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Short title: CB1-mediated inhibition of keratin expression

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Abbreviations: ACEA, arachidonoyl-chloro-ethanolamide; AEA, anandamide; CB1, cannabinoid receptor 1; ECS, endocannabinoid system; HF, hair follicle; KC, keratinocyte

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Introduction

Endocannabinoids as well as exocannabinoids (such as the active components of cannabis) are reported to control the functions of various types of cells *via* CB-dependent or independent manner (Kupczyk, Reich & Szepietowski 2009). The endocannabinoid system (ECS) consists of these CBs, their endogenous ligands (*i.e.* endocannabinoids, such as anandamide [AEA] and 2-arachidonoylglycerol), and enzymes responsible for endocannabinoid synthesis and degradation (Biro *et al.* 2009). ~~Since~~ In human skin, many different types of cells are now known to express functional cannabinoid receptors (CBs) (Biro *et al.* 2009; Czifra *et al.* 2012; Pucci *et al.* 2012; Roelandt *et al.* 2012; Stander *et al.* 2005; Sugawara *et al.* 2012; Telek *et al.* 2007; Toth *et al.* 2011). The ECS is increasingly appreciated as an important regulator of skin function in health and disease. ~~The ever-increasing portfolio of functions~~ Thus, the ECS has become implicated in ~~ranges from~~ pain (Khasabova *et al.* 2012; Walker & Hohmann 2005) and itch control (Stander, Reinhardt & Luger 2006), ~~via~~ and the modulation of inflammation (Klein 2005) and allergy (Karsak *et al.* 2007). ~~Additionally, it is important in~~ of skin mast cell activation and intracutaneous mast cell maturation from resident progenitors (Sugawara *et al.* 2012). ~~to the~~ Furthermore, it regulates ~~of~~ fibrosis (Akhmetshina *et al.* 2009), sebocyte differentiation (Dobrosi *et al.* 2008) and eccrine epithelial biology (Czifra *et al.* 2012). Nevertheless, the functions of CB-mediated signaling in human keratinocytes (KCs) *in situ* are as yet poorly understood.

We have previously shown that outer root sheath KCs of human hair follicles (HFs) express CB1. ~~and that~~ CB1 stimulation by the endocannabinoid, AEA, markedly inhibited human HF growth by inhibiting hair matrix KC proliferation and inducing apoptosis, thus leading to premature HF involution (catagen development). This was reversed by the CB1-specific antagonist, AM251 (Telek *et al.* 2007). Similarly, human epidermal KC express CBs, and their differentiation is regulated *via* CB1 (Maccarrone *et al.* 2003; Paradisi *et al.* 2008). AEA

also markedly suppresses human epidermal KC proliferation and induces apoptosis *via* CB1 *in vitro* and *in situ* (Toth *et al.* 2011). This suggests that the ECS could become a useful therapeutic target in the management of chronic hyperproliferative human skin diseases, such as psoriasis (Toth *et al.* 2011).

However, it remains unclear whether and how CB1-mediated signaling impacts on human KC differentiation, namely on the expression of hyperproliferation-associated keratins. Psoriasis is characterized by the upregulation of keratins K6 and K16 expression within lesional epidermis (Korver *et al.* 2006; Mommers *et al.* 2000). This pair of keratins is also prominently up-regulated in the epidermis under wound healing conditions in man and mice (Moll, Divo & Langbein 2008; Paladini *et al.* 1996; Rotty & Coulombe 2012) and is constitutively expressed in the outer root sheath KCs of human HFs (Langbein & Schweizer 2005; Moll, Divo & Langbein 2008). Since psoriasis is a chronic inflammatory, hyperproliferative dermatosis that, in addition to its anti-proliferative properties (Telek *et al.* 2007; Van Dross *et al.* 2012), might also profit from the well-recognized anti-inflammatory properties of CB1-mediated signaling (Sugawara *et al.* 2012; Wilkinson & Williamson 2007), we have investigated whether CB1 stimulation modulates K6 and K16 expression in human skin. This question was made particularly interesting in view of the most recent discovery that, in murine skin, K6 is not only a wound healing-associated marker keratin, but actively down-regulates KC migration during wound repair (Rotty & Coulombe 2012).

In order to answer this question, we used the CB1-specific agonist, arachidonoyl-chloro-ethanolamide (ACEA) (Harvey *et al.* 2012), and checked its effect on K6 expression *in situ*. This was done by utilizing full thickness human skin organ culture (Lu *et al.* 2007) as a physiologically and clinically relevant model to study multiple aspects of human skin biology under clinically relevant conditions *in vitro* (Bodo *et al.* 2010; Knuever *et al.* 2012; Langan *et al.* 2010; Lu *et al.* 2007; Sugawara *et al.* 2012). In order to confirm the CB1-specificity of the

observed effects of ACEA, we also used the CB1-specific antagonist, AM251 (Chanda *et al.* 2011).

~~Since~~ K16 serves as the type I keratin partner of K6 in the formation of intermediate filament heterodimers (Moll, Divo & Langbein 2008). ~~It is~~ also involved in epidermal barrier function (Grzanka *et al.* 2012; Thakoersing *et al.* 2012), and is upregulated in hyperproliferative conditions of the skin such as psoriasis (Iizuka *et al.* 2004) and atopic dermatitis (Grzanka *et al.* 2012). Therefore, we also examined the effects of ACEA on K16 expression.

HaCaT cells are a highly proliferating human KC line known to overexpress K6 (Ryle *et al.* 1989). Since HaCaT KCs share some other characteristics with psoriatic KCs and are often employed as surrogate “psoriatic” KCs (Balato *et al.* 2012; Farkas *et al.* 2001; George *et al.* 2010; Kim *et al.* 2011; Ronpirin & Tencomnao 2012; Saelee, Thongrakard & Tencomnao 2011), we also tested whether and how ACEA modulated K6 expression in these cells *in vitro*. In order to delineate whether any such effects on keratin expression resulted only indirectly from a possible down-regulation of KC proliferation by CB1 stimulation (Toth *et al.* 2011), double-labeling and quantitative immunohistomorphometry for both K6 and Ki-67 was performed. Finally, to investigate whether K6-expressing human KCs co-express CB1, double-immunolabeling for both antigens was employed.

Materials and Methods

Human skin organ culture

Isolated human skin samples obtained from elective plastic surgery procedures (32 pieces of skin fragments obtained by 4 mm punch biopsies from 4 individuals; 3 females and a male aged 26-74, average: 56.5; 3 skin samples were taken from the scalp and one was taken from the hip) were maintained in supplemented serum-free William’s E medium as previously

reported (Bodo *et al.* 2010; Knuever *et al.* 2012; Lu *et al.* 2007; Poeggeler *et al.* 2010). Human tissue collection and handling was performed according to Helsinki guidelines, after institutional review board ethics approval (University of Luebeck) and informed patient consent.

Skin samples were first incubated overnight to adapt to culture conditions after which the medium was replaced and vehicle or test substances were added. For human skin organ culture, skin samples were treated with ACEA (Sigma-Aldrich, Taufkirchen, Germany, 30 μ M) or AM251 (Sigma-Aldrich, 1 μ M), or the combination of them for 1-day after the overnight incubation (Sugawara *et al.* 2012). Following culturing for the time indicated, skin samples were cryoembedded and prepared for histology, immunohistochemistry/immunofluorescence and quantitative immunohistomorphometry (Ramot *et al.* 2010; Ramot *et al.* 2011; Sugawara *et al.* 2012). Each evaluation was performed on 2-4 sections of 2 skin fragments per each treatment group from 2-4 individuals.

Cell culture

Human immortalized HaCaT KCs (Boukamp *et al.* 1988) were cultured in DMEM (Sigma-Aldrich) supplemented with 10% fetal bovine serum (Invitrogen, Paisley, UK) and antibiotics (PAA Laboratories, Pasching, Austria). For qRT-PCR, the cells were cultured with ACEA (1 μ M) for 8h.

qRT-PCR

qRT-PCR was performed on an ABI Prism 7000 sequence detection system (Applied Biosystems/Life Technologies, Foster City, CA, USA) using the 5' nuclease assay as detailed

in our previous reports (Toth *et al.* 2011; Toth *et al.* 2009). Total RNA was isolated from HaCaT keratinocytes using TRIreagent (Applied Biosystems/Life Technologies, Foster City, CA, USA) and digested with recombinant RNase-free DNase-1 (Applied Biosystems) according to the manufacturer's protocol. After isolation, one μ g of total RNA was reverse-transcribed into cDNA by using High Capacity cDNA kit (Applied Biosystems) following the manufacturer's protocol. PCR amplification was performed by using specific TaqMan primer and probes (Applied Biosystems, assay ID: Hs01699178_g1 for human K6A). As internal housekeeping gene control, transcripts of cyclophilin A (PPIA) were determined (Assay ID: Hs99999904 for human PPIA). The amount of the K6A transcripts was normalized to the control gene using the Δ CT method.

Immunohistochemistry

For the detection of K6 in organ cultured human skin as well as cultured HaCaT KCs, indirect immunofluorescence staining was performed using mouse anti-human K6 antibody (Progen, [Ks6.KA12](#)) at 1:10 dilution as a primary antibody and rhodamine conjugated goat anti-mouse IgG (Jackson ImmunoResearch Laboratories, West Grove, PA) at 1:200 dilution in phosphate-buffered saline (PBS) as a secondary antibody.

To study the proliferation of epidermal KCs, double-immunostaining for K6 and Ki-67 was performed. Briefly, after the staining for K6 with FITC conjugated goat anti-mouse IgG (Jackson ImmunoResearch Laboratories) as a secondary antibody, sections were incubated overnight at 4°C with a mouse anti-human Ki-67 antibody (DAKO, Hamburg, Germany) at 1:20 in PBS. Sections were then washed with PBS, followed by incubation with rhodamine conjugated goat anti-mouse IgG (Jackson ImmunoResearch Laboratories) (1:200 in PBS, 45 min) at room temperature.

To investigate the localization of CB1 and K6, double immunostaining was performed. For CB1 immunostaining, the highly sensitive tyramide signal amplification (TSA) technique (Perkin Elmer, Boston, MA) was applied. Cryosections were incubated overnight at 4°C with rabbit anti-CB1 (Santa Cruz, CA, USA) at 1:400 diluted in TNB (Tris, NaOH, Blocking reagent, TSA kit; Perkin-Elmer). Thereafter, the samples were labeled with goat biotinylated antibody against rabbit IgG (Jackson ImmunoResearch Laboratories) at 1:200 in TNB for 45 min at room temperature. Sections were then stained with streptavidin-conjugated horseradish peroxidase (1:100, 30 min, TSA kit) and were finally incubated with rhodamine conjugated tyramide (1:50, TSA kit). The TSA method was applied according to the manufacturer's protocol. For the second primary labeling, mouse anti-human K6 antibody (Progen) was applied at 1:20 in PBS, overnight at 4°C. After the wash with PBS, the sections were incubated with FITC conjugated goat anti-mouse IgG (Jackson ImmunoResearch Laboratories) (1:200 in PBS, 45 min) at room temperature.

For K16 antigen detection, we used the LSAB (DCS, Germany) detection method (Ramot *et al.* 2009) using the K16-gp as primary antibody (guinea-pig, PROGEN, Heidelberg, Germany, dilution 1:1000, [GP-CK16](#)), and biotinylated goat anti-guinea pig as secondary antibody (Vector Laboratories, Burlingame, CA, USA, dilution 1:200). [HistoGreen \(Linaris, Wertheim-Bettingen, Germany\) was used as peroxidase substrate.](#)

For all immunostainings, the respective primary antibodies were omitted as negative controls, and morphological criteria and reproduction of the previously published intracutaneous expression patterns of the examined antigens were used as internal positive and negative controls (Moll, Divo & Langbein 2008). [For all experiments, control and treated sections were stained \(and later evaluated\) on the same day by the same investigator. To avoid staining biases, we calculated the relative staining intensity \(arbitrary intensity; 1 as control group\) among treatment groups per each individual and then pooled data from all of the experiments.](#)

High magnification images of K6/Ki67 double immunofluorescence and K6 immunofluorescence on HaCaT cells were taken by laser scanning confocal microscopy (Fluoview 300, Olympus Tokyo, Japan) running Fluoview 2.1 software (Olympus).

The staining intensity of K6 and K16 in defined reference areas was assessed by quantitative immunohistomorphometry using the ImageJ software (NIH) (Bodo *et al.* 2010; Ramot *et al.* 2010; Ramot *et al.* 2011). For epidermal evaluation, staining intensity was evaluated in the suprabasal cells, and for HaCaT cells, staining intensity was measured in the colonies formed. For K6 immunofluorescence intensity, skin sections from 4 different individuals were used, while K16 immunofluorescence intensity and %Ki67 were evaluated in skin sections from 2 patients. 4-6 sections per one individual (2 sections per investigated skin fragment) were used for each evaluation. Each section was evaluated in two/three different non-adjacent microscopic fields (x 200), and the mean intensity was measured, and considered as a value. Each treatment group was compared to the control group (average value), and relative change in expression was calculated. Highly comparable results were obtained from different sections from different individuals.

Statistical Analysis

Significance of difference between two groups was evaluated using Student's t-test for unpaired samples. For multiple comparisons, one-way analysis of variance (ANOVA) was used, followed by Bonferroni's multiple comparison test, using Prism 5.0 software (GraphPad Prism Program, GraphPad, San Diego, CA). *p* values <0.05 were regarded as significant. All data in the figures are expressed as mean + SEM. **p*<0.05, ***p*<0.01, ****p*<0.001 for the indicated comparisons.

Results and Discussion

The CB1-selective agonist, ACEA, down-regulates K6 protein expression *in situ*

First, we asked whether the CB1-specific synthetic agonist ACEA (Pertwee *et al.* 2010; Sugawara *et al.* 2012) can modulate the expression of keratin K6 in human skin. K6 staining intensity within the epidermis of full-thickness human skin that had been organ-cultured for 24 hrs under serum-free conditions in the presence of ACEA (30 μ M) or vehicle alone was assessed by quantitative immunohistomorphometry. This showed that K6 immunoreactivity (IR) was significantly reduced after ACEA treatment, compared to the vehicle control group (Fig. 1A and B).

This down-regulation was abrogated in part by the co-administration with the CB1-specific antagonist, AM251 (Pertwee *et al.* 2010; Sugawara *et al.* 2012) (Fig. 1A and B). Therefore, intraepidermal K6 protein expression in normal human skin *in situ* is down-regulated in a CB1-specific manner.

ACEA also down-regulates K16 protein expression *in situ*

Since K16 is the type I keratin partner of K6 in KCs and is thought to stabilize this keratin protein as a cytoskeletal heteropolymeric intermediate filament (Ramot *et al.* 2009), we next analyzed K16 IR in the epidermis of organ cultured human skin samples treated with ACEA. In line with the K6 protein expression data, K16 IR was also significantly down-regulated by ACEA *in situ* (Fig. 1C and D).

CB1-mediated signaling also regulates K6 expression in cultured, hyperproliferative human keratinocytes

In order to check whether the observed CB1-mediated effects on K6 regulation within intact human skin epithelium depend on intact epithelial-mesenchymal interactions between epidermis and dermis, or are likely to reflect a direct impact of CB1 ligands on epidermal keratinocytes, we next investigated K6 expression in cultured human HaCaT KCs. This transformed human KC line is well-appreciated to constitutively express K6 and to be hyperproliferative (just like human wounded and psoriatic KC) (Balato *et al.* 2012; Farkas *et al.* 2001; George *et al.* 2010; Kim *et al.* 2011; Ronpirin & Tencomnao 2012; Ryle *et al.* 1989; Saelee, Thongrakard & Tencomnao 2011). K6 is expressed in hyper-proliferative cells (Weiss, Eichner & Sun 1984) and both K6 expression and basal layer epidermal KC proliferation are increased in psoriasis lesions (Donetti *et al.* 2012; Griffiths & Barker 2007; Korver *et al.* 2006; Litvinov *et al.* 2011; Mommers *et al.* 2000).- ~~Since~~ HaCaT cells are known to express functional CB1 and CB2 (Leonti *et al.* 2010; Maccarrone *et al.* 2003; Paradisi *et al.* 2008; ~~Toth *et al.* 2011~~), and this had been confirmed previously by our group, both on the gene (RT-PCR analysis) and protein levels (immunocytochemistry and western blotting techniques) (Toth *et al.* 2011). ~~and thus, being~~ are a direct target of CB1-mediated signaling, this makes these KCs not only an instructive cell culture tool for evaluating the direct, dermis-independent role of CB1-mediated signaling in the regulation of keratin expression in human KCs, but may also provide first indications as to how the observed K6 expression could relate to wound healing and/or psoriasis.

In accordance with our human skin organ culture results, ACEA (1 μ M) significantly down-regulated K6 staining intensity of HaCaT cells *in vitro* (Fig. 2A and B). This was abrogated by the co-administration of the selective CB1 antagonist, AM251 (100 nM) (Fig. 2A and B). Unexpectedly, though, AM251 alone had a partial inhibitory effect on K6 expression, although not significant. This may be related to the fact that AM251 is an inverse agonist (Dono & Currie 2012; Fiori *et al.* 2011), and invites further study. The inhibitory effect of

ACEA on K6 protein expression was further confirmed by quantitative RT-PCR (Fig. 2C).

Therefore, while HaCaT cells may exhibit relatively low CB expression levels, at least under our assay conditions, they showed a vigorous response to CB ligands.

The CB1 specific agonist, ACEA significantly decreases human epidermal keratinocyte proliferation *in situ*

However, the observed effects of CB1-mediated signaling on epidermal K6 expression could simply reflect the appreciated anti-proliferative effects of CB1 agonists (Casanova *et al.* 2003; Hermanson & Marnett 2011; Toth *et al.* 2011). Moreover, K6 is overexpressed in hyper-proliferative and wounded KCs (Weiss, Eichner & Sun 1984), and both K6 expression and basal KC proliferation are increased in psoriatic epidermal lesions (Donetti *et al.* 2012; Griffiths & Barker 2007; Korver *et al.* 2006; Litvinov *et al.* 2011; Mommers *et al.* 2000; Navarro, Casatorres & Jorcano 1995). Therefore, we next assessed whether CB1 stimulation by CB1 specific agonist, ACEA, could affect human KC proliferation *in situ*.

Just as we had seen before with the non-selective endocannabinoid, AEA (Toth *et al.* 2011), the CB1-specific synthetic agonist, ACEA indeed significantly decreased human epidermal KC proliferation *in situ*. This effect was abrogated by the CB1-specific antagonist, AM251 (assessed by quantitative Ki-67 immunomorphometry, Fig. 3A and B).

The CB1 specific agonist, ACEA, significantly decreases K6 expression in suprabasal cells in a proliferation-independent manner

Therefore, it needed to be dissected whether or not CB1 also reduces K6 expression in a proliferation-independent manner. This was done by selectively assessing K6 expression in non-proliferative (i.e. Ki-67-negative) epidermal KCs *in situ*. We found that K6 IR within non-proliferative epidermis was also reduced by ACEA (Fig. 4A and B). Furthermore, K6-

expressing cells in the epidermis co-expressed CB1 *in situ* (Figure 4C), suggesting a direct effect of CB1-agonists on K6-expressing human epidermal KCs *in situ*. Thus, CB1 stimulation may affect K6 expression both, by reducing KC proliferation and by downregulating K6 expression directly *via* CB1 in a proliferation-independent manner.

Here we provide the first evidence that CB1-mediated signaling directly regulates K6/16 expression within normal human skin. Specifically, we show that CB1 stimulation downregulates expression of the hyper-proliferation-associated human keratins K6 *in vitro* and *in situ*, and inhibits human epidermal KC proliferation *in situ*. ~~Moreover we demonstrate that CB1-mediated signaling may regulate K6 expression both directly and indirectly (i.e. via inhibiting KC proliferation).~~

The effect of CB-mediated signaling in human KC biology remains to be fully explored. As we have also observed in isolated human skin (Fig. 4C), CB1 protein expression is detected mainly above the basal layer of the epidermis (Stander *et al.* 2005), i.e. above the compartment where KC proliferation normally occurs most prominently. Wilkinson and Williamson reported that the non-selective CB agonist HU210 inhibited KC proliferation. However, this could not be blocked by either CB1 or CB2 antagonists, suggesting that cannabinoids may also inhibit human KC proliferation through a non-CB1/CB2 mechanism (Wilkinson & Williamson 2007). Nevertheless, it has been previously shown that AEA, which can interact with CB1 on human KC (Biro *et al.* 2009), inhibited human KC proliferation *in situ* and *in vitro* (Toth *et al.* 2011).

Therefore, it was important to clarify whether specific CB1 stimulation inhibits human epidermal KC proliferation *in situ*. By using CB1-specific agonists and antagonists we clearly demonstrate that exclusive CB1 stimulation inhibited KC proliferation. Thus, CB1 is an important key regulator of human KC proliferation. Given the role of epidermal hyperproliferation in the pathobiology of psoriasis (Donetti *et al.* 2012; Griffiths & Barker

2007; Korver *et al.* 2006; Litvinov *et al.* 2011; Mommers *et al.* 2000; Navarro, Casatorres & Jorcano 1995), cannabimimetic agents that activate CB1, therefore, deserve consideration as a novel pharmacological strategy for treating psoriasis.

Furthermore, increased numbers of activated mast cells are often observed in and around lesional psoriatic skin, and increasing evidence suggests that mast cells are functionally important key immunocytes in the pathogenesis of psoriasis (Carvalho, Nilsson & Harvima 2010; Namazi 2005; Radosa *et al.* 2011; Suttle *et al.* 2012; Toruniowa & Jablonska 1988). Recently, we have shown that CB1 activation limits excessive mast cell activity and even inhibits mast cell maturation of resident, intracutaneous progenitors (Sugawara *et al.* 2012). Therefore, besides their anti-proliferative effects on human epidermal KCs, the anti-inflammatory (Richardson, Kilo & Hargreaves 1998) and mast cell-inhibitory properties of CB1 agonists in human skin (Sugawara *et al.* 2012) make them a particularly attractive class of agents in future psoriasis management.

It should be noted, that the constitutive level of K6 expression in organ-cultured human skin fragments is considerable higher than normal scalp skin *in vivo*. Presumably, this occurs as a response to tissue dissection and organ culture, which is well-known to elicit an immediate wound healing response in the epithelium. The latter rapidly starts to migrate over the wound edge in an attempt to enwrap the exposed skin mesenchyme (epiboly phenomenon [Stenn 1981; Brown C *et al.* 1991]). This constitutive up-regulation of K6 in organ-cultured normal human skin may greatly heighten the sensitivity to K6 expression-modulatory compounds, such as CB ligands, thus making skin organ culture a particularly sensitive and instructive tool for keratin research. At the same time, however, caution is advised in extrapolating from these observation made in what essentially reflects a wound healing milieu to healthy, unmanipulated human skin *in vivo*.

The current findings invite the speculation that the therapeutic down-modulation of K6 and/or K16 expression by CB1 agonists and other cannabinomimetics might become exploitable for the management of other dermatoses besides psoriasis, for example pachyonychia congenita (Hickerson *et al.* 2011; Zhao *et al.* 2011) and acne (Biro *et al.* 2009), and could be used to modulate KC migration-dependent reepithelialization in wound healing, similar to related findings in periodontal and intestinal wound repair (Kozono *et al.* 2010; Wright *et al.* 2005). Besides in psoriasis, epidermal K6 and K16 expression are also up-regulated under wound-healing conditions (Coulombe *et al.* 2004; Ishikawa *et al.* 2010; Luo *et al.* 2011). These intermediate filaments provide important structural support for the KCs that participate in epithelial regeneration during the wound healing process (Mazzalupo *et al.* 2003; Wong & Coulombe 2003). Intriguingly, K6 is intimately involved in regulating and optimizing epithelial cell migration by dynamically binding to Src (Rotty & Coulombe 2012). Therefore, the novel CB1-mediated *proliferation-independent* regulation of K6/K16 expression in human epidermis *in situ* revealed here may also impact on Src-mediated KC migration, namely under wound healing conditions and/or in hyperproliferative skin disease.

K6 is also constitutively expressed in a number of skin appendages. K6 is expressed in the suprabasal layers of the sebaceous gland duct, where it has been linked with the Hedgehog signaling pathway (Gu & Coulombe 2008). K6 is also expressed in the mammary gland, and K6-positive cells may represent progenitor cells involved in ductal hyperplasia (Nayak & Bhuiya 2011), or even mammary tumor formation (Bu *et al.* 2011). In fact, the potential role of cannabinoids in breast cancer therapy is currently being explored (Caffarel *et al.* 2012). Moreover, mutations in *KRT6A*, *KRT6B*, and *KRT16* underlie pachyonychia congenita, a rare autosomal dominant genodermatosis characterized by nail deformity, leukokeratosis and keratoderma (Wilson *et al.* 2011). Inhibiting expression of the mutated K6 protein may lead to improvement in pachyonychia congenita clinical symptoms (Hickerson *et al.* 2011; Zhao *et al.* 2011). On this basis, the current findings invite the speculation that the therapeutic down-

~~modulation of K6 and/or K16 expression by CB1 agonists and other cannabinomimetics might become exploitable for the management of other dermatoses besides psoriasis, for example pachyonychia congenita (Hickerson *et al.* 2011; Zhao *et al.* 2011) and acne (Biro *et al.* 2009), and could be used to modulate KC migration dependent reepithelialization in wound healing, similar to related findings in periodontal and intestinal wound repair (Kozono *et al.* 2010; Wright *et al.* 2005).~~

Conclusion

Our results suggest that cannabinoids and their receptors constitute a novel, clinically relevant control element of human ~~keratin-K6 and K16~~ expression. Therefore, cannabimimetic agents might be relevant for the treatment of several skin conditions related to aberrant K6/K16 expression, such as psoriasis and wound healing. In addition, the skin organ culture is shown to be a clinically and physiologically relevant model system for investigating the effect of CB1 specific agonist/antagonist on human skin.

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Figure Legends

Figure 1. The CB1 specific agonist, ACEA significantly inhibits K6 and K16 expression *in situ*. (aA) Representative images of K6 immunofluorescence with organ cultured human skin treated with ACEA/AM251 (1-day). (bB) Statistical analysis of K6 immunofluorescence intensity in organ cultured human skin (quantitative immunohistomorphometry, ImageJ); stimulation with ACEA (30 μ M), AM251 (1 μ M) or both for 1-day. n=9-22 skin sections/group. (cC) Representative images of K16 immunohistochemistry with organ cultured human skin samples with ACEA (1-day). (dD) Quantitative K16 immunohistomorphometry within the epidermis of organ-cultured human skin samples after 1-day of stimulation with ACEA 30 (μ M). n=4 skin sections/group. Data are expressed as mean + SEM. *p<0.05, **p<0.01.

Figure 2. The CB1 specific agonist, ACEA significantly inhibits K6 expression in cultured HaCaT cells. (aA) Representative images of K6 immunofluorescence of cultured HaCaT KCs with ACEA (1 μ M), AM251 (100 nM) or both for 1-day. (bB) Statistical analysis

of K6 immunofluorescence intensity of cultured HaCaT cells. n=6 colonies/group (eC)
 Statistical analysis of *K6* gene expression in HaCaT cells treated with vehicle control or
 ACEA (1 μ M) for 8h. Data are expressed as mean + SEM. *p<0.05; **p<0.01; ***p<0.001.

Figure 3. The CB1 specific agonist, ACEA significantly decreases human epidermal keratinocyte proliferation *in situ*. (aA) Representative images of K6 (green) and Ki67 (red) double-immunofluorescence with organ cultured human skin treated with ACEA/AM251 (1-day). (bB) Quantitative analysis of the percentage of Ki67+ KCs within organ cultured human epidermis. *p<0.05; ***p<0.001. n=5-12 skin sections/group.

Figure 4. The CB1 specific agonist, ACEA, significantly decreases K6 expression in suprabasal cells in a proliferation-independent manner. (aA) Representative images of K6 (green) and Ki-67 (red) double immunofluorescence. Dotted rectangles indicate the reference area for quantitative immunohistomorphometry of K6 fluorescence intensity. (bB) Quantitative analysis of K6 fluorescence intensity in non-proliferating (i.e. Ki67-negative) cells within human epidermis *in situ*. Data are expressed as mean + SEM. *p<0.05. n=5-7 skin sections/group. (eC) K6 (green) and CB1 (red) double-immunofluorescence study. Yellow arrows denote double-positive KCs.