIL groups. In all three cases, the funnel plots showed some asymmetry. Even on the basis of Egger test, we did not find a significant publication bias as the P values were greater than 0.1 (see Figures S23–S25).

4 | DISCUSSION

Our study, which included nearly half a million women from population-based studies, showed a positive association between *T. vaginalis* and cervical carcinogenesis. First, we investigated the association between *T. vaginalis* and HPV. We found that women with a *T. vaginalis* infection had higher odds of being diagnosed with a concomitant HPV infection than women who were *T. vaginalis*-negative. In the relation between *T. vaginalis* and cervical dysplasia, a significant association was found. When we evaluated the relationship between *T. vaginalis* and cervical cancer, we also found a statistically significant association resulting in higher odds of developing cervical cancer among women infected with *T. vaginalis*. Regarding our second clinical question in the HPV-positive population, we found a positive association between *T. vaginalis*, LSIL, HSIL, and cervical cancer.

As for strengths, our study is the first meta-analysis to investigate the relationship between *T. vaginalis*, HPV, and cervical lesions in detail. As a result of the large sample size in the assessed articles, we could include nearly half a million patients in the analysis. Moreover, most of the studies carried a low risk of bias.

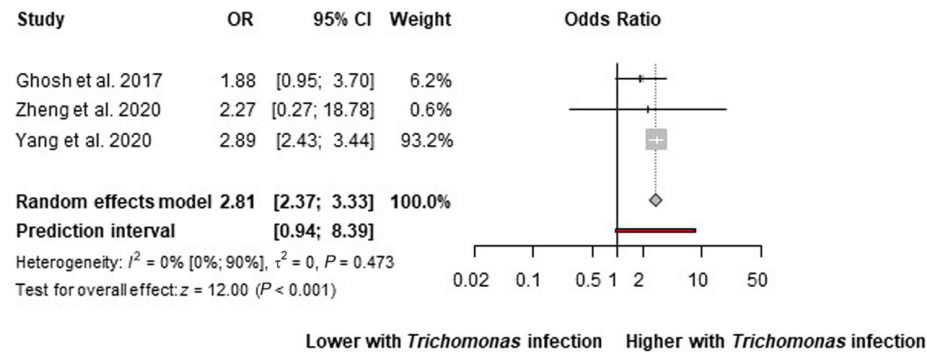
However, the results need to be interpreted together with the limitations. First, as none of the studies followed up the *T. vaginalis*-infected population, and all participants were screened for *T. vaginalis*, HPV, and cervical carcinogenesis simultaneously, we do not know

how *T. vaginalis* can contribute to the development of the outcomes. Second, as many studies did not perform multivariate analyses, we could not calculate pooled adjusted ORs. The inadequate control of confounders may lead to an underestimation or overestimation of the analyzed associations. Third, it is not clear whether *T. vaginalis* infection causes the cervical environment to be more susceptible to HPV infection and to the subsequent CIN, or whether cervical dysplasia makes the environment more attractive to *T. vaginalis* infection.⁵⁹ Fourth, in the diagnosis of cervical lesions, some studies used cytology, which is subjective, and it is a diagnostic method that is difficult to replicate.⁶⁰ Fifth, according to the GRADE assessment, the quality of evidence was low in six and very low in three outcomes. Sixth, not all HPVs are oncogenic though in the *T. vaginalis*-HPV association, 10 studies included non-oncogenic HPV strains in their investigation too.

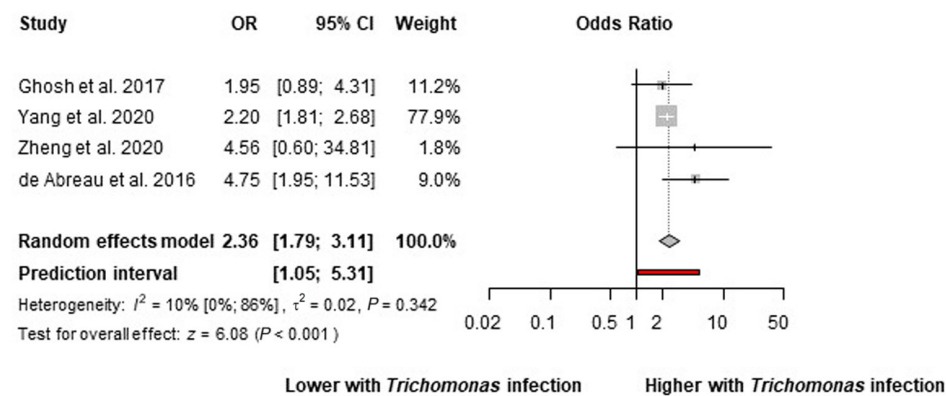
The association between *T. vaginalis* and HPV showed that STIs often coexist because of similar behavioral risk factors such as young age, a high number of sexual partners, and unprotected intercourse.^{16,61} Therefore, we cannot conclude that *T. vaginalis* infection affects HPV acquisition because both infections can be concomitantly present. The etiology of cervical cancer and most CIN are attributable to high-risk HPV types.⁵ Therefore, HPV could be a confounding factor for our cervix-related outcomes. However, if we only investigate the HPV-positive population, we could observe an even more increased association between *T. vaginalis* cervical dysplasia and cancer.

In contrast, not all HPV types carry the same oncogenic risk as HPV 16 and HPV 18, which cause around 70% of all cervical cancers worldwide.⁵ Therefore, HPV positivity in women does not represent a homogeneous population from an oncogenic point of view. Some prospective studies suggest that the likelihood of a persistent HPV infection increases in the presence of concomitant *T. vaginalis* infection.^{45,62} Behind this observation there is a presumption of how *T. vaginalis* can alter HPV clearance. *Trichomonas vaginalis* can cause micro-lesions in the cervical epithelium,

LSIL



HSIL



Cervical cancer

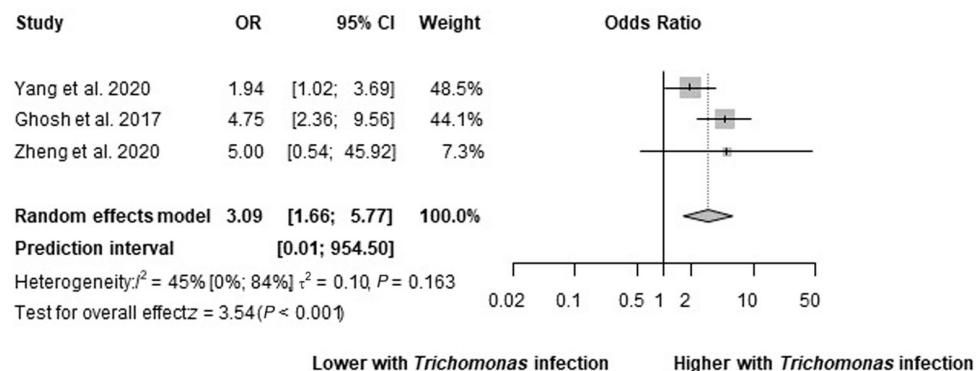


FIGURE 5 Forest plot of studies representing that *Trichomonas vaginalis* was associated with low-grade squamous intraepithelial lesions, high-grade squamous intraepithelial lesions, and cervical cancer in the HPV-positive population. CI, confidence interval; HSIL, high-grade squamous intraepithelial lesions; LSIL, low-grade squamous intraepithelial lesions.

decrease the protective mucus layer of the vagina, and induce proinflammatory cytokines through immune response, which can facilitate the spread of an HPV infection into the basal layer of the

cervical epithelium and induce persistent HPV infection.^{7,63,64} As persistent HPV infection occurs, the probability of cervical dysplasia increases, promoting cervical carcinogenesis.⁶⁵ Coexistence

of different genital infections, *Chlamydia trachomatis*, and bacterial vaginosis can also induce persistent HPV infection, resulting in cervical dysplasia progression.^{66,67}

When assessing ASCUS, we found a significant association with *T. vaginalis* infection. A Belgian study also found that women diagnosed with ASCUS had been HPV-negative but *T. vaginalis*-positive in a few cases, suggesting that *T. vaginalis* could also lead to ASCUS.¹⁶ In the cervical dysplasia group, we found the highest odds for cervical lesions when *T. vaginalis* was detected with PCR, probably because this method was the most sensitive for *T. vaginalis* detection.¹⁹ In the *T. vaginalis* and cervical dysplasia group, we found higher odds in the ASCUS and LSIL groups in South America, although we had only one article in the ASCUS group. The prevalence of *T. vaginalis* is deeply connected to socioeconomic variables, sexual behaviors, and access to health care. Without surveillance programs, the actual epidemiologic state of *T. vaginalis* is unknown. However, countries where the populations have higher incomes generally have a lower prevalence of *T. vaginalis*, and countries where the populations have lower incomes generally have a higher prevalence.¹³ In the sensitivity analysis, two articles could have altered our results. One of the outliers¹⁵ led to a lower association between *T. vaginalis* and cervical dysplasia. In this study, *T. vaginalis* was diagnosed with cytology, which is not a reference standard detection of *T. vaginalis*.¹⁹ The other article⁵⁶ came from an area of Brazil where poverty rate was high, and cervical cancer was the second most common cancer.³ These findings can explain the high ORs we experienced in the *T. vaginalis* and cervical dysplasia group.

Lipophosphoglycan (LPG), a virulence factor found on the surface of *T. vaginalis*, can induce immunologic reactions depending on the type of LPG. In reaction to these LPG particles, the host epithelial cells can secrete proinflammatory cytokines, interleukin-8 and macrophage inflammatory protein 3 α , which induce the inflammation of the cervix and the vagina. At the same time, other LPGs found on *T. vaginalis* can decrease the level of proinflammatory cytokines and evade immune reactions. This is in line with the clinical finding that *T. vaginalis* can often be asymptomatic or can cause persistent infection.⁶⁴

Inflammation of the cervix has been associated with an increased risk of CIN in one study.⁷ Another article found elevated levels of interleukin-6 and interleukin-8 in CIN and cervical cancer.⁶⁸ Generally, inflammation is considered a risk factor for developing many cancer types.⁶⁹ One study investigated the microbial component of the vagina in cervical cancer patients and non-cervical cancer patients, assuming that cervical cancer disrupts the vaginal microbiota and makes it attractive to infectious diseases. *Trichomonas vaginalis* is possibly less of a cofactor than a consequence of cervical cancer.⁷⁰ The intact state of the vaginal microbiome with *Lactobacillus* species is essential for protection against STIs. The abruption of this complex microbiome increases the probability of genital infections due to decreased defensive barriers.⁷¹ One study proved the proinflammatory synergism between vaginal dysbiosis and *T. vaginalis*; moreover, it suggested a surface biofilm that makes them more resistant

to antibiotic treatment.⁷² Overall, STIs and vaginal infections have been considered possible cofactors in the development of CIN and cervical cancer. In one meta-analysis, *Chlamydia trachomatis* was found to be associated with cervical cancer, whereas another meta-analysis also found an association between bacterial vaginosis and cervical lesions.^{61,67} Our findings also support the idea that STIs and vaginal infections might act as cofactors in the development of cervical cancer.

We believe that more studies are needed to control the confounding factors; therefore, the true effect of *T. vaginalis* on cervical carcinogenesis could be estimated in a more reliable way. Second, we recommend that clinicians who treat women in their practice always consider HPV infection and cervical lesions when diagnosing *T. vaginalis* infection. Even though we cannot conclude a causative relationship between *T. vaginalis* and cervical carcinogenesis, *T. vaginalis* is associated with HPV infection, cervical lesions, and cervical cancer, so a follow up of patients after the *T. vaginalis* diagnosis might be beneficial. Many countries have implemented HPV-based cervical cancer screening programs, which means a greater detection rate in HPV strains.⁷³ According to our study, *T. vaginalis* and HPV are associated; therefore, in the case of an HPV diagnosis, the screening and treatment of *T. vaginalis* are advisable because of its potential carcinogenic effect on the cervix.

In conclusion, our results showed that *T. vaginalis* infection might increase the odds of cervical lesions and cancer development in sexually active women. We advise clinicians to evaluate HPV and cervical dysplasia in the case of a *T. vaginalis* diagnosis.

AUTHOR CONTRIBUTIONS

Balázs Hamar contributed to conceptualization, project administration, and writing the original draft; Brigitta Teutsch and Péter Hegyi contributed to conceptualization, project administration, methodology, and writing—review & editing; Eszter Hoffmann and Zsombor Hunka contributed to data curation and writing—review & editing; Alex Váradi contributed to data curation, formal analysis, visualization, and writing—review & editing; Nándor Ács and Zsolt Melczer contributed to conceptualization, supervision, and writing—review & editing; and Péter Nyirády, Balázs Lintner, and Réka Juhász Hermáné contributed to conceptualization, and writing—review & editing. All authors certify that they have participated sufficiently in the work, including the concept, design, analysis, writing, and/or revision of the manuscript, to take public responsibility for the content of the work.

CONFLICT OF INTEREST STATEMENT

The authors have no conflicts of interest.

DATA AVAILABILITY STATEMENT

The data sets used in this study can be found in the full-text articles included in the systematic review and meta-analysis.

ORCID

Balázs Hamar  <https://orcid.org/0000-0003-1996-6035>

REFERENCES

1. Sung H, Ferlay J, Siegel RL, et al. Global cancer statistics 2020: GLOBOCAN estimates of incidence and mortality worldwide for 36 cancers in 185 countries. *CA Cancer J Clin*. 2021;71(3):209-249.
2. Simms KT, Steinberg J, Caruana M, et al. Impact of scaled up human papillomavirus vaccination and cervical screening and the potential for global elimination of cervical cancer in 181 countries, 2020-99: a modelling study. *Lancet Oncol*. 2019;20(3):394-407.
3. Giannini A, Bogani G, Vizza E, et al. Advances on prevention and screening of gynecologic tumors: are we stepping forward? *Healthcare*. 2022;10(9):1.
4. D'Oria O, Corrado G, Laganà AS, Chiantera V, Vizza E, Giannini A. New advances in cervical cancer: from bench to bedside. *Int J Environ Res Public Health*. 2022;19(12):2-3.
5. zur Hausen H. Papillomaviruses and cancer: from basic studies to clinical application. *Nat Rev Cancer*. 2002;2(5):342-350.
6. Mercer F, Johnson PJ. *Trichomonas vaginalis*: pathogenesis, symbiont interactions, and host cell immune responses. *Trends Parasitol*. 2018;34(8):683-693.
7. Castle PE, Hillier SL, Rabe LK, et al. An association of cervical inflammation with high-grade cervical neoplasia in women infected with oncogenic human papillomavirus (HPV). *Cancer Epidemiol Biomarkers Prev*. 2001;10(10):1021-1027.
8. Brusselaers N, Shrestha S, van de Wijgert J, Verstraelen H. Vaginal dysbiosis and the risk of human papillomavirus and cervical cancer: systematic review and meta-analysis. *Am J Obstet Gynecol*. 2019;221(1):9-18.e8.
9. Slattery ML, Robison LM, Schuman KL, et al. Cigarette smoking and exposure to passive smoke are risk factors for cervical cancer. *JAMA*. 1989;261(11):1593-1598.
10. Huh WK. Human papillomavirus infection: a concise review of natural history. *Obstet Gynecol*. 2009;114(1):139-143.
11. Hawes SE, Kiviat NB. Are genital infections and inflammation cofactors in the pathogenesis of invasive cervical cancer? *J Natl Cancer Inst*. 2002;94(21):1592-1593.
12. Van der Pol B. *Trichomonas vaginalis* infection: the most prevalent nonviral sexually transmitted infection receives the least public health attention. *Clin Infect Dis*. 2007;44(1):23-25.
13. Kissinger P. *Trichomonas vaginalis*: a review of epidemiologic, clinical and treatment issues. *BMC Infect Dis*. 2015;15:307.
14. Lazenby GB, Taylor PT, Badman BS, et al. An association between *Trichomonas vaginalis* and high-risk human papillomavirus in rural tanzanian women undergoing cervical cancer screening. *Clin Ther*. 2014;36(1):38-45.
15. Al-Awadhi R, Al-Shaheen A, Al-Juwaiser A, George SS, Sharma P, Kapila K. Prevalence of infectious organisms observed in cervical smears between 1997-2014 at Mubarak Al-Kabeer Hospital, Kuwait. *Sultan Qaboos Univ Med J*. 2018;18(3):e324-e328.
16. Donders GG, Depuydt CE, Bogers JP, Vereecken AJ. Association of *Trichomonas vaginalis* and cytological abnormalities of the cervix in low risk women. *PLoS One*. 2013;8(12):e86266.
17. Ghosh I, Muwonge R, Mittal S, et al. Association between high risk human papillomavirus infection and co-infection with *Candida* spp. and *Trichomonas vaginalis* in women with cervical premalignant and malignant lesions. *J Clin Virol*. 2017;87:43-48.
18. Zhang ZF, Begg CB. Is *Trichomonas vaginalis* a cause of cervical neoplasia? Results from a combined analysis of 24 studies. *Int J Epidemiol*. 1994;23(4):682-690.
19. Asmah RH, Agyeman RO, Obeng-Nkrumah N, et al. *Trichomonas vaginalis* infection and the diagnostic significance of detection tests among Ghanaian outpatients. *BMC Womens Health*. 2018;18(1):206.
20. Aslan DL, Gulbahce HE, Stelow EB, et al. The diagnosis of *Trichomonas vaginalis* in liquid-based pap tests: correlation with PCR. *Diagn Cytopathol*. 2005;32(6):341-344.
21. Yang S, Zhao W, Wang H, Wang Y, Li J, Wu X. *Trichomonas vaginalis* infection-associated risk of cervical cancer: a meta-analysis. *Eur J Obstet Gynecol Reprod Biol*. 2018;228:166-173.
22. Page MJ, McKenzie JE, Bossuyt PM, et al. The PRISMA 2020 statement: an updated guideline for reporting systematic reviews. *BMJ*. 2021;372:n71.
23. Cumpston M, Li T, Page MJ, et al. Updated guidance for trusted systematic reviews: a new edition of the Cochrane handbook for systematic reviews of interventions. *Cochrane Database Syst Rev*. 2019;10:ed000142.
24. van Zuuren EJ, Fedorowicz Z. Moose on the loose: checklist for meta-analyses of observational studies. *Br J Dermatol*. 2016;175(5):853-854.
25. Munn Z, Stern C, Aromataris E, Lockwood C, Jordan Z. What kind of systematic review should I conduct? A proposed typology and guidance for systematic reviewers in the medical and health sciences. *BMC Med Res Methodol*. 2018;18(1):5.
26. Roteli-Martins CM, Alves VA, Santos RT, Martinez EZ, Syrjänen KJ, Derchain SF. Value of morphological criteria in diagnosing cervical HPV lesions confirmed by in situ hybridization and hybrid capture assay. *Pathol Res Pract*. 2001;197(10):677-682.
27. McHugh ML. Interrater reliability: the kappa statistic. *Biochem Med*. 2012;22(3):276-282.
28. Hayden JA, van der Windt DA, Cartwright JL, Côté P, Bombardier C. Assessing bias in studies of prognostic factors. *Ann Intern Med*. 2013;158(4):280-286.
29. GradePro. <https://www.gradepro.org>
30. Mantel N, Haenszel W. Statistical aspects of the analysis of data from retrospective studies of disease. *J Natl Cancer Inst*. 1959;22(4):719-748.
31. Thompson SG, Turner RM, Warn DE. Multilevel models for meta-analysis, and their application to absolute risk differences. *Stat Methods Med Res*. 2001;10(6):375-392.
32. Guido Schwarzer JRC. *Gerta Rücker. Meta-Analysis with R*. Springer International Publishing Switzerland; 2015.
33. IntHout J, Ioannidis JP, Rovers MM, Goeman JJ. Plea for routinely presenting prediction intervals in meta-analysis. *BMJ Open*. 2016;6(7):e010247.
34. Verteramo R, Pierangeli A, Mancini E, et al. Human papillomaviruses and genital co-infections in gynaecological outpatients. *BMC Infect Dis*. 2009;9:2-5.
35. Noël JC, Fayt I, Romero Munoz MR, Simon P, Engohan-Aloghe C. High prevalence of high-risk human papillomavirus infection among women with *Trichomonas vaginalis* infection on monolayer cytology. *Arch Gynecol Obstet*. 2010;282(5):503-505.
36. Depuydt CE, Leuridan E, Van Damme P, Bogers J, Vereecken AJ, Donders GG. Epidemiology of *Trichomonas vaginalis* and human papillomavirus infection detected by real-time PCR in flanders. *Gynecol Obstet Invest*. 2010;70(4):273-280.
37. Caiyan X, Weiyuan Z, Minghui W, Songwen Z. Prevalence and risk factors of lower genital tract infections among women in Beijing, China. *J Obstet Gynaecol Res*. 2012;38(1):310-315.
38. Mendoza L, Mongelos P, Paez M, et al. Human papillomavirus and other genital infections in indigenous women from Paraguay: a cross-sectional analytical study. *BMC Infect Dis*. 2013;13(1):2-6.
39. Paesi SO, Aver L, Barea F, Vanni A, Roesch-Ely M. Human papillomavirus and infections of the lower genital tract in women with abnormal cervical cytological examination. *Sci Med*. 2013;23(1):41-46.
40. Liu J, Liu W, Liu Y, Zhou X, Zhang Z, Sun Z. Prevalence of microorganisms co-infections in human papillomaviruses infected women in northern China. *Arch Gynecol Obstet*. 2016;293(3):595-602.
41. Casillas-Vega N, Morfín-Otero R, García S, et al. Sexually transmitted pathogens, coinfections and risk factors in patients attending obstetrics and gynecology clinics in Jalisco, Mexico. *Salud Publica Mex*. 2016;58(4):437-445.

42. Camporiondo MP, Farchi F, Ciccozzi M, et al. Detection of HPV and co-infecting pathogens in healthy Italian women by multiplex real-time PCR. *Inf Med*. 2016;24(1):12-17.
43. Kim HS, Kim TJ, Lee IH, Hong SR. Associations between sexually transmitted infections, high-risk human papillomavirus infection, and abnormal cervical pap smear results in OB/GYN outpatients. *Journal of Gynecol Oncol*. 2016;27(5):2-6.
44. Costa-Lira E, Jacinto AHVL, Silva LM, et al. Prevalence of human papillomavirus, chlamydia trachomatis, and *Trichomonas vaginalis* infections in Amazonian women with normal and abnormal cytology. *Genet Mol Res*. 2017;16(2):2-5.
45. Lockhart A, Senkomago V, Ting J, et al. Prevalence and risk factors of *Trichomonas vaginalis* among female sexual workers in Nairobi, Kenya. *Sex Transm Dis*. 2019;46(7):458-464.
46. Ferré VM, Ekouevi DK, Gbeasor-Komlanvi FA, et al. Prevalence of human papillomavirus, human immunodeficiency virus and other sexually transmitted infections among female sex workers in Togo: a national cross-sectional survey. *Clin Microbiol Infect*. 2019;25(12):1560.e1-1560.e7.
47. Lv P, Zhao F, Xu X, Xu J, Wang Q, Zhao Z. Correlation between common lower genital tract microbes and high-risk human papillomavirus infection. *Can J Infect Dis Med Microbiol*. 2019;2019:1-6.
48. Cunha APA, Belfort IKP, Mendes FPB, et al. Human papillomavirus and its association with other sexually transmitted coinfection among sexually active women from the northeast of Brazil. *Interdiscip Perspect Infect Dis*. 2020;2020:1-8.
49. Wang W, Zhang XH, Li M, Hao CH, Liang HP. Association between vaginal infections and the types and viral loads of human papillomavirus: a clinical study based on 4,449 cases of gynecologic outpatients. *Can J Infect Dis Med Microbiol*. 2020;2020:1-6.
50. Yang M, Li L, Jiang C, et al. Co-infection with *Trichomonas vaginalis* increases the risk of cervical intraepithelial neoplasia grade 2-3 among HPV16 positive female: a large population-based study. *BMC Infect Dis*. 2020;20(1):642.
51. Zheng JJ, Miao JR, Wu Q, Yu CX, Mu L, Song JH. Correlation between HPV-negative cervical lesions and cervical microenvironment. *Taiwan J Obstet Gynecol*. 2020;59(6):855-861.
52. Taku O, Brink A, Meiring TL, et al. Detection of sexually transmitted pathogens and co-infection with human papillomavirus in women residing in rural eastern cape, South Africa. *PeerJ*. 2021;9:9.
53. Jary A, Teguede I, Sidibé Y, et al. Prevalence of cervical HPV infection, sexually transmitted infections and associated antimicrobial resistance in women attending cervical cancer screening in Mali. *Int J Infect Dis*. 2021;108:610-616.
54. Belfort IKP, Cunha APA, Mendes FPB, et al. *Trichomonas vaginalis* as a risk factor for human papillomavirus: a study with women undergoing cervical cancer screening in a northeast region of Brazil. *BMC Womens Health*. 2021;21(1):2-4.
55. Dey S, Pahwa P, Mishra A, Govil J, Dhillion PK. Reproductive tract infections and premalignant lesions of cervix: evidence from women presenting at the cancer detection Centre of the Indian Cancer Society, Delhi, 2000-2012. *J Obstet Gynaecol India*. 2016;66:441-451.
56. Amorim AT, Marques LM, Campos GB, et al. Co-infection of sexually transmitted pathogens and human papillomavirus in cervical samples of women of Brazil. *BMC Infect Dis*. 2017;17(1):769.
57. de Abreu ALP, Malaguti N, Souza RP, et al. Association of human papillomavirus, Neisseria gonorrhoeae and chlamydia trachomatis co-infections on the risk of high-grade squamous intraepithelial cervical lesion. *Am J Cancer Res*. 2016;6(6):1371-1383.
58. Gupta R, Singh N, Kalyan RK, Agrawal S. Case-control study to find Association of Common RTIs with CIN and cervical cancer. *Indian J Gynecol Oncol*. 2020;18(4):2-6.
59. Lin S, Zhang B, Lin Y, Lin Y, Zuo X. Dysbiosis of cervical and vaginal microbiota associated with cervical intraepithelial neoplasia. *Front Cell Infect Microbiol*. 2022;12:767693.
60. Kitchener HC, Castle PE, Cox JT. Chapter 7: achievements and limitations of cervical cytology screening. *Vaccine*. 2006;24(Suppl 3):S3/63-S3/70.
61. Zhu H, Shen Z, Luo H, Zhang W, Zhu X. Chlamydia trachomatis infection-associated risk of cervical cancer: a meta-analysis. *Medicine*. 2016;95(13):e3077.
62. Shew ML, Fortenberry JD, Tu W, et al. Association of condom use, sexual behaviors, and sexually transmitted infections with the duration of genital human papillomavirus infection among adolescent women. *Arch Pediatr Adolesc Med*. 2006;160(2):151-156.
63. Madeiro Da Costa RF, De Souza W, Benchimol M, Alderete JF, Morgado-Díaz JA. *Trichomonas vaginalis* perturbs the junctional complex in epithelial cells. *Cell Res*. 2005;15(9):704-716.
64. Fichorova RN, Yamamoto HS, Fashemi T, et al. *Trichomonas vaginalis* Lipophosphoglycan exploits binding to Galectin-1 and -3 to modulate epithelial immunity. *J Biol Chem*. 2016;291(2):998-1013.
65. Bogani G, Taverna F, Lombardo C, et al. Retrospective study of the influence of HPV persistence on outcomes among women with high-risk HPV infections and negative cytology. *Int J Gynaecol Obstet*. 2017;138(1):62-68.
66. Naldini G, Grisci C, Chiavarini M, Fabiani R. Association between human papillomavirus and chlamydia trachomatis infection risk in women: a systematic review and meta-analysis. *Int J Public Health*. 2019;64(6):943-955.
67. Gillet E, Meys JF, Verstraelen H, et al. Association between bacterial vaginosis and cervical intraepithelial neoplasia: systematic review and meta-analysis. *PLoS One*. 2012;7(10):e45201.
68. Tjong MY, van der Vange N, ten Kate FJ, et al. Increased IL-6 and IL-8 levels in cervicovaginal secretions of patients with cervical cancer. *Gynecol Oncol*. 1999;73(2):285-291.
69. Ames BN, Gold LS, Willett WC. The causes and prevention of cancer. *Proc Natl Acad Sci U S A*. 1995;92(12):5258-5265.
70. Kovachev SM. Cervical cancer and vaginal microbiota changes. *Arch Microbiol*. 2020;202(2):323-327.
71. Alimena S, Davis J, Fichorova RN, Feldman S. The vaginal microbiome: a complex milieu affecting risk of human papillomavirus persistence and cervical cancer. *Curr Probl Cancer*. 2022;46(4):100877.
72. Fichorova RN, Buck OR, Yamamoto HS, et al. The villain team-up or how *Trichomonas vaginalis* and bacterial vaginosis alter innate immunity in concert. *Sex Transm Infect*. 2013;89(6):460-466.
73. Smith MA, Sherrah M, Sultana F, et al. National experience in the first two years of primary human papillomavirus (HPV) cervical screening in an HPV vaccinated population in Australia: observational study. *BMJ*. 2022;376:e068582.

SUPPORTING INFORMATION

Additional supporting information can be found online in the Supporting Information section at the end of this article.

How to cite this article: Hamar B, Teutsch B, Hoffmann E, et al. *Trichomonas vaginalis* infection is associated with increased risk of cervical carcinogenesis: A systematic review and meta-analysis of 470000 patients. *Int J Gynecol Obstet*. 2023;163:31-43. doi:[10.1002/ijgo.14763](https://doi.org/10.1002/ijgo.14763)