

Hibernation stage predicts climatic responsiveness in noctuid moths

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| Keywords: | climate change, phylogenetic control, model selection |
| Abstract: | Emergence phenology has been shown to advance considerably in the past decades in many lepidopterans. Noctuid moths constitute a species-rich family of lepidopterans with a unique diversity of life history traits driving climatic responsiveness. In our study we aim to assess the role of life-history and ecological traits in climatic responsiveness of noctuid moths (Noctuidae) with a control for phylogenetic dependence. We used a long-term dataset of European noctuid moths collected by a light-trap in North-Eastern Hungary. As the study site is located in the intersection of several biogeographical zones harbouring a large number of noctuid moth species, our dataset provides a unique possibility to investigate the moths' climatic sensitivity. To estimate the role of life-history traits and ecological factors in driving lepidopterans' response to climatic trends, we employed three proxies related to the species' ecology (habitat affinity, food plant specialization and food type) and two robust types of life-history traits (migration strategy, and hibernation form). The degree of temporal shifts of various measures of emerging phenology was primarily related to hibernation stage, food type and migration strategy. Large-scale phylogenetic relatedness exerted little constraint in all models fitted on all measures of capture dates. Our results imply that noctuid moths overwintering as adults exhibited greater degrees of phenological shifts than species hibernating as larvae or pupae. It follows that this group is forced to hibernate as imagos in our climate and the prolonged autumnal activities of the last emerging individuals might be interpreted as a return |

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| | <p>to their original life-cycle. The missing subfamily-level phylogenetic signal in climatic responsiveness of noctuid moths implies that responses to climatic trends are primarily governed by life-history and ecological adaptations rather than large-scale phylogenetic relatedness.</p> |
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1 **Hibernation stage predicts climatic responsiveness in noctuid moths**

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22 Running head: Climatic responsiveness of noctuid moths

23 **Summary**

24

25 Emergence phenology has been shown to advance considerably in the past decades in
26 many lepidopterans. Noctuid moths constitute a species-rich family of lepidopterans with a
27 unique diversity of life history traits driving climatic responsiveness. In our study we aim to
28 assess the role of life-history and ecological traits in climatic responsiveness of noctuid moths
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31 is located in the intersection of several biogeographical zones harbouring a large number of
32 noctuid moth species, our dataset provides a unique possibility to investigate the moths'
33 climatic sensitivity. To estimate the role of life-history traits and ecological factors in driving
34 lepidopterans' response to climatic trends, we employed three proxies related to the species'
35 ecology (habitat affinity, food plant specialization and food type) and two robust types of
36 life-history traits (migration strategy, and hibernation form). The degree of temporal shifts of
37 various measures of emerging phenology was primarily related to hibernation stage, food type
38 and migration strategy. Large-scale phylogenetic relatedness exerted little constraint in all
39 models fitted on all measures of capture dates. Our results imply that noctuid moths
40 overwintering as adults exhibited greater degrees of phenological shifts than species
41 hibernating as larvae or pupae. It follows that this group is forced to hibernate as imagos in
42 our climate and the prolonged autumnal activities of the last emerging individuals might be
43 interpreted as a return to their original life-cycle. The missing subfamily-level phylogenetic
44 signal in climatic responsiveness of noctuid moths implies that responses to climatic trends

45 are primarily governed by life-history and ecological adaptations rather than large-scale
46 phylogenetic relatedness.

47

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49 **Introduction**

50

51 During the past decades an increasing number of studies have reported about the
52 effects of global climatic change on ecosystem structure and functioning (Parry et al. 2007,
53 Walther 2010, Burrows et al. 2011). These effects influence phenology, population dynamics
54 and life-history traits in a broad range of organisms, including a large number of endangered
55 taxa (Parmesan 2006, Forrest and Miller-Rushing 2010, Jenouvrier and Visser 2011).
56 Specifically, evidence is accumulating that temporal shifts in insect phenology as a response
57 to climatic alteration has accelerated over the past few decades (Singer and Parmesan 2010,
58 Hodgson et al. 2011). For instance, emergence phenology has been shown to advance
59 considerably in the past decades in many lepidopterans (Stefanescu et al. 2003, Hekkinen et
60 al. 2010, Diamond et al. 2011) and, studies have recently detected that the annual number of
61 generations of night-active moths has changed over the last decades (Altermatt 2010a, Kocsis
62 and Hufnagel 2011, Gimesi et al. 2012). Global warming is assumed to be one of the most
63 probable drivers of the increasing number of generations presumably by allowing more time
64 for activity (Altermatt 2010a). Nocturnal moths have also been found to respond strongly to
65 global warming by advancing spring flight periods but an immense number of species
66 diversely respond to climatic changes in this taxa (Hodgson et al. 2011, Valtonen et al. 2011).

67 European noctuid moths constitute an ideal group for studying the effects of climatic
68 trends on insect phenology, as they represent one of the most species-rich insect families
69 subdivided into a large number of subfamilies and tribes characterised by a high diversity of
70 life history, possible related to the observed variable climatic responsiveness (Karsholt and

71 Razowski 1996, references in Appendix 1). A specific explanation for variation in responses
72 may be that the ability of lepidopterans to track food plant phenology is often constrained by a
73 number of ecological and life history factors (van Asch and Visser 2007, Altermatt 2010b,
74 Breed et al. 2013). As a consequence, declining population sizes in butterflies (and also in
75 moths) are often attributed to spatial mismatches between their distribution area and changing
76 climatic envelopes, leading to regional or total extinctions of entire subspecies (Devictor et
77 al. 2012).

78 The developmental stage in which individuals hibernate has already been shown to
79 affect climatic responsiveness. For instance, species overwintering as adults were shown to
80 exhibit higher degrees of phenological flexibility than lepidopterans hibernating as larvae or
81 pupae, as these taxa can emerge immediately after hibernation (Diamond et al. 2011).

82 Recent studies have detected that climatic warming seem to enhance the range of
83 migratory movements of some moth species, as migrant populations might track climatic
84 trends faster owing to their increased mobility (Sparks et al. 2007).

85 Evidence is accumulating that climatic responsiveness shows contrasting regional
86 patterns across Europe, both in the frequency of supplementary generations and temporal
87 patterns of emergence phenology of lepidopterans (Tobin et al. 2008, Altermatt 2010a, Pöyry
88 et al. 2011, Kharouba et al. 2014). In our study we aim to assess the role of life-history and
89 ecological traits in climatic responsiveness in a species-rich lepidopteran family (Noctuidae)
90 with a control for phylogenetic dependence in eastern Europe where such investigations are
91 still scarce. To accomplish this objective, we used a long-term dataset of European noctuid
92 moths collected by a light-trap in Jósvalfö (Aggtelek karst, North-Eastern Hungary, 48.5°N,

93 20.51°E, 287 m a.s.l). This area provides a unique opportunity to investigate the moths'
94 climatic sensitivity, as this part of the Carpathian basin is located in the intersection of several
95 biogeographical zones harbouring 407 noctuid moth species, an unusually high number in
96 Central Europe (Varga 1999, 2012). Furthermore, the study area is surrounded by a broad
97 variety of habitats hosting a set of noctuid taxa highly representative for the region.

98 Although a recent study has shown that many life-history traits might have important
99 role in climatic responsiveness in lepidopterans, we aim to assess only those which are more
100 likely to be relevant for noctuid moths (Radchuk et al. 2013). Hence, we employed three
101 proxies related to the species' ecology (habitat affinity, characterised as habitat specialist or
102 generalist; food plant specialization and food type, defined as herbaceous or woody) and two
103 robust types of life-history traits (migration strategy, and hibernation form) already shown to
104 affect the climatic sensitivity of lepidopterans (Forrest and Miller-Rushing 2010, Diamond et
105 al. 2011).

106 Specifically, our hypotheses are as follows. (1) Species with broader habitat range
107 would be less responsive to climatic change, as these noctuids are less constrained by climate-
108 induced changes in habitat phenology. (2) Food-generalist noctuid moths would exhibit
109 smaller degrees of advancement in emergence phenology (as estimated by capture dates) than
110 specialist species as they are less dependent on the phenology of one or a few specific food
111 plants. (3) Migrant moths would track climatic variability to a greater degree than resident
112 ones, as they have more developed sensitivity to weather conditions promoting flexible onsets
113 of northward migration (Sparks et al. 2007, Alerstam et al. 2011). (4) Finally, noctuids which
114 hibernate as adults might track climatic trends faster than species hibernating as larvae or

115 pupae as these moths can emerge immediately after finishing the hibernation (Nyman et al.
116 2012).

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118

119 **Materials and methods**

120

121 *Study area*

122

123 The moth fauna of the Aggtelek karst area has been intensively surveyed over the last
124 decades of the last century (Varga 1999, Szabó et al. 2007). We collected data during a long
125 term light trap survey. The light trap was established in 1988 on the North-East-facing slope
126 of the Tohonya Valley in Aggtelek National Park, North-East Hungary (48.1° N20.51° E, 287
127 m). The light trap was located in an abandoned orchard, presently used as a semi-dry
128 haymeadow, surrounded by scattered rests of karstic oak-hornbeam (*Waldsteinio-Carpinetum*)
129 and xerothermic oak forests (*Corno-Quercetum pubescentis*) and hazel shrub (*Coryletum*
130 *avellanae*) communities (Szabó et al. 2007).

131

132

133 *Climate data*

134

135 Weather data were recorded by an automatic meteorological station, located 50 meters
136 from the light trap. Out of a numerous set of climatic variables, daily mean temperature and

137 daily precipitation were available for the studied period.

138 As it was not possible to obtain meteorological data between 1996-2005, we compared
139 existing data with those of the nearest meteorological station at Miskolc (N48.1, E20.4), 60
140 kilometres from the light-trap by computing Pearson correlation coefficients. Since the
141 strength of correlation between the monthly mean temperatures and precipitation sums was
142 highly significant (median $r=0.89$, minimum $r=0.68$, all $p < 0.01$) for all months, we
143 considered meteorological data from Miskolc representative for the study site. To assess
144 temporal patterns in climatic predictors relevant for lepidopteran biology, we calculated
145 monthly and seasonal (March to May for spring, June to August for summer, September to
146 November for autumn and December to February for winter) means and variances of daily
147 average temperature and monthly as well as seasonal precipitation sums and variances.
148 Further, we computed temporal patterns in cumulative temperature, a biologically more
149 relevant predictor of climatic responsiveness than temperature itself (Pöyry et al. 2011). The
150 computation went as follows: We set a temperature limit of 5.0°C, and summed daily average
151 temperature values above this limit for the following periods in each year,: (1) January 1-
152 March 31 (2) January 1-April 31 (3) January 1-May 31 (4) March 1-May 31. To detect
153 temporal trends in weather proxies, we fitted linear regressions on these variables as a
154 function of years.

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158

159 *Noctuid data*

160

161 The Jermy-type light trap (Szabó et al. 2007) equipped with a 125 W mercury bulb
162 was activated in 1988 and operated in each year for a 20 year period until 2007. In each year
163 the light trap was operated from the first days of March until the first days of November (with
164 a small, 1-2 day variation per year due to accidentally unfavourable weather conditions),
165 totalling 4788 days (mean \pm SD = 239.4 \pm 39 days per year). Captured specimens were
166 collected on a daily basis and were stored in cotton layers or pinned until identification.
167 Species-level identification was carried out by SSz and ZV and the data were entered into a
168 database by recording date, taxon and number of individuals.

169 During the study period 128,045 individuals of 417 taxa were collected. To acquire
170 robust measures on climatic responsiveness, we used only the data of species with records
171 from at least ten years spanning for at least 15 years, yielding 200 species in total. In the
172 following step we retained only species with more than twenty individuals caught in total in
173 every year, in order to obtain reliable estimates of the annual population development. As a
174 result, we used the records of 70 species in subsequent analyses (filtered species hereafter;
175 Appendix 2). To test the robustness of the above filtering conditions, we removed noctuids
176 with less than ten distinct capture dates per year from the set of 200 species and repeated all
177 analyses.

178 To test the effects of first and final operation dates of the light trap, we calculated the
179 distribution of capture dates of filtered species. This shows that the percentage of first and last
180 capture dates within seven days from first and last operation dates, respectively, were less

181 than 0.1%. Therefore it is unlikely that between year variation in first and final operation
182 dates can influence phenological patterns.

183 To derive standard measures of climatic responsiveness for each species, we calculated
184 slopes of linear regressions fitted on years using the following estimates of emergence
185 phenology as response variables: (1) first, (2) median and (3) last julian dates (day-of-year
186 number) of capture and (4) activity length (defined as the number of days between first and
187 last capture dates), calculated using corresponding capture dates. We refer to these response
188 variables as climatic responsiveness of first, median and last julian days as well as activity
189 length. High slopes from these regressions indicate that a given species can faster respond to
190 climatic changes. The linear regression slopes were then applied in phylogenetic least squares
191 models as response variables to investigate what ecological and life history traits influence
192 climatic responsiveness.

193

194

195 *Ecological and life-history predictors*

196

197 We obtained data on ecological variables and life-history traits from the series of
198 Noctuidae Europae (Ronkay et al. 2001, Hacker et al. 2002, Fibiger et al. 2009, Fibiger et al.
199 2010). Habitat affinity was classified as specialist or generalist. Feeding strategy was
200 characterized by diet specialization (categorised as food specialist or food generalist) and food
201 type (classified according to the food type of caterpillars: herbaceous or woody). Migration
202 strategy has the categories of resident and migratory. Hibernation type was classified as

203 imago, pupal and larval stages (Varga et al. 2005).

204 All used data on the species' life history and ecology are presented in Appendix 2.

205

206

207 *Phylogenetic and statistical analyses*

208

209 In the first step we constructed a composite tree (Appendix 3) using the data of the
210 most recent molecular survey applying gradual branch lengths (Mitchell et al. 2006, Zahiri et
211 al. 2011). To account for the lack of knowledge of species-level tree topology, we included
212 only tribal-level information provided by the most recent molecular phylogenetic analyses
213 (Mitchell et al. 2006, Zahiri et al. 2011).

214 To evaluate the degree of association between climatic responsiveness measures and
215 ecological and life-history predictors, we fitted Phylogenetic Generalized Least Squares
216 (PGLS) models controlling for the non-independence among taxa by incorporating a
217 variance–covariance matrix that represents their phylogenetic relationships (Martins and
218 Hansen 1999, Pagel 1997, 1999). The degree of phylogenetic dependence (λ) was set to the
219 most appropriate value obtained for each model by likelihood ratio statistics (Freckleton et al.
220 2002). In the following step we calculated the strength of phylogenetic relatedness in each
221 trait separately by quantifying the maximum-likelihood value of λ with its statistical
222 significance estimated by likelihood ratio tests (i.e. by comparing the log-likelihood of the
223 model in which λ is set to the most appropriate value to the log-likelihood of a model in
224 which $\lambda = 0$). As recent studies on the phylogeny of Noctuidae have suggested a number of

225 major changes in the structure of the phylogenetic tree (references provided in Appendix 4),
226 we repeated assessing phylogenetic signal in response and predictor variables by formulating
227 linear mixed models that include only genus nested in subfamily as a random term, and
228 compared model performance by ANOVA tests.

229 After model fitting, the relative importance of life-history traits and ecological
230 variables was calculated using information-theoretic model-comparison (Burnham and
231 Anderson 2002). First we obtained the values of Akaike's information criterion corrected for
232 small sample size (AIC_c) and the corresponding Akaike weight of each model (ω). Second, we
233 selected models with substantial support ($\Delta_i = AIC_i - AIC_{\min} < 2.0$) and calculated model-
234 averaged parameter estimates (β) and unconditional standard errors (SE_u ; Burnham and
235 Anderson 2002) of each variable by the sums of their Akaike weights across all models with
236 substantial support containing the given predictor. Akaike differences in the range 0-2 indicate
237 substantial level of empirical support of a given model, whereas $\Delta_i > 7$ implies very weak
238 support (Burnham and Anderson 2002).

239 For all analyses we used the R statistical computing environment (R Development
240 Core Team, 2013) including its "ape" (Paradis et al. 2011), "geiger" (Harmon et al. 2009),
241 "MuMIn" (Barton 2011) and "nlme" (Pinheiro et al. 2009) packages.

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243

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246

247 **Results**

248

249 *Climatic patterns*

250

251 During the 20 years of data collection monthly mean temperature has significantly
252 increased in June ($b = 0.140$ °C/year, $F_{1,17} = 7.292$, $p = 0.015$). Seasonal mean and cumulative
253 temperature has not shown significant trends. Further, we detected no clear temporal patterns
254 in variance of monthly temperature and all precipitation metrics (results not shown).

255

256

257 *Temporal trends in capture data*

258

259 Noctuids showed a considerable amount of variation in slopes of various estimates of
260 emergence phenology as a function of years. Out of the 70 taxa in our study, 38 shifted their
261 first capture to earlier dates with four of them exhibiting significant advance. Median capture
262 dates, indicative of the climatic responsiveness of the bulk of the population, showed similar
263 patterns with 34 noctuids emerging earlier but the trend was only significant for a single
264 species. Last julian dates of annual records have advanced in 27 taxa, four of which were
265 significant (Appendix 2).

266 In contrast, first capture of 32 noctuids shifted to later dates with only a single species
267 showing significant patterns. Median capture date was detected to change to later dates in 36
268 noctuids including three species with significant trends. Last capture dates of 43 noctuids

269 have shifted later, five of which exhibited significant temporal patterns. Although activity
270 length shortened in 28 species with significant trends in three species, it became longer in 42
271 noctuids including significant relationships for nine taxa (Appendix 2).

272 On average, noctuids shifted their first capture dates by 4.705 (SE=0.902) days to later
273 dates after 1998 than in the first half of the study period across all species considered.

274 Similarly, median capture occurred 4.525 (SE=2.210) days later in the second period than in
275 the first one. In contrast, overall last emergence shifted by 7.547 (SE=1.325) days to later
276 dates, representing a delay almost twice as long as the delay at the beginning of the flight
277 season. As a result, activity length increased by 2.842 (SE= 1.151) days calculated over the
278 complete set of species.

279

280

281 *Ecological and life history correlates of phenological change*

282

283 Phylogenetic signal (λ) was zero for all measures of all response and proxy variables
284 (Table 1). Quantitatively similar results were obtained by linear mixed models with the
285 random factor genus nested within subfamily (results not shown).

286 First capture dates were affected by significant effects of food type and migration
287 strategy – parameter estimates of which had confident intervals not overlapping with zero.
288 Noctuids with larvae feeding primarily on woods are shifting first emergence to later dates
289 while species feeding on herbaceous plants advance first capture dates ($\Sigma = 0.826$; Table 2, 3).

290 Our results show that migratory strategy has emerged as the second most important

291 proxy of climatic responsiveness based on its Akaike sums in PGLS-models fitted on first
292 capture dates ($\Sigma = 0.749$; Table 2, 3): resident noctuids advanced climatic responsiveness
293 measured as first capture dates to a greater degree than migratory species. Parameter
294 estimates for all other variables have confidence intervals overlapping with zero, therefore we
295 consider their effects as non-significant.

296 Overall, hibernation stage has emerged with the highest Akaike scores calculated over
297 supported models fitted on the slope of last capture dates ($\Sigma = 1.0$, Table 2). Hibernation stage
298 was the only predictor of last capture dates and activity length in the best supported models:
299 species hibernating as imagos shifted last capture to later dates and prolonged activity length
300 to a greater degree than noctuids hibernating as larvae or pupae (Table 2, Appendix 5; Fig 1,
301 Fig 2).

302 We obtained qualitatively similar results for species with more than ten capture dates
303 per year (results not shown.)

304

305

306 Discussion

307

308 In this study we aimed to (i) describe the extent of phenological responses presumably
309 caused by ongoing climate change in a species-rich group of noctuid moths and (ii)
310 investigate the possible ecological and life history correlates of these responses.

311 Considering noctuid phenology, we detected no clear patterns in changes of emergence
312 phenology, which might be related to the diversity in a number of life history traits of the

313 studied species.

314 Our findings show that the degree of temporal shifts of various measures of emerging
315 phenology was primarily related to hibernation stage, food type and migration strategy.
316 Specifically, hibernation stage is the strongest predictor of climatic responsiveness measured
317 as last capture dates and activity length in noctuid moths -, similarly to the results of Diamond
318 et al. (2011). This might indicate that noctuid moths overwintering as adults exhibited greater
319 degrees of phenological shifts than species hibernating as larvae or pupae. However, the
320 difference between the reactions of the bulk of the population and last active individuals to
321 climatic trends seems to be different in our case. Since noctuids hibernating as adults showed
322 no shifts in median emergence to earlier dates, the last individuals of these moths have
323 prolonged capture dates over the study period in comparison to species hibernating in larval
324 or pupal form. Prolonged autumn activity may allow longer time for replenishing fat reserves
325 before hibernation which can increase survival chances and improve reproductive success.
326 Parallely, all of these species are winter-active moths in the southern part of their distribution
327 areas which group is forced to hibernate as imagos in our climate and the prolonged autumn
328 activities of the last emerging individuals might be interpreted as a return to their original life-
329 cycle (László et al. 2000).

330 Food type was included in best supported models for climatic responsiveness
331 measured as first capture dates: noctuids feeding on herbaceous plants advanced first capture
332 dates to a greater degree than species connected to woody materials. Although a recent survey
333 of British plants showed that annuals were more likely than perennials to advance phenology
334 (Fitter and Fitter 2002), our results imply that moths feeding on herbaceous plants are able to

335 track plant phenology faster than noctuids feeding on woody plants which evokes the
336 following hypotheses. First, as herbaceous plants are shorter-lived on average than woody
337 taxa, their herbivores are forced to synchronise phenology to a greater degree than moth
338 larvae feeding on woody plants. Second, woody plants are on average larger than herbaceous
339 species implying higher levels of phenological variance of plant parts consumed by
340 herbivores in individual trees which allows for a wider range of phenological synchronisation.
341 However the complexity of climatic responses driven by trophic interactions is enhanced by
342 that the highly diverse family of Noctuidae is connected to a great variety of larval food
343 plants. For instance, majority of these species is connected to forested habitats and majority of
344 spring- and autumn-active moths are linked to woody food plants (details shown in Appendix
345 2). A large group of genera (Appendix 2) use a wide variety of food plants which are most
346 often herbaceous (grasses for mostly monovoltine *Apamea*, and often Urticaceae for the
347 phenologically more plastic Plusiinae), or partly herbaceous and woody (Acronictinae)
348 (Appendix 2).

349 Migration strategy has emerged as a supported predictor of climatic responsiveness.
350 Migrant noctuids have shifted first captures to later dates than resident moths which implies
351 that migratory species arriving from southern regions may detect warming spring weather
352 later than locally hibernating taxa. (Alerstam et al. 2011). Indeed, migrant moths need time to
353 reach Central Europe and thus are outcompeted in advancements of climatic responsiveness
354 measured as first capture dates by locally wintering moths hibernating as imagos.

355 Throughout our analyses we detected no phylogenetic relatedness on the subfamily
356 level in climatic responsiveness of noctuid moths. This result is in line with a number of

357 investigations showing that the advancement in spring phenology is not driven by common
358 descent in a broad range of taxa (Végvári et al. 2010, Diamond et al. 2011). The lack of
359 phylogenetic signal in emergence phenology of the species in our dataset evokes several
360 explanations (1) climatic sensitivity is a species-specific response (2) phenotypic plasticity
361 might play a role in driving phenology dynamics as a response to climatic variability,
362 paralleling the findings of recent studies across a number of taxonomic levels (Hoffmann and
363 Sgrò 2011). However, the resolution of current phylogenetic studies of noctuid moths does not
364 allow us to draw more general conclusions on the existence of phylogenetic signal in climatic
365 responsiveness of night-active lepidopterans. The similar results of Diamond et al. (2011)
366 showing that phylogenetic relatedness among species predicted very little variation in
367 butterflies' phenological responses imply that climatic responsiveness in lepidopterans can be
368 treated as a species-specific reaction rather than driven by evolutionary relatedness. Further,
369 lacking phylogenetic relatedness in climatic sensitivity probably contributes to fast
370 adaptations to global change, observed in a wide range of insect taxa (Altermatt 2010a).

371 Our study implies recommendations for lepidopteran conservation, as a reflection to
372 current declining trends in a number of insect species on a global scale. We found that noctuid
373 moths hibernating as adults are able to track climatic trends faster than species overwintering
374 in less developed stages which suggest higher declining probabilities in these taxa,
375 considering current climatic predictions (Stocker et al. 2013). Our result that resident moths
376 are responding to ongoing climatic patterns to a greater degree than migratory taxa call for
377 more intensive survey of migrating lepidopteran populations.

378 In sum, we have demonstrated in a diverse set of noctuid moth species that temporal

379 trends in emergence phenology (which is associated with a measure of temperature increase in
380 the study area) is primarily related to hibernation stage, diet specialisation and migration
381 strategy.

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506 **Table 1.** λ -statistics for phylogenetic signal in the investigated response variables and
507 predictors. $\lambda = 0$ indicates no phylogenetic signal.

508
509

| Variable | λ | Likelihood ratio | P |
|------------------------------|-----------|------------------|-------|
| Slope of first capture date | 0.000 | 0.000 | 0.999 |
| Slope of median capture date | 0.000 | 0.000 | 0.999 |
| Slope of last capture date | 0.000 | 0.000 | 0.999 |
| Slope of activity length | 0.000 | 0.000 | 0.997 |
| Migration type | 0.000 | 0.000 | 0.999 |
| Hibernation type | 0.000 | 0.000 | 0.422 |
| Food specialisation | 0.543 | 0.898 | 0.343 |
| Food type | 0.811 | 2.249 | 0.134 |
| Habitat affinity | 0 | 0.001 | 0.98 |

510

511 **Table 2.** Akaike sums, model-averaged parameter estimates (beta), standard errors (SE_u) and
 512 95% confidence intervals for predictors across best supported models ($\Delta_i < 2.0$)

513

| Slope of first emergence date | Slope of median emergence date | Slope of last emergence date | Slope of activity length |
|------------------------------------------------------------------------------------------------------|------------------------------------------------------------------------------------------------------|------------------------------------------------------------------------------------------------------|------------------------------------------------------------------------------------------------------|
| Food type $\Sigma = 0.826$ $\beta(SE_u) = -0.395 (0.145)$ 95% CI = (-0.678; -0.111) | Migration strategy $\Sigma = 0.483$ $\beta(SE_u) = -0.497 (0.575)$ 95% CI = (-1.625; 0.631) | Hibernation stage $\Sigma = 1.0$ $\beta(SE_u) = -2.674 (0.543)$ 95% CI = (-3.737; -1.61) | Hibernation stage $\Sigma = 1.0$ $\beta(SE_u) = -3.018 (0.608)$ 95% CI = (-4.21; -1.826) |
| Migration strategy $\Sigma = 0.749$ $\beta(SE_u) = -0.547 (0.243)$ 95% CI = (-1.023; -0.07) | Food type $\Sigma = 0.404$ $\beta(SE_u) = -0.376 (0.365)$ 95% CI = (-1.091; 0.339) | Migration strategy $\Sigma = 0.246$ $\beta(SE_u) = -0.227 (0.406)$ 95% CI = (-1.023; 0.57) | Migration strategy $\Sigma = 0.282$ $\beta(SE_u) = 0.265 (0.467)$ 95% CI = (-0.651; 1.18) |
| Habitat affinity $\Sigma = 0.296$ $\beta(SE_u) = 0.252 (0.15)$ 95% CI = (-0.099; 0.548) | Habitat affinity $\Sigma = 0.299$ $\beta(SE_u) = 0.206 (0.36)$ 95% CI = (-0.499; 0.912) | Food type $\Sigma = 0.209$ $\beta(SE_u) = -0.217 (0.301)$ 95% CI = (-0.807; 0.373) | Habitat affinity $\Sigma = 0.213$ $\beta(SE_u) = -0.212 (0.287)$ 95% CI = (-0.774; 0.35) |
| Food specialisation $\Sigma = 0.212$ $\beta(SE_u) = 0.224 (0.165)$ 95% CI = (-0.099; 0.548) | Food specialisation $\Sigma = 0.293$ $\beta(SE_u) = 0.052 (0.404)$ 95% CI = (-0.74; 0.843) | Food specialisation $\Sigma = 0.183$ $\beta(SE_u) = 0.151 (0.293)$ 95% CI = (-0.424; 0.725) | Food type $\Sigma = 0.195$ $\beta(SE_u) = 0.099 (0.339)$ 95% CI = (-0.564; 0.763) |
| Hibernation stage $\Sigma = 0.151$ $\beta(SE_u) = 0.386 (0.303)$ 95% CI = (-0.208; 0.981) | Hibernation stage $\Sigma = 0.278$ $\beta(SE_u) = 0.097 (0.755)$ 95% CI = (-1.384; 1.577) | Habitat affinity $\Sigma = 0.142$ $\beta(SE_u) = -0.018 (0.252)$ 95% CI = (-0.512; 0.475) | Food specialisation $\Sigma = 0.186$ $\beta(SE_u) = -0.06 (0.331)$ 95% CI = (-0.708; 0.588) |

514

515 **Table 3.** Supported models ($\Delta_i < 7.0$) fitted on all types of response variables.

516

| response | Model formula | df | AICc | Δ_i | w |
|--------------------------------|------------------------------------------|---------|---------|------------|-------|
| Slope of first emergence date | foodtype + migratory | 4 | 123.811 | 0 | 0.279 |
| | foodtype + habpr2 + migratory | 5 | 124.904 | 1.092 | 0.161 |
| | foodtype | 3 | 125.321 | 1.509 | 0.131 |
| | foodspec + foodtype + migratory | 5 | 125.964 | 2.152 | 0.095 |
| | foodtype + hiber3 + migratory | 6 | 127.997 | 4.186 | 0.034 |
| | foodspec + foodtype + habpr2 + migratory | 6 | 128.104 | 4.293 | 0.033 |
| | foodspec + foodtype | 4 | 128.213 | 4.402 | 0.031 |
| | foodtype + habpr2 + hiber3 + migratory | 7 | 128.514 | 4.702 | 0.027 |
| | migratory | 3 | 128.692 | 4.881 | 0.024 |
| | habpr2 + hiber3 + migratory | 6 | 128.772 | 4.961 | 0.023 |
| | foodspec | 3 | 128.832 | 5.021 | 0.023 |
| | habpr2 + migratory | 4 | 128.974 | 5.162 | 0.021 |
| | hiber3 | 4 | 128.999 | 5.187 | 0.021 |
| | foodspec + migratory | 4 | 129.064 | 5.253 | 0.02 |
| | hiber3 + migratory | 5 | 129.069 | 5.258 | 0.02 |
| | foodtype + habpr2 | 4 | 129.11 | 5.299 | 0.02 |
| | foodtype + hiber3 | 5 | 129.653 | 5.842 | 0.015 |
| habpr2 | 3 | 130.275 | 6.463 | 0.011 | |
| foodspec + hiber3 + migratory | 6 | 130.285 | 6.473 | 0.011 | |
| Slope of median emergence date | migratory | 3 | 246.815 | 0 | 0.123 |
| | foodtype | 3 | 247.018 | 0.203 | 0.111 |
| | foodtype + migratory | 4 | 247.695 | 0.88 | 0.079 |
| | hiber3 | 4 | 247.858 | 1.043 | 0.073 |

| | | | | | |
|------------------------------|------------------------------------------|---|---------|-------|-------|
| | foodspec | 3 | 247.881 | 1.067 | 0.072 |
| | habpr2 | 3 | 247.996 | 1.181 | 0.068 |
| | habpr2 + migratory | 4 | 248.586 | 1.771 | 0.051 |
| | hiber3 + migratory | 5 | 248.847 | 2.032 | 0.044 |
| | foodspec + migratory | 4 | 249 | 2.185 | 0.041 |
| | foodspec + foodtype | 4 | 249.3 | 2.485 | 0.035 |
| | foodtype + habpr2 | 4 | 249.545 | 2.731 | 0.031 |
| | foodtype + habpr2 + migratory | 5 | 249.724 | 2.91 | 0.029 |
| | foodtype + hiber3 | 5 | 249.754 | 2.939 | 0.028 |
| | foodspec + foodtype + migratory | 5 | 250.039 | 3.224 | 0.024 |
| | foodspec + hiber3 | 5 | 250.2 | 3.385 | 0.023 |
| | foodspec + habpr2 | 4 | 250.22 | 3.406 | 0.022 |
| | habpr2 + hiber3 | 5 | 250.382 | 3.568 | 0.021 |
| | foodtype + hiber3 + migratory | 6 | 250.552 | 3.738 | 0.019 |
| | habpr2 + hiber3 + migratory | 6 | 250.76 | 3.946 | 0.017 |
| | foodspec + habpr2 + migratory | 5 | 250.878 | 4.063 | 0.016 |
| | foodspec + hiber3 + migratory | 6 | 251.176 | 4.361 | 0.014 |
| | foodspec + foodtype + habpr2 | 5 | 251.805 | 4.99 | 0.01 |
| | foodspec + foodtype + habpr2 + migratory | 6 | 252.089 | 5.275 | 0.009 |
| | foodspec + foodtype + hiber3 | 6 | 252.117 | 5.303 | 0.009 |
| | foodtype + habpr2 + hiber3 | 6 | 252.41 | 5.595 | 0.007 |
| | foodtype + habpr2 + hiber3 + migratory | 7 | 252.678 | 5.864 | 0.007 |
| | foodspec + habpr2 + hiber3 | 6 | 252.709 | 5.895 | 0.006 |
| | foodspec + foodtype + hiber3 + migratory | 7 | 252.992 | 6.177 | 0.006 |
| | foodspec + habpr2 + hiber3 + migratory | 7 | 253.188 | 6.373 | 0.005 |
| Slope of last emergence date | hiber3 | 4 | 203.804 | 0 | 0.395 |
| | hiber3 + migratory | 5 | 205.899 | 2.095 | 0.139 |
| | foodtype + hiber3 | 5 | 206.245 | 2.44 | 0.117 |

| | | | | | |
|--------------------------|----------------------------------------|---|---------|-------|-------|
| | foodspec + hiber3 | 5 | 206.554 | 2.75 | 0.1 |
| | habpr2 + hiber3 | 5 | 207.16 | 3.355 | 0.074 |
| | foodtype + hiber3 + migratory | 6 | 208.222 | 4.418 | 0.043 |
| | foodspec + hiber3 + migratory | 6 | 208.585 | 4.781 | 0.036 |
| | habpr2 + hiber3 + migratory | 6 | 209.142 | 5.338 | 0.027 |
| | foodspec + foodtype + hiber3 | 6 | 209.159 | 5.354 | 0.027 |
| | foodtype + habpr2 + hiber3 | 6 | 209.627 | 5.823 | 0.022 |
| | foodspec + habpr2 + hiber3 | 6 | 209.815 | 6.011 | 0.02 |
| | | | | | |
| Slope of activity length | hiber3 | 4 | 219.033 | 0 | 0.362 |
| | hiber3 + migratory | 5 | 220.947 | 1.914 | 0.139 |
| | foodtype + hiber3 | 5 | 221.667 | 2.634 | 0.097 |
| | habpr2 + hiber3 | 5 | 221.717 | 2.684 | 0.095 |
| | foodspec + hiber3 | 5 | 221.778 | 2.745 | 0.092 |
| | habpr2 + hiber3 + migratory | 6 | 223.106 | 4.073 | 0.047 |
| | foodtype + hiber3 + migratory | 6 | 223.564 | 4.531 | 0.038 |
| | foodspec + hiber3 + migratory | 6 | 223.695 | 4.662 | 0.035 |
| | foodspec + foodtype + hiber3 | 6 | 224.438 | 5.405 | 0.024 |
| | foodtype + habpr2 + hiber3 | 6 | 224.462 | 5.428 | 0.024 |
| | foodspec + habpr2 + hiber3 | 6 | 224.479 | 5.446 | 0.024 |
| | foodtype + habpr2 + hiber3 + migratory | 7 | 225.877 | 6.844 | 0.012 |
| | foodspec + habpr2 + hiber3 + migratory | 7 | 225.961 | 6.928 | 0.011 |

517

518 Predictors are abbreviated as:

519 hiber3: hibernation stage

520 migratory: migration strategy

521 habpr2: habitat affinity

522 foodtype: food type

523 foodspec: food specialisation

524 **Figure legends**

525

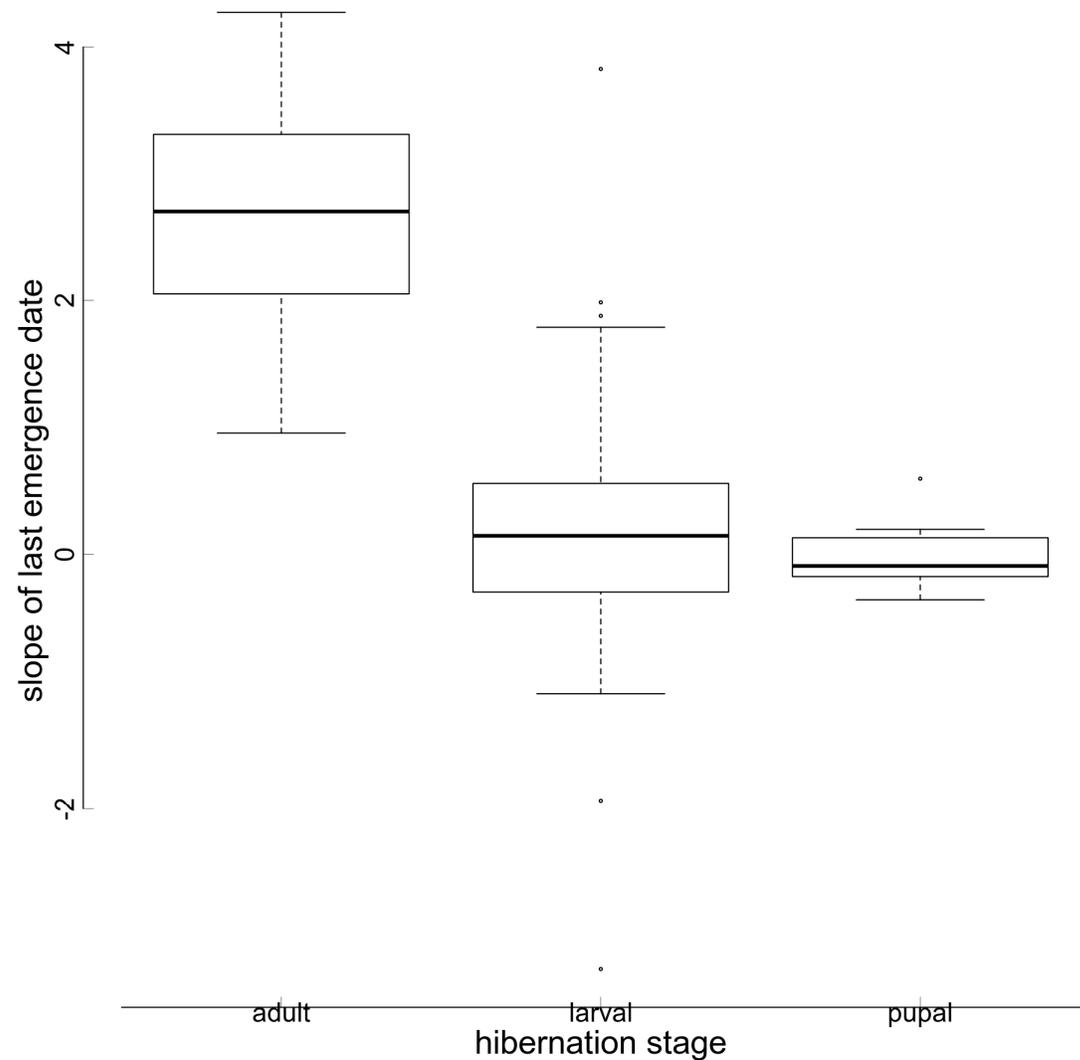
526 **Fig 1.** Slope of last capture date as a function of hibernation stage in noctuid moths.

527

528 **Fig 2.** Slope of activity length as a function of hibernation stage in noctuid moths.

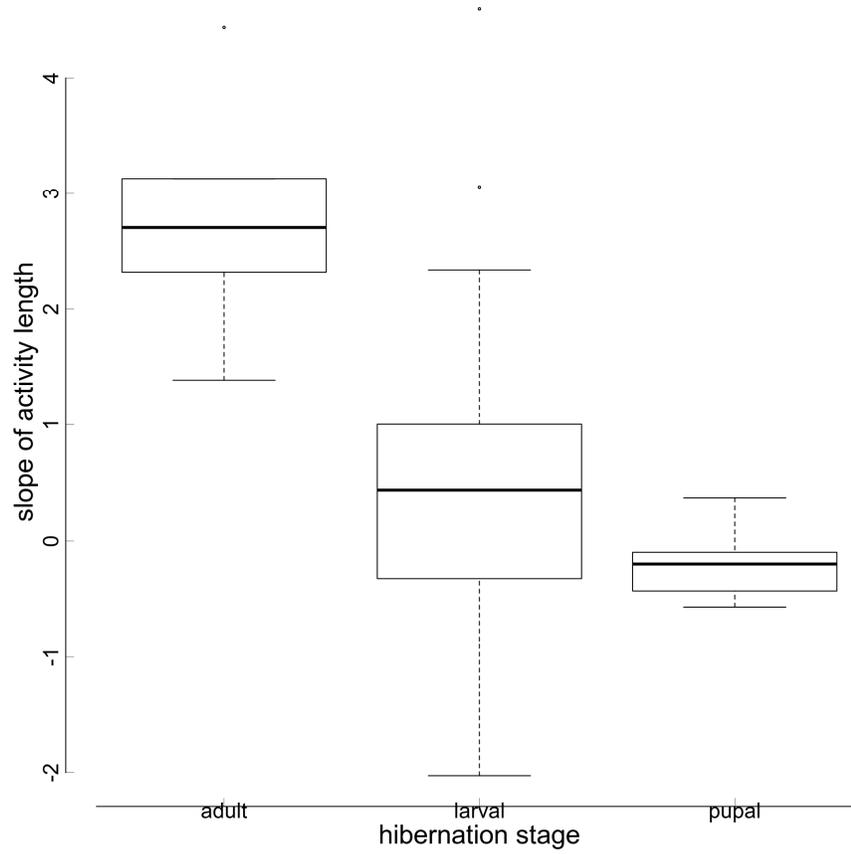
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529 **Fig. 1.**
530



531
532
533

534 Fig. 2.



Only

Appendix 1. References for ecological and life-history traits of Noctuid moths.

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Appendix 2. Life-history traits, ecological predictors and measures of climatic responsiveness of noctuid moths. In the table we use the following abbreviations: FED=first emergence date; LED= last emergence date; MED=median emergence date.

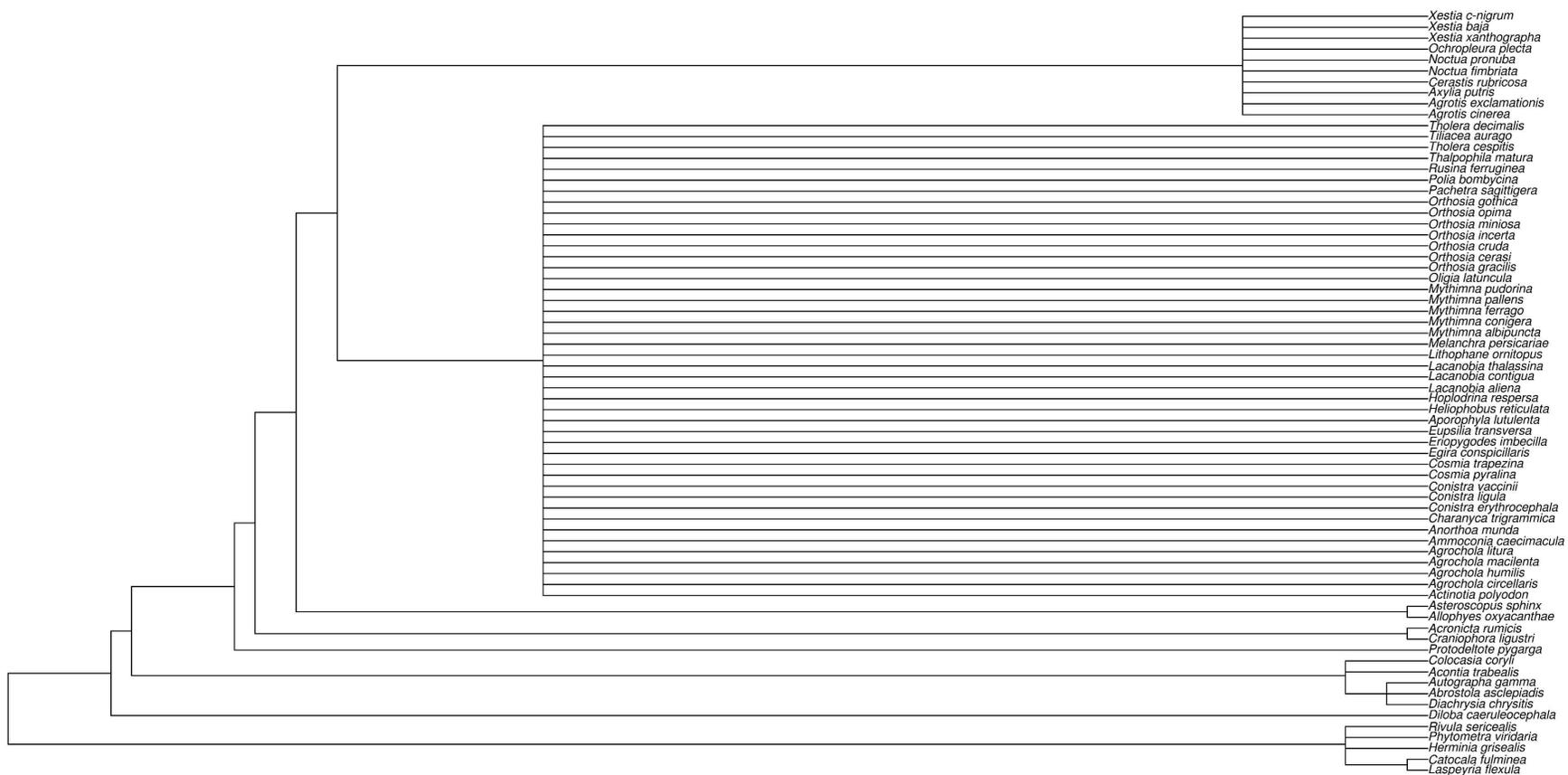
| Taxon | Subfamily | Hibernation stage | Migration strategy | Habitat affinity | Food specialization | Food type | FED slope (p-value) | LED slope (p-value) | MED slope (p-value) | Slope of activity length (p-value) |
|------------------------------|--------------|-------------------|--------------------|------------------|---------------------|------------|---------------------|---------------------|---------------------|------------------------------------|
| <i>Abrostola asclepiadis</i> | Plusiinae | larval | resident | stenoecious | specialist | herbaceous | -0.383 (-0.383) | 1.415 (0.119) | -0.101 (0.949) | 1.798 (0.013) |
| <i>Acontia trabealis</i> | Acontiinae | larval | resident | euryoecious | gene | herbaceous | -1.641 (-1.641) | 1.418 (0.158) | 1.152 (0.697) | 3.058 (0.08) |
| <i>Acronicta rumicis</i> | Acronictinae | larval | resident | euryoecious | generalist | herbaceous | -0.743 (-0.743) | 1.043 (0.26) | -1.155 (0.55) | 1.786 (0.036) |
| <i>Actinotia polyodon</i> | Xyleninae | larval | resident | stenoecious | generalist | herbaceous | 0.253 (0.253) | 0.056 (0.951) | -0.759 (0.619) | -0.198 (0.824) |
| <i>Agrochola circellaris</i> | Xyleninae | egg | resident | stenoecious | generalist | woody | 0.854 (0.854) | 0.562 (0.334) | 1.249 (0.141) | -0.292 (0.743) |
| <i>Agrochola humilis</i> | Xyleninae | egg | resident | stenoecious | generalist | woody | -0.218 (-0.218) | 0.511 (0.053) | 0.537 (0.376) | 0.729 (0.157) |
| <i>Agrochola litura</i> | Xyleninae | egg | resident | stenoecious | generalist | woody | 0.102 (0.102) | 0.567 (0.006) | 0.701 (0.169) | 0.465 (0.277) |
| <i>Agrochola macilenta</i> | Xyleninae | egg | resident | stenoecious | specialist | woody | 0.1 (0.1) | 0.378 (0.444) | 0.103 (0.859) | 0.277 (0.699) |
| <i>Agrotis cinerea</i> | Noctuinae | larval | resident | stenoecious | generalist | herbaceous | -0.24 (-0.24) | -0.663 (0.003) | -0.554 (0.172) | -0.423 (0.131) |
| <i>Agrotis exclamationis</i> | Noctuinae | larval | resident | euryoecious | generalist | herbaceous | -0.774 (-0.774) | 0.387 (0.576) | -0.868 (0.4) | 1.161 (0.097) |
| <i>Allophyes oxyacanthae</i> | Psaphidinae | egg | resident | stenoecious | specialist | woody | -0.141 (-0.141) | 0.804 (0.005) | 0.951 (0.027) | 0.944 (0.018) |
| <i>Ammoconia caecimacula</i> | Xyleninae | egg | resident | stenoecious | specialist | herbaceous | -0.192 (-0.192) | 0.284 (0.336) | -0.801 (0.078) | 0.476 (0.182) |
| <i>Anorthoa munda</i> | Hadeninae | pupal | resident | stenoecious | specialist | woody | 0.494 (0.494) | 0.065 (0.821) | 0.684 (0.276) | -0.429 (0.326) |
| <i>Aporophyla lutulenta</i> | Xyleninae | larval | resident | euryoecious | generalist | herbaceous | 0.074 (0.074) | 1.512 (0.168) | 1.012 (0.606) | 1.438 (0.135) |
| <i>Asteroscopus sphinx</i> | Psaphidinae | egg | resident | stenoecious | specialist | woody | 0.239 (0.239) | 0.348 (0.408) | 0.133 (0.777) | 0.11 (0.764) |
| <i>Autographa gamma</i> | Plusiinae | larval | migratory | euryoecious | generalist | herbaceous | 0.403 (0.403) | -0.608 (0.403) | 1.406 (0.501) | -1.012 (0.326) |

| | | | | | | | | | | |
|--------------------------------|--------------|--------|-----------|-------------|------------|------------|-----------------|----------------|----------------|----------------|
| <i>Axylia putris</i> | Noctuinae | larval | migratory | euryoecious | generalist | herbaceous | -0.377 (-0.377) | -0.057 (0.939) | -0.239 (0.878) | 0.321 (0.654) |
| <i>Catocala fulminea</i> | Catocalinae | larval | resident | stenoecious | specialist | woody | 0.622 (0.622) | -0.812 (0.051) | -0.582 (0.408) | -1.434 (0.001) |
| <i>Cerastis rubricosa</i> | Catocalinae | pupal | resident | stenoecious | generalist | herbaceous | 0.129 (0.129) | -0.107 (0.63) | -0.248 (0.662) | -0.236 (0.623) |
| <i>Charanyca trigrammica</i> | Xyleninae | larval | resident | stenoecious | generalist | herbaceous | -0.679 (-0.679) | -0.143 (0.741) | 0.196 (0.706) | 0.536 (0.319) |
| <i>Colocasia coryli</i> | Pantheinae | larval | resident | stenoecious | generalist | woody | 0.582 (0.582) | 0.15 (0.787) | -0.355 (0.846) | -0.432 (0.432) |
| <i>Conistra erythrocephala</i> | Xyleninae | adult | resident | stenoecious | generalist | woody | -0.163 (-0.163) | 4.273 (0.09) | 0.309 (0.94) | 4.435 (0.092) |
| <i>Conistra ligula</i> | Xyleninae | adult | resident | stenoecious | generalist | woody | 0.182 (0.182) | 3.313 (0.305) | 6.599 (0.094) | 3.131 (0.351) |
| <i>Conistra vaccinii</i> | Xyleninae | adult | resident | stenoecious | generalist | woody | 0.002 (0.002) | 2.706 (0.336) | -0.937 (0.814) | 2.704 (0.328) |
| <i>Cosmia pyralina</i> | Xyleninae | larval | resident | stenoecious | generalist | woody | -0.178 (-0.178) | 1.985 (0.052) | -0.124 (0.877) | 2.163 (0.042) |
| <i>Cosmia trapezina</i> | Xyleninae | larval | resident | euryoecious | generalist | woody | -0.399 (-0.399) | -0.121 (0.767) | -1.723 (0.121) | 0.279 (0.696) |
| <i>Craniophora ligustri</i> | Acronictinae | larval | resident | stenoecious | specialist | woody | -0.735 (-0.735) | 0.531 (0.422) | -0.346 (0.851) | 1.267 (0.145) |
| <i>Diachrysia chrysitis</i> | Plusiinae | larval | resident | stenoecious | generalist | herbaceous | -0.948 (-0.948) | -0.118 (0.881) | -0.84 (0.58) | 0.83 (0.286) |
| <i>Diloba caeruleocephala</i> | Dilobinae | egg | resident | euryoecious | generalist | woody | -0.313 (-0.313) | 0.706 (0.025) | 0.311 (0.523) | 1.019 (0.011) |
| <i>Egira conspicillaris</i> | Hadeninae | pupa | resident | stenoecious | generalist | herbaceous | 0.232 (0.232) | 0.602 (0.167) | 0.367 (0.562) | 0.37 (0.408) |
| <i>Eriopygodes imbecilla</i> | Hadeninae | larval | resident | stenoecious | generalist | herbaceous | -0.424 (-0.424) | 0.097 (0.82) | 0.268 (0.553) | 0.521 (0.24) |
| <i>Eupsilia transversa</i> | Xyleninae | adult | resident | euryoecious | generalist | woody | -0.267 (-0.267) | 2.051 (0.45) | -2.568 (0.491) | 2.318 (0.39) |
| <i>Herminia grisealis</i> | Herminiinae | larval | resident | euryoecious | generalist | herbaceous | -1.237 (-1.237) | -3.263 (0.012) | -2.453 (0.343) | -2.026 (0.224) |
| <i>Heliophobus reticulata</i> | Hadeninae | larval | resident | stenoecious | specialist | herbaceous | 0.002 (0.002) | -0.383 (0.342) | -0.5 (0.213) | -0.385 (0.375) |
| <i>Hoplodrina respersa</i> | Xyleninae | larva | resident | stenoecious | generalist | herbaceous | -0.816 (-0.816) | -1.062 (0.042) | -2.162 (0.056) | -0.246 (0.756) |
| <i>Lacanobia aliena</i> | Hadeninae | larval | resident | stenoecious | specialist | herbaceous | -0.011 (-0.011) | 0.146 (0.812) | 0.404 (0.73) | 0.157 (0.839) |
| <i>Lacanobia contigua</i> | Hadeninae | larval | resident | stenoecious | generalist | herbaceous | -0.828 (-0.828) | 1.281 (0.025) | 3.708 (0.002) | 2.109 (0.004) |
| <i>Lacanobia</i> | Hadeninae | larval | resident | stenoecious | generalist | herbaceous | 0.313 (0.313) | 0.389 (0.61) | -0.29 (0.824) | 0.076 (0.92) |

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|-----------------------|---------------|--------|-----------|-------------|------------|------------|-----------------|----------------|----------------|----------------|
| thalassina | | | | | | | | | | |
| Laspeyria flexula | Aventiinae | larval | resident | stenoecious | generalist | herbaceous | -1.402 (-1.402) | 0.061 (0.898) | -3.215 (0.039) | 1.462 (0.015) |
| Lithophane ornitopus | Xyleninae | adult | resident | stenoecious | generalist | woody | -0.428 (-0.428) | 0.96 (0.181) | -2.277 (0.569) | 1.388 (0.108) |
| Melanchra persicariae | Hadeninae | larval | resident | stenoecious | generalist | herbaceous | -0.025 (-0.025) | -0.223 (0.752) | 0.838 (0.27) | -0.198 (0.785) |
| Mythimna albipuncta | Hadeninae | larval | migratory | euryoecious | generalist | herbaceous | -0.123 (-0.123) | 1.879 (0.059) | 0.207 (0.918) | 2.002 (0.065) |
| Mythimna conigera | Hadeninae | larval | resident | stenoecious | generalist | herbaceous | 0.412 (0.412) | -1.092 (0.005) | -1.489 (0.061) | -1.504 (0.01) |
| Mythimna ferrago | Hadeninae | larval | resident | euryoecious | generalist | herbaceous | -0.922 (-0.922) | 0.064 (0.927) | -0.119 (0.888) | 0.986 (0.213) |
| Mythimna pallens | Hadeninae | larval | migratory | euryoecious | generalist | herbaceous | -2.107 (-2.107) | -1.933 (0.228) | -1.611 (0.451) | 0.175 (0.911) |
| Mythimna pudorina | Hadeninae | larval | resident | stenoecious | generalist | herbaceous | 0.011 (0.011) | 0.933 (0.161) | 1.171 (0.191) | 0.921 (0.186) |
| Noctua fimbriata | Noctuidae | larval | migratory | euryoecious | generalist | herbaceous | 0.686 (0.686) | 0.331 (0.567) | 0.931 (0.498) | -0.355 (0.509) |
| Noctua pronuba | Noctuidae | larval | migratory | euryoecious | generalist | herbaceous | 0.876 (0.876) | -0.33 (0.612) | -0.9 (0.649) | -1.205 (0.308) |
| Ochropleura plecta | Noctuidae | larval | migratory | euryoecious | generalist | herbaceous | -0.034 (-0.034) | -0.13 (0.889) | 0.359 (0.799) | -0.096 (0.911) |
| Oligia latruncula | Xyleninae | larval | resident | stenoecious | specialist | herbaceous | -1.16 (-1.16) | -0.211 (0.669) | -1.55 (0.04) | 0.949 (0.194) |
| Orthosia cerasi | Hadeninae | pupal | resident | euryoecious | generalist | woody | 0.393 (0.393) | -0.174 (0.701) | 0.78 (0.152) | -0.567 (0.198) |
| Orthosia cruda | Hadeninae | pupal | resident | euryoecious | generalist | woody | 0.26 (0.26) | 0.131 (0.749) | 0.087 (0.88) | -0.129 (0.771) |
| Orthosia gothica | Hadeninae | pupal | resident | euryoecious | generalist | woody | 0.208 (0.208) | -0.265 (0.598) | -0.69 (0.256) | -0.474 (0.326) |
| Orthosia gracilis | Hadeninae | pupal | resident | stenoecious | specialist | woody | 0.261 (0.261) | -0.125 (0.735) | 1.192 (0.042) | -0.386 (0.49) |
| Orthosia incerta | Hadeninae | pupal | resident | euryoecious | generalist | woody | -0.197 (-0.197) | -0.356 (0.394) | 1.145 (0.072) | -0.159 (0.7) |
| Orthosia miniosa | Hadeninae | pupal | resident | stenoecious | specialist | woody | 0.238 (0.238) | 0.197 (0.59) | -0.424 (0.411) | -0.041 (0.905) |
| Orthosia opima | Hadeninae | pupal | resident | stenoecious | specialist | woody | 0.023 (0.023) | -0.074 (0.921) | 0.122 (0.893) | -0.097 (0.882) |
| Pachetra sagittigera | Hadeninae | larval | resident | stenoecious | generalist | herbaceous | 0.062 (0.062) | -0.409 (0.585) | -1.058 (0.087) | -0.471 (0.528) |
| Phytometra viridaria | Phytometrinae | larval | resident | euryoecious | generalist | herbaceous | -0.767 (-0.767) | 3.828 (0.504) | 0.690 (0.893) | 4.595 (0.554) |
| Polia bombycina | Hadeninae | larval | resident | stenoecious | generalist | herbaceous | 0.127 (0.127) | -0.477 (0.496) | -0.313 (0.563) | -0.604 (0.45) |

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|-----------------------------|--------------|--------|-----------|-------------|------------|------------|-----------------|----------------|----------------|----------------|
| <i>Protodeltote pygarga</i> | Eustrotiinae | larval | resident | euryoecious | generalist | herbaceous | -0.094 (-0.094) | -0.746 (0.275) | 0.068 (0.956) | -0.652 (0.332) |
| <i>Rivula sericealis</i> | Rivulinae | larval | resident | euryoecious | generalist | herbaceous | -0.841 (-0.841) | -0.306 (0.77) | -1.09 (0.442) | 0.535 (0.623) |
| <i>Rusina ferruginea</i> | Xyleninae | larval | resident | stenoecious | generalist | herbaceous | -0.864 (-0.864) | 0.422 (0.376) | 0.152 (0.809) | 1.286 (0.009) |
| <i>Thalpophila matura</i> | Xyleninae | larval | resident | stenoecious | generalist | herbaceous | 0.221 (0.221) | -0.284 (0.437) | 0.198 (0.689) | -0.505 (0.415) |
| <i>Tholera cespitis</i> | Hadeninae | egg | resident | euryoecious | generalist | herbaceous | 0.102 (0.102) | 0.538 (0.213) | 0.533 (0.1) | 0.436 (0.467) |
| <i>Tholera decimalis</i> | Hadeninae | egg | resident | euryoecious | specialist | herbaceous | -0.025 (-0.025) | 0.114 (0.663) | 0.1 (0.747) | 0.139 (0.654) |
| <i>Tiliacea aurago</i> | Xyleninae | egg | resident | euryoecious | specialist | woody | 0.335 (0.335) | 1.096 (0.004) | 0.599 (0.476) | 0.761 (0.145) |
| <i>Xestia baja</i> | Noctuinae | larval | resident | stenoecious | generalist | herbaceous | -0.702 (-0.702) | 0.101 (0.621) | -0.297 (0.704) | 0.803 (0.11) |
| <i>Xestia c-nigrum</i> | Noctuinae | larval | migratory | euryoecious | generalist | herbaceous | -0.548 (-0.548) | 1.789 (0.079) | 0.212 (0.901) | 2.338 (0.028) |
| <i>Xestia xanthographa</i> | Noctuinae | larval | resident | euryoecious | generalist | herbaceous | 0.59 (0.59) | -0.375 (0.093) | -0.004 (0.995) | -0.965 (0.042) |

Appendix 3. Phylogenetic tree of noctuid taxa used in PGLS models



Appendix 4. References for creating the phylogenetic tree of noctuid taxa.

Mitchell, A. et al. (2006) Systematics and evolution of the cutworm moths (Lepidoptera : Noctuidae): evidence from two protein-coding nuclear genes. Syst. Ent. 31, 21-46.

Speidel, W. et al. (1996) The phylogeny of the Noctuidae (Lepidoptera). Syst. Ent. 21, 219-251.

Zahiri, R. et al. (2011) A new molecular phylogeny offers hope for a stable family level classification of the Noctuoidea (Lepidoptera). Zool. Script. 40, 158-173.

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|-----------|---------------|----------------|-----------------|-----------------|-----------------|------------------|-----------------|
| | $\beta(SE_u)$ | 0.347 (0.34) | -0.547 (0.243) | 0.252 (0.15) | 0.224 (0.165) | -0.395 (0.145) | 0.386 (0.303) |
| | 95% CI | (-0.32; 1.015) | (-1.023; -0.07) | (-0.042; 0.546) | (-0.099; 0.548) | (-0.678; -0.111) | (-0.208; 0.981) |
| LED slope | 0 | 2.66 (0.438) | | | | | -2.671 (0.536) |
| | 1 | 2.867 (0.593) | -0.206 (0.398) | | | | -2.671 (0.539) |
| | 2 | 2.66 (0.439) | | | | -0.212 (0.296) | -2.629 (0.541) |
| | 3 | 2.66 (0.44) | | | 0.149 (0.287) | | -2.731 (0.551) |
| | 4 | 2.675 (0.482) | | -0.018 (0.242) | | | -2.675 (0.542) |
| | 5 | 2.929 (0.6) | -0.268 (0.406) | | | -0.249 (0.303) | -2.621 (0.544) |
| | 6 | 2.912 (0.601) | -0.252 (0.407) | | 0.181 (0.293) | | -2.744 (0.554) |
| | 7 | 2.861 (0.599) | -0.232 (0.439) | 0.039 (0.266) | | | -2.663 (0.546) |
| | 8 | 2.66 (0.442) | | | 0.099 (0.3) | -0.183 (0.311) | -2.674 (0.562) |
| | 9 | 2.699 (0.484) | | -0.048 (0.246) | | -0.221 (0.303) | -2.636 (0.547) |
| | 10 | 2.714 (0.489) | | -0.067 (0.258) | 0.175 (0.306) | | -2.755 (0.563) |
| | | | | | | | |
| | $\beta(SE_u)$ | 2.718 (0.496) | -0.227 (0.406) | -0.018 (0.252) | 0.151 (0.293) | -0.217 (0.301) | -2.674 (0.543) |
| | 95% CI | (1.746; 3.691) | (-1.023; 0.57) | (-0.512; 0.475) | (-0.424; 0.725) | (-0.807; 0.373) | (-3.737; -1.61) |
| MED slope | 0 | 0.282 (0.512) | -0.358 (0.54) | | | | |
| | 1 | 0.195 (0.27) | | | | -0.366 (0.336) | |
| | 2 | 0.731 (0.617) | -0.536 (0.555) | | | -0.449 (0.347) | |
| | 3 | 0.225 (0.608) | | | | | 0.076 (0.745) |
| | 4 | -0.058 (0.185) | | | 0.081 (0.387) | | |
| | 5 | -0.137 (0.257) | | 0.161 (0.331) | | | |
| | 6 | 0.282 (0.514) | -0.559 (0.593) | 0.301 (0.363) | | | |
| | 7 | 0.693 (0.822) | -0.468 (0.551) | | | | 0.076 (0.746) |

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|----------|---------------|-----------------|-----------------|-----------------|----------------|-----------------|
| | 8 | 0.282 (0.516) | -0.391 (0.553) | 0.131 (0.395) | | |
| | 9 | 0.201 (0.307) | | -0.018 (0.398) | -0.369 (0.348) | |
| | 10 | 0.123 (0.364) | | 0.1 (0.337) | -0.347 (0.344) | |
| | 11 | 0.708 (0.62) | -0.701 (0.602) | 0.261 (0.363) | -0.426 (0.35) | |
| | 12 | 0.225 (0.611) | | | -0.26 (0.412) | 0.128 (0.753) |
| | 13 | 0.725 (0.626) | -0.542 (0.564) | 0.033 (0.401) | -0.443 (0.357) | |
| | 14 | 0.225 (0.612) | | 0.056 (0.399) | | 0.054 (0.767) |
| | 15 | -0.139 (0.26) | | 0.155 (0.35) | 0.026 (0.409) | |
| | 16 | 0.111 (0.668) | | 0.142 (0.336) | | 0.105 (0.752) |
| | 17 | 0.777 (0.83) | -0.552 (0.563) | | -0.336 (0.42) | 0.143 (0.753) |
| | 18 | 0.652 (0.825) | -0.673 (0.604) | 0.308 (0.367) | | 0.138 (0.751) |
| | 19 | 0.282 (0.517) | -0.563 (0.599) | 0.289 (0.379) | 0.051 (0.41) | |
| | 20 | 0.723 (0.834) | -0.498 (0.564) | | 0.121 (0.406) | 0.028 (0.769) |
| | 21 | 0.134 (0.375) | | 0.112 (0.353) | -0.054 (0.416) | -0.356 (0.353) |
| | 22 | 0.714 (0.628) | -0.699 (0.607) | 0.269 (0.378) | -0.04 (0.416) | -0.432 (0.359) |
| | 23 | 0.225 (0.615) | | | -0.015 (0.418) | -0.264 (0.433) |
| | 24 | 0.137 (0.673) | | 0.111 (0.342) | -0.237 (0.421) | 0.146 (0.76) |
| | 25 | 0.732 (0.835) | -0.731 (0.612) | 0.281 (0.37) | -0.304 (0.423) | 0.193 (0.758) |
| | 26 | 0.112 (0.68) | | 0.142 (0.359) | 0.001 (0.425) | 0.104 (0.782) |
| | 27 | 0.785 (0.84) | -0.559 (0.572) | | 0.041 (0.422) | -0.325 (0.437) |
| | 28 | 0.66 (0.84) | -0.675 (0.61) | 0.301 (0.386) | 0.026 (0.425) | 0.126 (0.781) |
| | | | | | | |
| | $\beta(SE_u)$ | 0.298 (0.598) | -0.497 (0.575) | 0.206 (0.36) | 0.052 (0.404) | -0.376 (0.365) |
| | 95% CI | (-0.874; 1.469) | (-1.625; 0.631) | (-0.499; 0.912) | (-0.74; 0.843) | (-1.091; 0.339) |
| | | | | | | |
| Slope of | 0 | 2.795 (0.49) | | | | -3.01 (0.601) |

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|-----------------|---------------|----------------|----------------|----------------|-----------------|-----------------|
| activity length | | | | | | |
| | 1 | 2.585 (0.665) | 0.21 (0.446) | | | -3.01 (0.604) |
| | 2 | 2.795 (0.494) | | | 0.099 (0.333) | -3.03 (0.608) |
| | 3 | 2.94 (0.538) | | -0.182 (0.27) | | -3.046 (0.605) |
| | 4 | 2.795 (0.494) | | | -0.07 (0.322) | -2.982 (0.618) |
| | 5 | 2.622 (0.667) | 0.396 (0.488) | -0.279 (0.296) | | -3.066 (0.607) |
| | 6 | 2.552 (0.675) | 0.243 (0.457) | | 0.132 (0.341) | -3.036 (0.612) |
| | 7 | 2.56 (0.675) | 0.235 (0.457) | | -0.1 (0.329) | -2.97 (0.622) |
| | 8 | 2.795 (0.497) | | | -0.047 (0.338) | 0.085 (0.35) |
| | 9 | 2.934 (0.543) | | -0.173 (0.276) | 0.064 (0.339) | -3.057 (0.613) |
| | 10 | 2.941 (0.547) | | -0.182 (0.289) | 0.001 (0.342) | -3.047 (0.63) |
| | 11 | 2.596 (0.678) | 0.415 (0.496) | -0.27 (0.3) | | 0.102 (0.343) |
| | 12 | 2.619 (0.679) | 0.397 (0.493) | -0.275 (0.312) | -0.013 (0.344) | -3.06 (0.632) |
| | | | | | | |
| | $\beta(SE_u)$ | 2.757 (0.567) | 0.265 (0.467) | -0.212 (0.287) | -0.06 (0.331) | 0.099 (0.339) |
| | 95% CI | (1.646; 3.867) | (-0.651; 1.18) | (-0.774; 0.35) | (-0.708; 0.588) | (-0.564; 0.763) |
| | | | | | | (-4.21; -1.826) |