1	Csilla Kovács - Péter Balling - Zoltán Bihari - Antal Nagy – Erzsébet Sándor
2	
3	Incidence of grapevine trunk diseases is influenced by soil, topology and vineyard age, but not by Diplodia
4	seriata infection rate in the Tokaj Wine Region, Hungary
5	
6	
7	1. Csilla Kovács
8	University of Debrecen, Faculty of Agricultural and Food Sciences and Environmental Management, Institute of
9	Food Science
10	Böszörményi út 138., H-4032 Debrecen, Hungary
11	<u>k.csilla20@gmail.com</u> , +3652508444/88598
12	
13	2. Péter Balling
14	Research Institute for Viticulture and Oenology
15	Könyves Kálmán u. 54., H-3915 Tarcal, Hungary
16	balling.peter@tarcalkutato.hu, +3647380148
17	
18	3. Zoltán Bihari
19	Research Institute for Viticulture and Oenology
20	Könyves Kálmán u. 54., H-3915 Tarcal, Hungary
21	<u>biharitokaj@gmail.com</u> , +3647380148
22	
23	4. Antal Nagy
24	University of Debrecen, Faculty of Agricultural and Food Sciences and Environmental Management, Institute of
25	Plant Protection
26	Böszörményi út 138., H-4032 Debrecen, Hungary
27	<u>nagyanti76@gmail.com</u> , +3652508444/88128
28	
29	5. Erzsébet Sándor (corresponding author)
30	University of Debrecen, Faculty of Agricultural and Food Sciences and Environmental Management, Institute of
31	Food Science
32	Böszörményi út 138., H-4032 Debrecen, Hungary
33	<u>karaffaem@yahoo.co.uk</u> , +3652508444/88067

34 Abstract

35 Grapevine Trunk Diseases (GTD) are of great importance worldwide, including Hungary, a Center European 36 country with long wine producing history. Several GTD pathogens have been described till now in Europe, but 37 only a few from Hungary. The presence of a GTD pathogen in the vine does not necessarily result in the immediate 38 appearance of disease symptoms, and information on the importance of environmental factors related to disease 39 incidence are still limited. The aim of this research was to assess the occurrence of GTD in the Tokaj Wine Region, 40 and to determine the biotic and abiotic factors influencing disease incidence. Five vineyards within 15 km radius 41 - each with different topology, soil types, varieties and age - were studied for three consecutive years (2013 -42 2015). The incidence of GTD-infection was determined every year for each vineyard. Diplodia seriata was isolated 43 with incidence ranging from 50 to 100 %, while *Diaporthe* spp. were the only other – minor – GTD pathogen 44 found. Topology and soil type appeared to be major abiotic factors affecting incidence of GTD symptom. Disease 45 incidence was also positively correlating with the age of the vineyards, and it was in fact found to be the definitive 46 biotic factor regarding incidence. In contrast, D. seriata infection rate appeared unrelated to disease incidence or 47 to any of the biotic or abiotic factors investigated.

Key words: disease incidence, Furmint, Hárslevelű, Diplodia seriata

51 Introduction

48 49

50

Grapevine trunk diseases (GTDs) restrict the productivity and longevity of vineyards, causing significant economic
losses (Scheck et al. 1998; Siebert 2001). Eutypa, Botryosphaeria, Phomopsis dieback, Esca disease complex, and
Petri disease are considered the major GTDs worldwide, and the causal pathogens attack the woody perennial
organs of the vine and ultimately lead to the death of the plant (Lehoczky 1974; Larignon and Dubos 1997;
Rolshausen et al. 2004; Úrbez-Torres et al. 2006; Kotze et al. 2011; Úrbez-Torres 2011; Bertsch et al. 2012; ÚrbezTorres et al. 2013; Úrbez-Torres et al. 2014; Fontaine et al. 2015).

- Grapes are the largest fruit crop in Hungary, with high economic value (Hungarian Central Statistical Office,
 2014). Moreover there is also great cultural importance in the historic Tokaj Wine Region, which is listed on the
 UNESCO World Heritage since 2002 as a producer of the world's oldest botrytized "aszú" wines. The actual size
 of cultivated areas in the Tokaj Wine Region is 5050 hectares, growing almost inclusively indigenous varieties
 (Furmint, Hárslevelű, Sárgamuskotály (yellow muscat), Zéta, Kövérszőlő, Kabar) (Hungarian Central Statistical
 Office, 2014). GTDs have significant effects on the profitability of grape production in the Tokaj Wine Region of
 Hungary, causing almost 110000 USD (32 million HUF) in losses annually (Bihari et al. 2015).
- 65 Hewitt et al. (1957) observed that specific symptoms cannot be detected on the diseased trunks every year. Latest 66 results have also proved that abiotic factors affect the appearance and the severity of the disease (Lecomte et al. 67 2011). GTDs presents in different forms: (i) slow but chronic disease development over many years (grapevine 68 leaf stripe disease, young esca or 'Phaeotracheomycosis of grapevine'), or (ii) rapid, acute disease development 69 (apoplexy, esca proper) within one season (Surico et al. 2006). GTDs are caused by many different Ascomycete 70 fungi, including Diatrypaceaous and Botryosphaeriaceous species, Phomopsis species, Phaeomoniella and 71 Phaeoacremonium species, as well as Basidiomycetous fungi such as Fomitiporia mediterranea (Kuntzmann et 72 al. 2010; Kotze et al. 2011; Bertsch et al. 2012; Úrbez-Torres et al. 2014). GTD incidence has increased during 73 the last few decades (Úrbez-Torres et al. 2014). Esca incidence has reached 60% to 80% in some old vineyards in

- southern Italy (Pollastro et al. 2000; Sidoti et al. 2000; Surico et al. 2000; Romanazzi et al. 2006; Calzarano and
 Di Marco 2007). GTD incidence may vary between closely located vineyards possibly due to microclimate, soil
 composition or water supply, suggesting abiotic environmental factors are likely to have a role in the development
 of the disease (Lecomte et al.; 2011; Bertsch et al. 2012).
- GTDs have been reported previously in Hungary. Lehoczky (1974) described the black dead arm disease of
 grapevine, which is caused by *Botryosphaeria stevensii* (R. A. Schoemaker), in the Tokaj Wine Region. Later *Phaeoacremonium hungarium* (S. Essakhi, L. Mugnai, G. Surico & P. W. Crous (Pers.)) was identified as a new *Phaeoacremonium* species in the Tokaj Wine Region (Essakhi et al. 2008). The disease incidence of the esca was
 reported to increase from 2% to almost 13%, between 2003 and 2007 in Hungary (Dula 2011). However, there is
 limited information about the current extent and distribution of GTD in Hungary, especially in the Tokaj Wine
 Region.
- 85 The effective control of GTD faces several problems. Although the susceptibility of the different cultivars are 86 different (Marchi 2011; Romanazzi et al. 2009; Andreini et al. 2014; Murolo and Romanazzi 2014) and the 87 rootstock may also affect the frequency of the symptoms (Marchi 2011; Murolo and Romanazzi 2014) there is no 88 grapevine cultivar known to be resistant to GTD. Once GTD pathogens have entered the woody tissues, they 89 proliferating inside the vine, where fungicides may have difficulty reaching them. Sodium arsenite, the previously 90 used most effective chemical (Mugnai et al. 1999), was banned because of its human carcinogenic, and 91 environmental toxic properties (Kuntzmann et al. 2010). Prevention of planting material (Gramaje et al. 2015) and 92 appropriate protection of pruning wounds (Eskalen et al. 2007; Sosnowski et al. 2008; Rolshausen et al. 2010; 93 Kotze et al. 2011; Sosnowski et al. 2013) are the most important techniques to prevent disease. The appropriate 94 time for pruning by avoiding spore dispersal periods (Petzoldt et al. 1983; Rooney-Latham et al. 2005; Eskalen et 95 al. 2007; Úrbez-Torres and Gubler 2008; Rolshausen et al. 2010; Fontaine et al. 2015) and pathogen free planting 96 materials (Billones-Baaijens et al. 2015) may also prevent infection and decrease disease incidence.
- 97 The aims of this research were (i) to investigate the incidence of GTD in different vineyards of the Tokaj Wine
 98 Region, Hungary; and (ii) to identify possible biotic (age, variety, endophytic fungi) and abiotic (plantation
 99 characteristics, soil, weather conditions, vintage) factors affecting disease incidence.

101 Material and Methods

100

102 Vineyard characteristics and meteorological data

- The survey was carried out in five vineyards (Bakonyi, Dorgó, Szemere, Szarvas and Várhegy) in the Tokaj Wine
 Region during 2013 and 2015, located close to each other, within 15 km (Fig. 1).
- 105 The Bakonyi and Szarvas vineyards were planted on cambisols, while the Szemere, Dorgó, and Várhegy vineyards 106 were planted on slope sediment from luvisols soil. All training systems were mid-high cordon, except for Szemere, 107 which was Guyot (Table 1.). The rootstock 125AA was used only in Várhegy for the variety Zéta (Vitis vinifera), 108 and Teleki 5C was used in all the other vineyards with varieties Furmint and Hárslevelű (V. vinifera). There was 109 also difference on the direction of the rows on the slopes, as the Bakonyi vineyard is teracced, while the other 110 vineyards are on steep slopes. All vineyards had only one variety, except Bakonyi which had both Furmint (4F), 111 and Hárslevelű (4H). Data from Szarvas vineyard was also divided into two parts: the upper part of the vineyard 112 with 5 - 8% slope (5S), and a lower, flat part at the foot of the hill (5L), thus data of seven sites were draw into 113 the analysis (Table 1.).

Meteorological data were collected with MILLIMET Weather Station (Boreas Kft., Hungary) in the region.
Minimum, maximum and average temperature, as well as total amount of precipitation was calculated from the
data for each month.

117

130

141

118 Disease incidence (DI) and collection of samples

119 The evaluations of external GTD symptoms were carried out between May and August in each year from 2013-120 2015, with visual inspections in the vineyards. . The disease incidence (DI) was calculated as the ratio of the 121 number of plants expressing foliar symptoms with wood necrosis or dieback in each vineyard over the number of 122 all vines and multiplying by 100. Samples were collected from each vineyard at several sampling times each year 123 between May and September from the trunks with typical GTD symptoms. Only the upper parts of the plants 124 (cordon) were sampled to leave the plants alive for further analysis. Samples were taken from all symptomatic 125 plants for isolation. Moreover representative random sampling was performed to determine the infection rate of 126 D. seriata in each vineyard. A minimum of three samples were collecting from each row for this representative 127 sampling. We sampled at least 3% of vines in each vineyard except Bakonyi, where all plant were analyzed. 128 Woody samples were transported in a cooler (under 10°C) to the laboratory at the University of Debrecen within

48 hours to determine the colonizing fungal microflora of the wood tissues.

131 Isolation of fungi from woody tissues

132 We established the infection rate (IR) (percentage of infected vines in the sample) of *D. seriata* and *Diaporthe* sp. 133 to characterize and compare the infection level in vineyards on the basis of microbiological analysis made in 2015. 134 Wood chips from debarked and surface sterilized (10% Noemagnol solution than washing twice with sterile 135 distilled water) plant samples were cut from both browned and adjacent apparently healthy wood tissues, and 136 placed on malt extract agar medium (MEA, Scharlau, Spain) in 90 mm diameter Petri dishes with a sterile scalpel, 137 under aseptic conditions following the method of Abreo et al. (2013), with modification described in Kovács et al. 138 (2014). Plates were incubated at room temperature in darkness for 7 to 14 days, and mycelial fragments from 139 emerging fungal colonies were transferred to new 2% MEA plates (Crous et al. 2006). Isolates were maintained 140 on MEA. Mycelial or conidial suspensions were also stored in 50% glycerol at -80°C.

142 Morphological and molecular identification of the isolated fungi

- Pure fungal cultures were used for the taxonomic identification of the fungi based on morphological and cultural features (mycelial color, cell wall structure). Pycnidia formation structures (presence, absence) on potato-dextrose agar medium with streptomycine sulfate (PDA, Scharlau, Spain) and conidia (color, shape, size) or conidiophores were examined under microscope. The morphological identification of *Diplodia seriata* were based on Crous (2006), and *Phomopsis* species on Mostert et al. (2001) and Van Niekerk et al. (2005).
- Molecular identification with sequence analysis of the Internal Transcribed Spacer (ITS) region of the rDNA was
 performed for isolates whose sporulation was not observed. Randomly chosen isolates from each morphological
 group were also sequenced.
- Fungal DNA was isolated from fresh mycelium scraped at the surface of PDA described previously (Assadollahi
 et al. 2013). DNA concentrations were measured by NanoDrop 2000 (Thermo Scientific).

Amplifications of 25 µl PCR reaction containing 12.5 µl 2 X PCR Master Mix (GoTaq Green Master (Promega),
40-40 pmol of each primer, 20-40 ng of genomic DNA and nuclease free water were run. ITS4 and ITS5 primers
(White et al. 1990) (Integrated DNA Technologies, Inc.) were used to amplify the full length of ITS region, with
the following amplification protocol: 3 min initial denaturing at 95°C, followed by 5 cycles of 1 min at 95°C, 1
min annealing at 50°C, 1 min at 72°C and 25 cycles of 1 min at 90°C, 1 min annealing at 50°C, 1 min at 72°C and
15 min final extension at 72°C. Purified amplification products were sequenced by Microsynth Austria GmbH.

The sequences were manually aligned and compared with those deposited in the NCBI GenBank database using the BLAST program (Altschul et al. 1990). The ITS sequences were also submitted to the NCBI GenBank (www.ncbi.nlm.nih.gov) for the species identification and KU377167-KU377290 accession numbers were obtained.

164 Statistical analyses

163

165

182 183 184

185

Binominal tests were used to analyze deviation of the infection rate in the sites from the summarized proportion in the whole sample. The relationship between IR and DI was analyzed with correlation analysis. Beyond that the effect of location (seven studied site), age of plantations (below or above 15 years old in 2013), vintage (three studied years), topology (slope and horizontal), soil types (luvisols and cambisoils) and grapevine variety (Furmint and Hárslevelű) on the DI were analyzed. The analyses were made on the whole samples in each case except the Várhegy vineyard, where only Zéta variety is grown, so it was excluded from the comparison of different grapevine variety.

Three categories were used to characterize the temporal stability of symptom appearance between 2013 and 2015:
 occasional – symptoms appeared only one year, stable – symptoms appeared twice and continuous – symptoms
 appeared in all three studied years. The proportion of the three categories was calculated for each site and the effect
 of above mentioned biotic and abiotic parameters on it was analyzed.

The Kolmogorov-Smirnov test for normality and the Levene test were used to test normality and the equality of
variance assumptions of parametric tests. Considering that our data did not meet these assumptions in case of
multiple comparison of groups Kruskall-Wallis nonparametric test were used. Pairs showing significant
differences were compared by Mann-Whitney U-test. The paired comparisons were also made by Mann-Whitney
U-test (Reiczigel et al. 2007). Statistical analyses were performed by SPSS 21.0 (Ketskeméty et al. 2011).

Results

The percentage of DI of GTD varied from 0.17 to 42.11% in the three-year study according to observations made on 22794 grapevines (Table 2). Large areas of chlorosis and deterioration between the veins were observed on the leaves of the diseased plants together with small black spots on the cross section of the cordon. Mean DI showed significant differences (Kruskal-Wallis test: H=19.375 df=6, n=21, p=0.004) in the different vineyards. The highest mean DI was detected in the Bakonyi vineyard. Here both Furmint (4F) and Hárslevelű (4H) varieties showed significantly higher DI than other vineyards and there was also significant difference between them. An intermediate mean DI value was observed in the Szarvas vineyard with significant difference between higher (5S)

- and lower (5L) part of this site. The Dorgó vineyard also showed intermediate DI, while the Szemere and Várhegyvineyards had the lowest mean DI (Fig. 2.).
- 195The most frequently identified GTD pathogens from the upper part of the symptomatic grapevine woody tissues196was *D. seriata*, while *Diaporthe* sp. was identified with much lower rates from the 558 samples taken in 2015. The197mean infection rate (IR) of *D. seriata* was 76% with large variance among studied sites. Infection rate of each site198significantly differed from the mean value (Binomial test p<0.05). The mean IR of *Diaporthe* sp. was 4% without199significant deviation from this mean (Binomial test p>0.05) (Table 2). There was low correlation between IR and200DI ($r^2=0.4818$). The success of isolation was established in the ratio of infected plant to symptomatic plants (*D. seriata*: 75.51%, 406/558 plants; *Diaporthe* sp.: 3.71% 19/558 plants; see details in Table 3).
- D. seriata could be isolated from most of the symptomatic plants. Its isolation rate varied between 94 and 100%.
 Diaporthe species were isolated with much lower frequencies, while other *Botryosphaeria* sp. were detected only
 in one year (2013) from one vineyard (Bakonyi).
- Non-GTD pathogen, endophytic fungi (*Trichoderma* sp., *Alternaria* sp., *Mucor* sp., *Penicillium* sp, *Epicoccum* sp., *Fusarium* sp. and *Aspergillus* sp.) were also isolated with up to 100% isolation rates from symptomatic plants.
 Alternaria, and *Fusarium* species were the most frequently isolated non-GTD pathogens isolated with 43-100%, and 0-55% of the symptomatic plants.
- Among the studied abiotic and biotic factors topology, soil type and age of the vineyard showed significant effect on the disease incidence (DI) (Table 4). However variety and vintage (year) did not have a significant effect on the DI. There was no significant difference on the average DI of the vineyards at the different vintages (Kruskall-Wallis test: H=0.0967, df=2, n=21, p=0.953). DI was the highest in 2014 in Szemere and Szarvas (sloped), but decreased during the three years in Dorgó, but did not change in Várhegy.
- In case of topology, significantly lower DI was detected in vineyards planted on slopes (Dorgó,Szemere, Várhegy), than in the terraced Bakonyi. This difference could be seen even within the same vineyard (Szemere) with different topology (5S and 5L, see above in the text and Fig. 2. and Table 1.). Plantations on luvisols soil also had significantly lower DI than on the cambisols with similar differences in mean values to that seen with topology (Table 4).
- Although in the Bakonyi vineyard there was a large difference in mean DI between two varieties Furmint (4F) and Hárslevelű (4H) (Fig 2). However DI of Furmint and Hárslevelű varieties was not statistically different across all vineyards with different age. There were no differences in the DI between years either. Because of the relatively short study period, the trends in temporal changes of DI could not been analysed but different tendencies could be observed in the different vineyards (Table 1. The Várhegy vineyard which was planted with the variety Zéta on rootstock 125AA, had the lowest disease incidence (0.17%) of all those examined (Table 1., Fig. 2.).
- The age of vineyard had an effect, with DI in the older vineyards (over 15 years) were significantly higher,comparing to the younger ones (under 15 years) (Table 4.).
- Neither studied abiotic or biotic factors, nor DI showed significant effect on *D. seriata* infection rate of the
 vineyards. Similarly no correlation was found between isolation rate of *D. seriata* from symptomatic plants and
 any of the studied factors.
- Only eight symptomatic plants could be detected in the Várhegy vineyard during the three years of the study, with
 one vine expressing symptoms over two years, but none with continuous disease incidence was observed (Table
 5). Therefore it was excluded from the statistical analysis of the symptom stability. The ratio of occasional disease

233

235

incidence was significantly (Mann-Whitney U test, p=0.0244) higher in younger vineyards (younger than 15 years 234 at 2013), than stable or continuous disease incidence (Figure 3.).

236 Discussion

237 Grapevine trunk diseases are the most threatening problem to wine industries with increasing disease incidence 238 worldwide (Bertsch et al. 2012; Úrbez-Torres et al. 2013). It is a complex disease with unique characteristics, 239 infection can be latent without visible disease symptoms for years (Lehoczky 1974; Cristinzio 1978; Phillips 2002; 240 Auger et al. 2004; Marchi et al. 2006; Van Niekerk et al. 2006; Savocchia et al. 2007; Rego et al. 2008), but the 241 factors affecting the disease appearance are still only suspected. The influence of the weather conditions has been 242 reported may affect symptom appearance. Increased disease incidence was reported in case of irrigation (Bertsch 243 et al. 2012) or high amount of precipitation (Lehoczky 1974; Hewitt 1988). Bruno et al. (2007); Marchi et al (2006) 244 and Andolfi et al. (2009) explained that increase with facilitated transport of the fungal toxins responsible for foliar 245 symptoms. Moreover Surico et al. (2006) and Bertsch et al. (2012) reported increased disease appearance when 246 drought weather was followed by rainy period. Sosnowski et al. (2007) reported that both temperature and rainfall 247 were related to eutypa dieback symptom development. Different biotic factors, like variety (Marchi 2011; Maher 248 et al. 2012; Murolo and Romanazzi 2014), rootstock type (Marchi 2011; Murolo and Romanazzi 2014) age of the 249 plant (Mugnai et al. 1999) was also indicated may affect the disease incidence.

- 250 The detection rate of the reported GTD pathogen D. seriata was high, but variable (50 - 100 %) in the vineyards 251 when both symptomatic and asymptomatic plants were included in the test. This infection rate was similar to 252 previously reported isolation rate of GTD pathogens, particularly D. seriata in other studies from symptomatic 253 vines (Bruez et al. 2014). It was concluded that infection rate did not affect symptom appearance, expressed as 254 disease incidence rate in this study. It must be mentioned, however that only the upper part was sampled for fungal 255 isolation, therefore GTD pathogens that may exist in the lower parts of the vine were not included.
- 256 There was significant difference in the disease incidence of GTD at the different vineyards. DI was much higher 257 in the Bakonyi vineyard, than in Dorgó vineyard with similar characteristics except soil type and topology. The 258 Bakonyi vineyard is terraced, whereas the other examined vineyards with significantly lower disease incidence 259 were planted with rows on the slopes. The difference of the DI could be detected even within the same vineyard 260 (Szemere) with different topology. The DI was significantly higher on horizontal (5L) part, than on 5-8% slope 261 (5S). Surico et al. (2000), and Robotic and Bosancik (2007) similarly found higher disease incidence in vineyards 262 on gentler slope, than on sleep ones.
- 263 Soil type was the other tested abiotic factor for GTD. Szemere, Dorgó, Várhegy vineyards were on luvisols, while 264 vineyards Bakonyi and Szarvas on cambisols. DI of the vineyards on cambisols was significantly higher, 265 comparing with the other soil type. Cambisol is considered as a soil with high water capacity (Rhoton and 266 Markewich 2006). Soil with high water reserve was reported to enhance esca symptoms (Guérin-Dubrana et al. 267 2005), and DI of GTD (Kovács et al. 2016).
- 268 Regarding the varieties, higher disease incidence was observed on Furmint, than on Hárslevelű at Bakonyi 269 vineyard in each year. However Furmint had lower disease incidence in the other two younger (age > 15 years) 270 vineyards (Szemere and Dorgó), than in Szarvas vineyard planted later (age < 15 years) with Hárslevelű variety 271 (Table 2, Fig. 2). Interestingly, Várhegy vineyard 3 had the lowest disease incidence (0.17%), among the examined 272 ones. It had unique characteristics regarding variety (Zéta) and rootstock (125AA), comparing to the others. It was

- among the youngest examined plantation (12 years old in 2013), and its disease incidence was lower, than Szemere
 vineyard with the same age.
- Although the weather conditions were different in the three years of the study, neither temperature nor amount of precipitation resulted differences of the DI The trends of the DI changes were different in vineyards on slope (1-3 and 5S), however vineyards 4F, 4H, and 5L, with horizontal topology showed an increasing DI between 2013 and 2015. There was significant difference in DI of 5L and 5S (different topology) at 2013, and 2015, while the difference in the GTD symptom expression has disappeared at 2014, when higher amount of precipitation was observed during the summer (green berries growth) period on the slopes as well (Table 2).
- The majority of vines expressed disease symptoms in only one of the three years. The average disease incidence rate, regarding the individual vineyards was also the highest (62.42%) for the plants expressing symptoms only once. Occasional disease incidence ranged above 50% (55.17-80%) in all vineyards, except the 4H (Bakonyi with Hárslevelű variety), where stable disease expression occurred at 68%. Interestingly, not this vineyard had the highest DI value.
- 286 It was concluded, that topology and soil type, and were the most important abiotic factors for disease incidence 287 (DI) of the GTD symptom appearance expressed by foliar symptoms or dieback. The higher DI may have resulted 288 by increased pathogen activity (growth or toxin production), or facilitated toxin transport (Bruno et al 2007; Marchi 289 et al. 2006). Among the biotic factors, age of the vineyards had the highest detectable impact for the DI, resulting 290 a significantly higher rate of diseased plants in the vineyards with older plants. It also may have caused by several 291 factors, e.g. cumulated infection of the different GTD pathogens, moreover the change of the plant physiology and 292 plant resistance, or the endophytic microbiota. Further studies are necessary to explain the role of the soil type, 293 topology in the increase of the diseased plants in the vineyards.

295 Acknowledgements

294

303 304

The authors would like to thank Mark Sosnowski the critical review of the manuscript. This work was supported by the TÁMOP 4.2.4.A/2-11-1-2012-0001 project (Nemzeti Kiválóság Program Hazai hallgatói, illetve kutatói személyi támogatást biztosító rendszer kidolgozása és működtetése konvergencia program). The project is cofinanced by the European Union and the European Social Fund. E. Sándor was supported by the Research Grant of the University of Debrecen. The research was supported by through the New National Excellence Program of the Ministry of Human Capacities, Hungary, and COST Action FA1303. The research also has been undertaken with the support of the OTKA No. K1325.

References

- Abreo, E., Martinez, S., Bettucci, L., & Lupo, L. (2013). Characterization of *Botryosphaeriaceae* species
 associated with grapevines in Uruguay. *Australasian Plant Pathology*, *42*, 241–249.
- Altschul, S. F., Gish, W., Miller, W., Myers, E. W., & Lipman, D. J. (1990). Basic local alignment search tool. *Journal of Molecular Biology*, *215*, 403–410.
- 309 Amponsah, N. T., Jones, E. E., Ridgway, H. J, & Jaspers, M. V. (2011). Identification, potential inoculum sources
- and pathogenicity of botryosphaeriaceous species associated with grapevine dieback disease in New Zealand.
- 311 *European Journal of Plant Pathology*, 131. 467–482. /DOI 10.1007/s10658-011-9823-1/

- Asadollahi M., Fekete, E., Karaffa L., Flipphi M., Árnyasi M., Esmaeili, M., Váczy K.Z., & Sándor E. (2013).
 Comparison of *Botrytis cinerea* populations isolated from two open-field cultivated host plants. *Microbiological Research*, *168*, 379-388.
- Bertsch, C., Ramírez-Suero, M., Magnin-Robert, M., Larignon, P., Chong, J., Abou-Mansour, E., Spagnolo, A.,
 Clément, C. & F. Fontaine. (2013). Grapevine trunk diseases: complex and still poorly understood. *Plant Pathology*, 62(2), 243–265.
- 318 Bihari, Z., Zsigrai, Gy., Éles, S. & Balling, P. (2015). Impact of the Vine Training System on the Occurrence of
- 319 Grapevine Trunk Diseases. Workshop, Cognac, Sustainable control of GTDs: current state and future prospects.
- 320 Abstracts of oral and poster presentation given at the first COST Action FA 1303 workshop on Grapevine Trunk
- 321 Diseases, Cognac, France, 23-24 June 2015., *Phytopathologia Mediterranea*, 54(2) 433. ISSN (print): 0031-9465,
- 322 ISSN (online): 1593-2095 /DOI: 10.14601/Phytopathol_Mediterr-16495/
- Billones-Baaijens, R., Jaspers, M. V.,, Allardi, A., Hong, Y., Ridgway, H., J., & Jones, E. (2015). Management of
 Botryosphaeriaceae species infection in grapevine propagation materials. *Phytopathologia Mediterranea*, 54(2)
 355–367. /DOI: 10.14601/Phytopathol_Mediterr-16159/
- Bruno, G. (2007). Effects of three esca-associated fungi on *Vitis vinifera* L.: V. Changes in the chemical and
 biological profile of xylem sap from diseased cv. Sangiovese vines Physiological and *Molecular Plant Pathology*,
 71, 210–229.
- Calzarano, F., & Di Marco, S. (2007). Wood discoloration and decay in grapevines with esca proper and their
 relationship with foliar symptoms. *Phytopathologia Mediterranea*, 46, 96–101.
- Crous, P. W., Slippers B., Wingfield M. J., Rheeder J., Marasas, W. F. O., Philips, A. J. L., Alves, A., Burgess T.,
 Barber, P., & Groenewald, J. Z. (2006). Phylogenetic lineages in the Botryosphaeriaceae. *Studies in Mycology*,
 55, 235–253. 2006.
- 334 Dula, T. (2011). Propagation material borne fungus pathogens causing early stock decay in vineyards.
 335 Növénvvédelem, 47, 461–468.
- Eskalen, A., Feliciano, A.J. & Gubler, W.D. (2007). Susceptibility of grapevine pruning wounds and symptom
 development in response to infection by *Phaeoacremonium aleophilum* and *Phaeomoniella chlamydospora*. *Plant Disease*, 91, 1100–1104.
- Fontaine, F., Pinto, C., Vallet, J., Clément, C., Gomes, A.C., & Spagnolo, A. (2015). The effects of grapevine
- trunk diseases (GTDs) on vine physiology. *European Journal of Plant Pathology*, 1-15. / DOI 10.1007/s10658015-0770-0/
- Gramaje, D., Mostert, L.,& Groenewald, J. Z. (2015). *Phaeoacremonium*: From esca disease to
 phaeohyphomycosis. Fungal Biology, 119. 759–783.
- 344 Guérin-Dubrana, L., Destrac-Irvine, A., Goutouly, J. P., Letouze, A., & Gaudillére, J. P. (2005).
- Relationship between incidence of esca and black dead arm foliar symptom expression in the vineyard, ecophysiological indicators and cultural practices. *Phytopathologia Mediterranea*,44, 110. (abstract)
- 347 Hewitt, W. B. (1957). Some manifestations of black measles of grapevines. *Phytopathology*. 47. 16. (abstract)
- Hewitt, W. B., & Pearson, R. C. (1988). *Diplodia* Cane Dieback and Bunch Rot. In R. C. Pearson, R. C. & A. C.
- 349 Goheen, A. C. (ed.) Compendium of Grape Diseases. (pp 25-26) American Phytopathological Society Press. St.
- 350 Paul, USA.

- 351 Higgins, D. G., & Sharp, P. M. (1988). "CLUSTAL: A package for performing multiple sequence alignment on a
- **352** microcomputer". *Gene*, *73*(1), 237–244.
- 353 Hungarian Central Statistical Office (2014).
- Ketskeméty, L., Izsó L., & Könyves Tóth E. (2011). Bevezetés az IBM SPSS Statistics programmendszerbe. Artéria
 Stúdió Kft, Budapest. 1-576.
- Kotze, C., Van Niekerk, J., Mostert, L., Halleen, F., & Fourie, P. (2011). Evaluation of biocontrol agents for
 grapevine pruning wound protection against trunk pathogen infection. *Phytopathologia Mediterranea*, 50, S247–
 S263.
- Kovács, Cs., Csótó, A., Rakonczás, N., & Sándor, E. (2016). Mikroklimatikus viszonyok szerepe a szőlő
 tőkebetegségeinek tünet megjelenésére. *Georgikon Napok Conference publication*, In Press.
- Kovács, Cs., Peles, F., Xie, H., Szojka, A., Hajdu, G., Bihari, Z., & Sándor, E, (2014). Isolation and identification
 of endophytic fungi connected to Grapevine Diseases, from the Tokaj wine region, Hungary. *Acta Agraria Debreceniensis*, 56, 61–66.
- Kuntzmann, P., Villaume, S., Larignon, P., & Bertsch, C. (2010). Esca, BDA and Eutypiosis: foliar symptoms,
 trunk lesions and fungi observed in diseased vine stocks in two vineyards in Alsace. *Vitis*, 49(2) 71–76.
- Larignon, P., & Dubos, B. (1997). Fungi associated with esca disease in grapevine. *European Journal of Plant Pathology*, *103*, 147–157.
- Larkin, M. A., Blackshields, G., Brown, N. P., Chenna, R., Mc Gettigan, P. A., Mc William, H., Valentin, F.,
 Wallace, I. M., Wilm, A., Lopez, R., Thompson, J. D., Gibson, T. J., & Higgins, D. G. (2007). ClustalW and
 ClustalX version 2.0. *Bioinformatics Advance Access*, 1-2.
- 371 Lecomte, P., Darrieutort, G., Laveau, C., Blancard, D., Louvet, G., Goutouly, J. P., Rey, P., & Guérin-Dubrana,
- L. (2011). Impact of biotic and abiotic factors on the development of Esca decline. *Integrated protection and production in viticulture*, */IOBC/ wprs Bulletin*, 67, 171–180.
- Lehoczky, J. (1974). Black dead arm disease of grapevine caused by *Botryosphaeria stevensii* infection. *Acta Phytopathologica Academie Scientiarum Hungaricae*, 9, 319–327.
- Marchi, G. (2001). Susceptibility to esca of various grapevine (*Vitis vinifera*) cultivars grafted on different
 rootstocks in a vineyard in the province of Siena (Italy). *Phytopathologia Mediterranea* 40, 27–36.
- Marchi, G., Peduto, F., Mugnai L., Di Marco, S., Calzarano, F., & Surico, G. (2006). Some observations on the
 relationship of manifest and hidden esca to rainfall. *Phytopathologia Mediterranea*, 45, 117–126.
- Mostert, L, Crous, P. W., Kang, J. C., & Phillips, A. J. L. (2001). Species of *Phomopsis* and a *Libertella* sp.
 occurring on grapevines with specifi c reference to South Africa: morphological, cultural, molecular and
 pathological characterization. *Mycologia*, 93, 146–167.
- 383 Mugnai, L., A., Graniti, A., & Surico G. (1999). Esca (black measels) and brown wood-streaking: two old and
 384 elusive diseases of grapevines. *Plant Disease*, 83, 404–416.
- 385 Murolo, S., & Romanazzi, G. (2014). Effect of grapevine cultivar, rootstock and clone on esca disease. *Australian* 386 *Plant Pathology*, 43, 215–221.
- Petzoldt, C. H., Sall, M. A., & Moller, W. J. (1983). Factors determining the relative number of ascospores released
 by *Eutypa armeniacae* in California. *Plant Disease*, 67, 857–860.
 - 10

- Pollastro, S., Dongiovanni, C., Abbatecola, A., & Faretra, F. (2000). Observations on the fungi associated with
 esca and on spatial distribution of esca symptomatic plants in Apulian (Italy) vineyards. *Phytopathologia Mediterranea*, 39, 206–210.
- Rego, C, Vaz, A, Nascimento, T, Cabral, A, Oliveira, H. (2008). Diseases incited *Botryosphaeriaceae* fungi in
 Portuguese vineyards. *Phytopathologia Mediterranea*, 48, 181.
- Reiczigel, J., Harnos A., & Solymosi, N. (2007). Biostatisztika nem statisztikusoknak. Pars Kft. Nagykovácsi,
 Hungary. 1–455.
- Robotic, V., & Bosancic, R. (2007). Notes on the relationship of manifest Esca disease to vineyard slope. *Phytopathologia Mediterranea*, 46(1), 124.
- Romanazzi, G., Murolo, S., Pizzichini, L., & Nardi, S. (2006). Grapevine esca disease in Marche region: first
 results. *Proceedings of the symposium "Giornate Fitopatologiche"*, 2, 289–290.
- 400 Romanazzi, G., Murolo, S., Pizzichini, L., & Nardi, S. (2009). Esca in young and mature vineyards, and molecular
 401 diagnosis of the associated fungi. *European Journal of Plant Pathology*, *125*(2), 277–290.
- 402 Rolshausen, P. E., Trouillas, F. P., & Gubler, W.D. (2004). Identification of Eutypa lata by PCR-RFLP. *Plant*403 *Disease*, 88, 925–929.
- Rolshausen, P. E., Úrbez-Torres, J. R., Rooney-Latham, S., Eskalen, A., Smith, R. J., & Gubler W., D. (2010).
 Evaluation of Pruning Wound Susceptibility and Protection against fungi Associated with Grapevine Trunk
 Diseases. *American Journal of Enology and Viticulture*, 61(1), 113–119.
- 407 Rooney-Latham, S., Eskalen, A., & Gubler, W. D. (2005). Occurrence of *Togninia minima* perithecia in esca408 affected vineyards in California. *Plant Disease*, *89*, 867–871.
- Scheck, H. J., Vasquez, S. J., Fogle, D., & Gubler, W. D. (1998). Grape growers report losses to black-foot and
 grapevine decline. *California Agriculture*, *52*, 19–23.
- Schoch, C. L., Seifert, K. A., Huhndorf, S., Robert, V., Spouge, J. L., Levesque, C. A., Chen, W., & Fungal
 Barcoding Consortium. (2012). Nuclear ribosomal internal transcribed spacer (ITS) region as an universal DNA
- 413 barcode marker for Fungi. *Proceedings of the National Academy of Sciences*, 109, 6241–6246.
- Sidoti, A., Buonocore, E., Serges, T., & Mugnai, L. (2000). Decline of young grapevines associated with *Phaeoacremonium chlamydosporum* in Sicily (Italy). *Phytopathologia Mediterranea*, *39*, 87–91.
- 416 Siebert, J. B. (2001). Eutypa: The economic toll on vineyards. *Wines and Vines, April*, 50–56.
- Sosnowski, M. R., Creaser, M. L., Wicks, T. J., Lardner, R., & Scott, E. S. (2008). Protecting grapevine wounds
 from infection by *Eutypa lata*. *Australian Journal of Grape and Wine Research*, *14*, 134–142.
- 419 Sosnowski, M. R., Loschiavo, A. P., Wicks, T. J., & Scott, E. S. (2013). Evaluating treatments and spray
- 420 application for the protection of grapevine pruning wounds from infection by *Eutypa lata*. *Plant Disease*, 97,
 421 1599–1604.
- Sosnowski, M. R., Shtienberg, D., Creaser, M. L., Wicks, T. J., Lardner, R., & Scott, E. S. (2007). The influence
 of climate on foliar symptoms of eutypa dieback in grapevines. *Phytopathology*, *97*, 1284–1289.
- 424 Surico, G., Marchi, G., Braccini, P., & Mugnai, L. (2000). Epidemiology of esca in some vineyards in Tuscany
 425 (Italy). *Phytopathologia Mediterranea*, *39*, 190–205.
- 426 Tamura, K., Peterson, D., Peterson, N., Stecher, G., Nei, M., & Kumar, S. (2011). MEGA5: molecular evolutionary
- 427 genetics analysis using maximum likelihood, evolutionary distance, and maximum parsimony methods. *Molecular*
- **428** *Biology and Evolution*, 28, 2731–2739.

- Thompson, J. D., Gibson, T. J., Plewniak, F., Jeanmougin, F., & Higgins, D. G. (1997). The CLUSTAL_X
 windows interface: flexible strategies for multiple sequence alignment aided by quality analysis tools. *Nucleic Acids Research*, 25, 4876–4882.
- 432 Úrbez-Torres, J. R., Leavitt, G. M., Voegel, T. M., & Gubler, W. D. (2006). Identification and distribution of
 433 *Botryosphaeria* spp. associated with grapevine cankers in California. *Plant Disease*, *90*, 1490–1503.
- 434 Úrbez-Torres, J. R. (2011). The status of *Botryosphaeriaceae* species infecting grapevines. Review.
 435 *Phytopatpology Mediterranea*, 50, 5–45.
- 436 Úrbez-Torres, J. R., & Gubler, W. D. (2008). Double pruning, a potential method to control Bot canker disease of
 437 grapes, and susceptibility of grapevine pruning wounds to infection by *Botryosphaeriaceae*. *Phytopathologia* 438 *Mediterranea*, 48, 185. (abstract)
- 439 Úrbez-Torres, J. R., Haag, P., Bowen, P., & O'Gorman, D. T. (2014). Grapevine trunk diseases in British
 440 Columbia: Incidence and characterization of the fungal pathogens associated with esca and Petri diseases of
 441 grapevine. *Plant Disease*, *98*, 469–482.
- 442 Úrbez-Torres, J. R., Peduto, F., Smith, R. J., & Gubler, W. D. (2013). Phomopsis dieback: A grapevine trunk
 443 disease caused by *Phomopsis viticola* in California. *Plant Disease*, 97. 1571-1579.
- Van Niekerk, J. M., Groenewald, J. Z., Farr, D. F., Fourie, P. H., Halleen, F., & Crous, P. W. (2005). Reassessment
 of *Phomopsis* species on grapevine. *Australasian Plant Pathology*, 34. 27–39.
- 446 White, T. J., Bruns, T. D., Lee, S. B., & Taylor, J. W. (1990). Amplification and direct sequencing of fungal
- 447 ribosomal RNA genes for phylogenetics. In M. A., Innis, D. H., Gelfand, J. J., Sninsky, & T. J., White (ed.) PCR
- 448 Protocols: a guide to methods and applications. (pp 317–322) Academic Press Inc., New York.

449

450	Legend for figures
451	
452	Fig 1. Sampling sites in the Tokaj Wine Region, Hungary in 2013-2015. 1. Szemere vineyard; 2. Dorgó
453	vineyard; 3. Várhegy vineyard; 4. Bakonyi vineyard; 5. Szarvas vineyard)
454	
455	Fig. 2 Mean disease incidence (DI% \pm SE/SD) at the different vineyards in the Tokaj Wine Region, Hungary
456	between 2013 and 2015. Letters indicate significant differences (p<0.05) according to Mann-Whitney U-test.
457	Number of sampling sites according to Fig 1.
458	
459	Fig. 3 Ratio of different disease stability categories (mean \pm SE/SD) in the studied sampling sites with different
460	age in the Tokaj Wine Region, Hungary, between 2013 and 2015.