

PROOF COVER SHEET

Author(s): Attila Mokánszki, Emese Varga Tóthné, Béla Bodnár, Zoltán Tándor, Zsuzsanna Molnár, Attila Jakab, Anikó Ujfalusi, and Éva Oláh

Article title: Is sperm hyaluronic acid binding ability predictive for clinical success of intracytoplasmic sperm injection: PICSU vs. ICSI?

Article no: UAAN_A_948102

Enclosures: 1) Query sheet
2) Article proofs

Dear Author,

Please check these proofs carefully. It is the responsibility of the corresponding author to check against the original manuscript and approve or amend these proofs. A second proof is not normally provided. Informa Healthcare cannot be held responsible for uncorrected errors, even if introduced during the composition process. The journal reserves the right to charge for excessive author alterations, or for changes requested after the proofing stage has concluded.

The following queries have arisen during the editing of your manuscript and are marked in the margins of the proofs. Unless advised otherwise, submit all corrections using the CATS online correction form. Once you have added all your corrections, please ensure you press the "Submit All Corrections" button.

Please review the table of contributors below and confirm that the first and last names are structured correctly and that the authors are listed in the correct order of contribution.

Contrib. No.	Prefix	Given name(s)	Surname	Suffix
1		Attila	Mokánszki	
2		Emese Varga	Tóthné	
3		Béla	Bodnár	
4		Zoltán	Tándor	
5		Zsuzsanna	Molnár	
6		Attila	Jakab	
7		Anikó	Ujfalusi	
8		Éva	Oláh	

AUTHOR QUERIES

Q1: Confirm: HBA: HA-binding assay, HA-binding ability

Q2: Note change to [Ménézo and...]

Q3: Confirm change to phrasing: The ICSI group included 42 couples where the sperm number and the HBA score was low and unsuitable for PICSU.

Q4: Confirm wording: ... the sample was mixed and a pipette of 7-10 μ l was placed near the center of the chamber.

Q5: Confirm phrasing: . . . embryos were put to . . .

Q6: Re: Eighteenth World Congress on Fertility and Sterility. . . Add location of meeting/add pp if Proceedings

Q7: Confirm page reference re WHO

Q8: Please provide last page range.

Q9: Please provide better quality artworks for all the figures.

6 RESEARCH ARTICLE

7
8
9 **Is sperm hyaluronic acid binding ability predictive for clinical success of**
 10 **intracytoplasmic sperm injection: PICSI vs. ICSI?**

11 Attila Mokánszki^{1*}, Emese Varga Tóthné², Béla Bodnár², Zoltán Tándor², Zsuzsanna Molnár¹, Attila Jakab³,
 12 Anikó Ujfalusi¹, and Éva Oláh⁴

13 ¹Department of Laboratory Medicine, ²Assisted Reproduction Center, Kaali Institute, ³Department of Obstetrics and Gynecology, and
 14 ⁴Clinical Genetic Center, Department of Pediatrics, Clinical Center, University of Debrecen, Debrecen, Hungary

15 **Abstract**

16 Although intracytoplasmic sperm injection (ICSI) is now a widely-used technique, it is still of
 17 interest to improve our knowledge as to which is the best spermatozoon to be selected for ICSI.
 18 Infertile men have increased risks of producing aneuploid spermatozoa. Using hyaluronic acid
 19 (HA)-binding sperm selection may reduce the genetic risks such as chromosomal aberrations of
 20 offspring. In the present study we examined the clinical success of ICSI with HA-selected sperm
 21 ('physiologic' ICSI, PICSI) compared to conventional ICSI, as well as the necessity to differentiate
 22 patients according to the initial HA-binding assay result (HBA score) and whether the sperm
 23 concentration or HBA score can provide additional information. We observed a significantly
 24 higher fertilization rate (FR) of the PICSI group with >60% HBA, implantation rate (IR) of the
 25 PICSI group with ≤60% HBA, and clinical pregnancy rate (CPR) in every PICSI group compared
 26 to the ICSI groups ($p < 0.01$). We also observed a significantly higher life birth rate (LBR) in the
 27 PICSI group with ≤60% HBA compared to ICSI patients with ≤60% HBA ($p < 0.001$). The
 28 pregnancy loss rate (PLR) was significantly lower in PICSI patients compared to the ICSI group
 29 ($p < 0.0001$). The FR, IR, CPR, and LBR of the PICSI group with <50% HBA were significantly
 30 higher and the PLR was lower than in the ICSI group with <50% HBA ($p < 0.01$). A statistically
 31 significant correlation was found between the sperm concentration and the HA-binding
 32 capacity ($r = 0.62$, $p < 0.001$). We found a closer relationship between HBA score and FR
 33 ($r = 0.53$, NS) than between sperm concentration and FR ($r = 0.14$, NS). HBA could be considered
 34 for sperm selection prior to ICSI because of its success and apparant ability to reduce genetic
 35 complications. However, this must be extended to a larger study.

36
 37 **Abbreviations:** CPR: clinical pregnancy rate; FR: fertilization rate; FSH: follicle stimulating
 38 hormone; GnRH: gonadotropin releasing hormone; HA: hyaluronic acid; HBA: HA-binding assay,
 39 HA-binding ability; hCG: human chorionic gonadotropin; HspA2: heat shock-related 70 kDa
 40 protein 2; ICSI: intracytoplasmic sperm injection; IR: implantation rate; IVF: *in vitro* fertilization;
 41 LBR: life birth rate; NS: non-significant; PICSI: ICSI with HA-selected sperm ('physiologic' ICSI);
 42 PLR: pregnancy loss rate; r: Pearson correlation coefficient; 2PN: two-pronuclear zygote, the
 43 appearance of two pronuclei is the first sign of successful fertilization

44
 45
 46 **Introduction**

47 Oligozoospermic men requiring intracytoplasmic sperm
 48 injection (ICSI) often carry sperm populations characterised
 49 by an increase in chromosomal aberrations and a compro-
 50 mised DNA integrity. A higher incidence of numerical
 51 [Palermo et al. 2000; Simpson and Lamb 2001; Van
 52 Steirteghem et al. 2002] and structural chromosomal aberrations
 53 [Bonduelle et al. 2002] have been associated in studies
 54 in the embryos resulting from ICSI.

55
 56
 57
 58 *Address correspondence to Attila Mokánszki, Department of
 59 Laboratory Medicine, Clinical Center, University of Debrecen,
 60 Nagyerdei krt. 98, Debrecen, H-4032, Hungary. Tel: +36-52-255-114.
 E-mail: mokanszki.attila@med.unideb.hu

Keywords

Clinical success, hyaluronic acid (HA)-binding
 capacity (HBA score), intracytoplasmic
 sperm injection (ICSI), 'physiologic' ICSI
 (PICSI), sperm concentration

History

Received 3 March 2014
 Revised 3 June 2014
 Accepted 8 June 2014
 Published online ■■■

The selection of sperm for ICSI is commonly done via the
 microscopic assessment of motility and morphology. Sperm
 classified as normal morphology are found to host chromo-
 somal aberrations [Celik-Ozenci et al. 2004]. Disomic and
 diploid sperm have been found in all categories of morpho-
 logical classification [Zavaczki et al. 2006].

Hyaluronic acid (HA) is thought to be critical within the
 female reproductive tract when selecting functionally com-
 petent sperm during *in vivo* fertilization. The human oocyte is
 surrounded by the cumulus oophorus, whose major compo-
 nent is HA, a high molecular weight glycosaminoglycan.
 Developmentally mature sperm were found to bind to HA gels
 similarly to the binding between sperm and zona pellucida.
 The binding of sperm to HA *in vitro* is a selection process.
 In another study it has been demonstrated that there are

121 exceptions when motile sperm do not bind to HA [Huszar
122 et al. 2003].

123 Simultaneously with cytoplasmic extrusion in spermiogen-
124 esis, there is also a remodeling of the plasma membrane that
125 facilitates the formation of the zona pellucida- and HA-
126 binding sites [Huszar et al. 1997, 2003]. HA-binding
127 associated with the presence of the HA receptors on the
128 sperm surface is related to sperm development [Huszar and
129 Vigue 1993]. Sperm with HA-binding ability are viable
130 having either intact or slightly capacitated acrosomal status
131 and appear devoid of significant DNA degradation [Huszar
132 et al. 2007; Yagci et al. 2010].

133 Diminished expression of the heat shock-related 70 kDa
134 protein 2 (HspA2), a testis-specific chaperone protein, part of
135 the meiotic synaptonemal complex, causes meiotic defects
136 leading to aneuploidies [Kovanci et al. 2001]. There is a
137 relationship between diminished sperm development (asso-
138 ciated with oligozoo/asthenozoo/teratozoospermia), low
139 levels of HspA2 expression, increased frequency of chromo-
140 somal aneuploidies, the presence of apoptotic process, and
141 fragmented DNA [Huszar and Vigue 1993; Huszar et al.
142 2000, 2003, 2007; Yagci et al., 2010]. *In vitro* solid-state HA-
143 binding facilitates the selection of individual mature sperm
144 with low levels of chromosomal aneuploidies [Jakab et al.
145 2005]. Based on the percentage of bound sperm, three binding
146 zones were established: excellent (>80%), moderate (60-
147 80%), and low (<60%). The HA sperm selection method for
148 ICSI might reduce the potential genetic complications and
149 adverse public health effects of ICSI [Jakab et al. 2005].

150 In the present study we examined (1) the clinical success of
151 ICSI with HA-selected sperm ('physiologic' ICSI, PICSI)
152 compared to the conventional ICSI, (2) the necessity to
153 differentiate patients according to the initial HA-binding
154 assay result, and (3) whether the sperm concentration or the

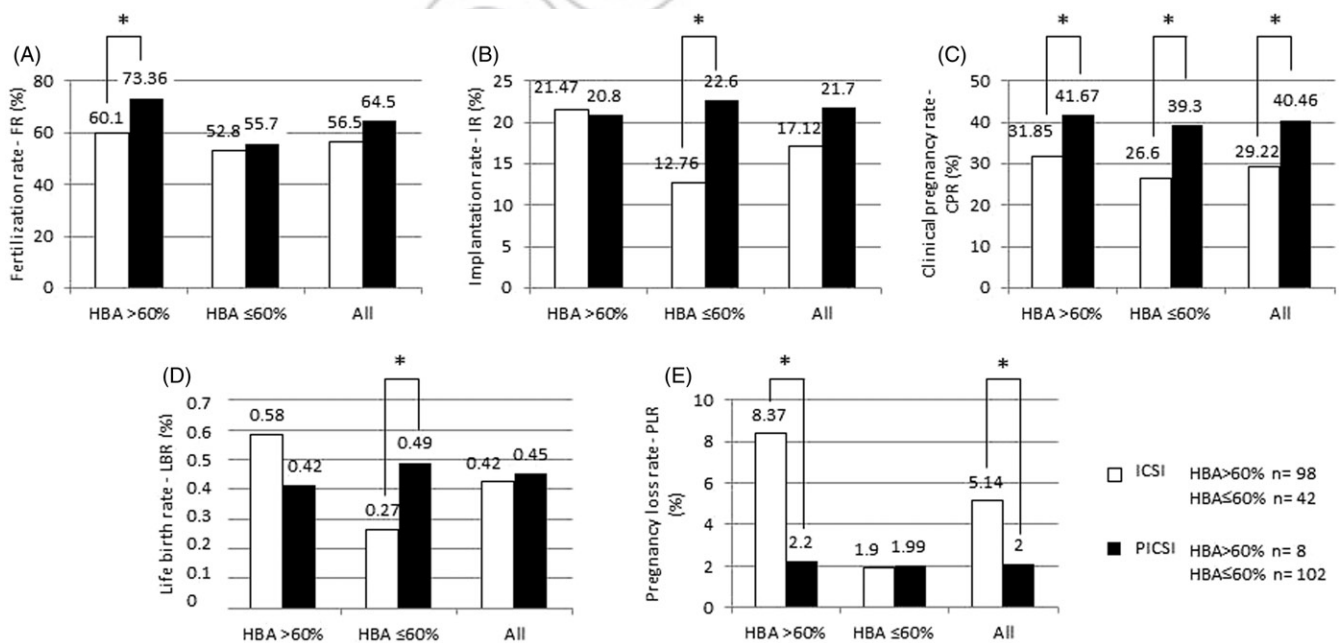
181 HA-binding ability can give more information about fertil-
182 ization outcome. For this purpose, (i) we analyzed the clinical
183 outcome (fertilization rate (FR), implantation rate (IR),
184 clinical pregnancy rate (CPR), life birth rate (LBR), and
185 pregnancy loss rate (PLR)) of 250 infertile couples (idiopathic
186 infertile couples or infertility caused by male factor infertility)
187 conceived by PICSI or ICSI, (ii) we carried out initial HA-
188 binding score of all male partners, formed different groups
189 according to the results, and then analyzed the clinical
190 outcome, and finally (iii) we studied the correlation between
191 the sperm concentration, HA-binding capacity, and fertiliza-
192 tion rate.

193 Results

194 Clinical outcome of PICSI vs. ICSI

195 An average of 10.6 Metaphase II oocytes and 7.9 2PN zygotes
196 were produced. The average fertilization rate was 62.7%.
197 The male patients demonstrated average sperm concentration
198 of $33.3 \times 10^6/\text{mL}$ with 52.6% HBA score. In all cases the
199 morphology of the embryos was normal (<30% fragmenta-
200 tion; [WHO 2010]). The results of Study 1 are summarized in
201 Figure 1.

202 In the ICSI group the average sperm concentration proved
203 to be $39.2 \times 10^6/\text{mL}$, the HBA score 62.5%, the FR 56.5%, the
204 IR 17.12%, the CPR 29.22%, the LBR 0.42%, and the PLR
205 5.14%, respectively. In the ICSI group, where the HBA score
206 was >60%, the average sperm concentration was $47.4 \times 10^6/\text{mL}$,
207 HBA score was 75.7%, FR was 60.14%, IR was 21.47%,
208 CPR was 31.85%, LBR was 0.58%, and PLR was 8.37%. In
209 the ICSI group with HBA score $\leq 60\%$ the parameters found
210 were as follows: average sperm concentration: $20.1 \times 10^6/\text{mL}$,
211 the HBA score: 31.7%, the FR: 52.85%, the IR: 12.76%, the
212 CPR: 26.6%, the LBR: 0.27%, and the PLR: 1.9%.



177 Figure 1. Clinical outcome of PICSI vs. ICSI. (A) Fertilization rate (FR) of patients with HBA >60%, patients with HBA ≤60%, and all patients; (B)
178 Implantation rate (IR) of patients with HBA >60%, patients with HBA ≤60%, and all patients; (C) Clinical pregnancy rate (CPR) of patients with HBA
179 >60%, patients with HBA ≤60%, and all patients; (D) Life birth rate (LBR) of patients with HBA >60%, patients with HBA ≤60%, and all patients;
180 (E) Pregnancy loss rate (PLR) of patients with HBA >60%, patients with HBA ≤60%, and all patients. Statistical significance (*) is indicated at
181 $p < 0.05$. HBA: hyaluronic acid (HA) binding ability; ICSI: intracytoplasmic sperm injection; PICSI: ICSI with HA-selected sperm ('physiologic' ICSI).

In the PICS group the average sperm concentration was $25.6 \times 10^6/\text{mL}$ associating with HBA score of 34.8%; the FR was 64.5%; the IR 21.7%, the CPR 40.46%, the LBR 0.45%, and the PLR 2%. In the PICS group, where the HBA score was $>60\%$, the average sperm concentration was measured as $35.5 \times 10^6/\text{mL}$, the HBA score 66.7%, the FR 73.36%, the IR 20.8%, the CPR 41.67%, the LBR 0.42%, and the PLR 2.2%. In the PICS group with HBA score $\leq 60\%$ the same parameters are as follows: average sperm concentration: $24.8 \times 10^6/\text{mL}$, the HBA score: 32.3%, the FR: 55.7%, the IR: 22.6%, the CPR: 39.3%, the LBR: 0.49%, and the PLR: 1.99%.

The FR of the PICS group with $>60\%$ HBA was significantly higher than that in the ICSI group with $>60\%$ HBA ($p < 0.01$). The IR of the PICS group with $\leq 60\%$ HBA proved to be significantly higher than that in the ICSI group with $\leq 60\%$ ($p < 0.001$). The CPR was significantly higher in every PICS group compared to the ICSI groups ($p < 0.01$). We have observed a significantly higher LBR in the PICS group with $\leq 60\%$ HBA compared to ICSI patients with the same HBA ratio ($\leq 60\%$; $p < 0.001$). PLR was significantly lower in PICS patients and in the PICS group with above 60% HBA compared to the ICSI group and the ICSI patients with $>60\%$ HBA, respectively ($p < 0.0001$).

Characterization according to the HBA score

Patients were further differentiated into two groups: HA-excellent ($>70\%$) and HA-low bound sperm ($<50\%$) groups based upon their HA binding capacity (%). The ICSI group with excellent HBA consisted of 69 couples and the ICSI group with $<50\%$ HBA contained 32 patients. The PICS group with excellent HBA consisted of six couples and the PICS group with $<50\%$ HBA contained 87 patients. The results of Study 2 are summarized in Figure 2.

In the ICSI group, where the HBA score was $>70\%$, the average sperm concentration was $54.1 \times 10^6/\text{mL}$, the HBA score 84.1%, the FR 70.14%, the IR 21.5%, the CPR 35.8%, the LBR 0.58%, and the PLR 8.3%, respectively. In the ICSI group with $<50\%$ HBA the average sperm concentration proved to be $16.13 \times 10^6/\text{mL}$, the HBA score 24%, the FR 47.24%, the IR 12.5%, the CPR 30.8%, the LBR 0.26%, and the PLR 9.15%. In the PICS group, where the HBA score was $>70\%$, the average sperm concentration was $54.1 \times 10^6/\text{mL}$, the HBA score 83.1%, the FR 73.4%, the IR 20.8%, the CPR 41.7%, the LBR 0.4%, and the PLR 2.2%. In the PICS group with $<50\%$ HBA the average sperm concentration was $24.1 \times 10^6/\text{mL}$, the HBA score 28.5%, the FR 55.42%, the IR 24.02%, the CPR 41.2%, the LBR 0.5%, and the PLR 4.65%.

The FR, IR, CPR, and LBR of the PICS group with $<50\%$ HBA were significantly higher and the PLR was significantly lower than those in the ICSI group with $<50\%$ HBA ($p < 0.01$). The PLR of the PICS group with $>70\%$ HBA proved to be significantly lower than that in the ICSI group with $>70\%$ HBA ($p < 0.0001$).

Correlation analysis between sperm concentration, HA-binding capacity, and fertilization rate

The Pearson correlation (r) between the sperm concentration and HA-binding capacity was determined by comparing all samples in the ICSI and PICS groups. The Pearson correlation between the sperm concentration and FR and between the HBA score and FR independent of treatment (ICSI or PICS), were respectively calculated. The results of the correlation analysis are summarized in Figure 3.

A statistically significant positive correlation was found between the sperm concentration and the HA-binding

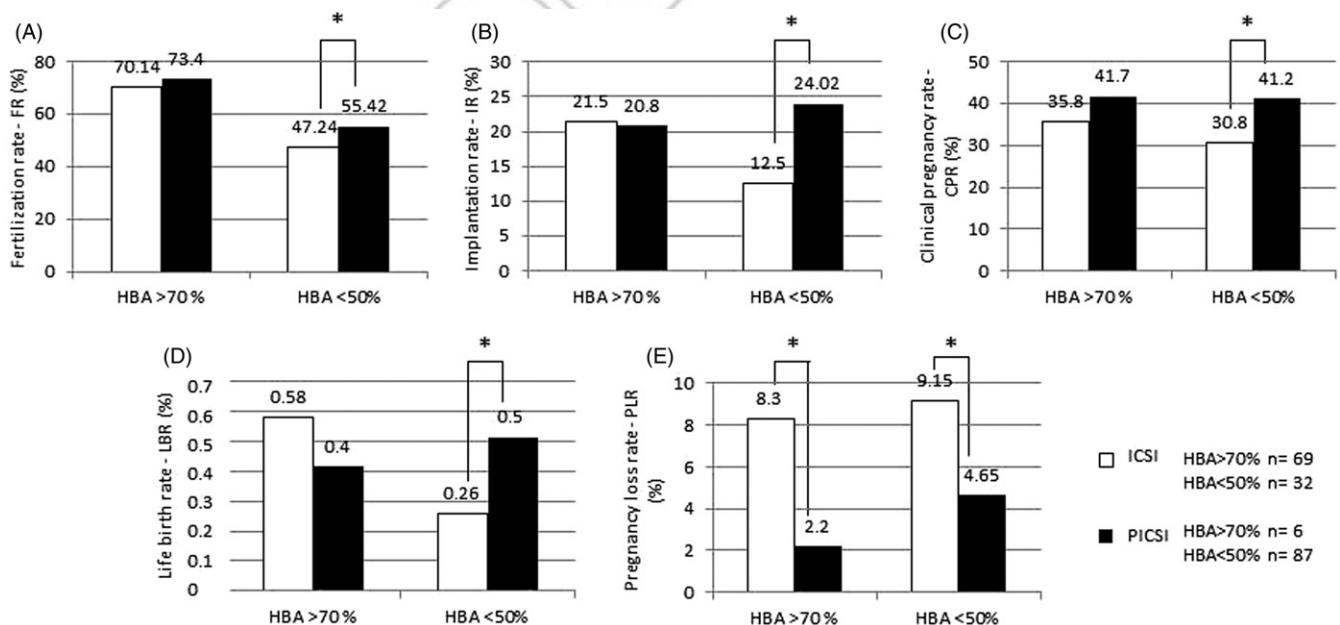


Figure 2. Clinical outcome of PICS vs. ICSI of patients with HA-excellent (HBA score $>70\%$) and HA-low bound sperm (HBA score $<50\%$). (A) Fertilization rate (FR) of patients with HBA $>70\%$ and patients with HBA $<50\%$; (B) Implantation rate (IR) of patients with HBA $>70\%$ and patients with HBA $<50\%$; (C) Clinical pregnancy rate (CPR) of patients with HBA $>70\%$ and patients with HBA $<50\%$; (D) Life birth rate (LBR) of patients with HBA $>70\%$ and patients with HBA $<50\%$; (E) Pregnancy loss rate (PLR) of patients with HBA $>70\%$ and patients with HBA $<50\%$. Statistical significance (*) is indicated at $p < 0.05$. HBA: hyaluronic acid (HA) binding ability; ICSI: intracytoplasmic sperm injection; PICS: ICSI with HA-selected sperm ('physiologic' ICSI).

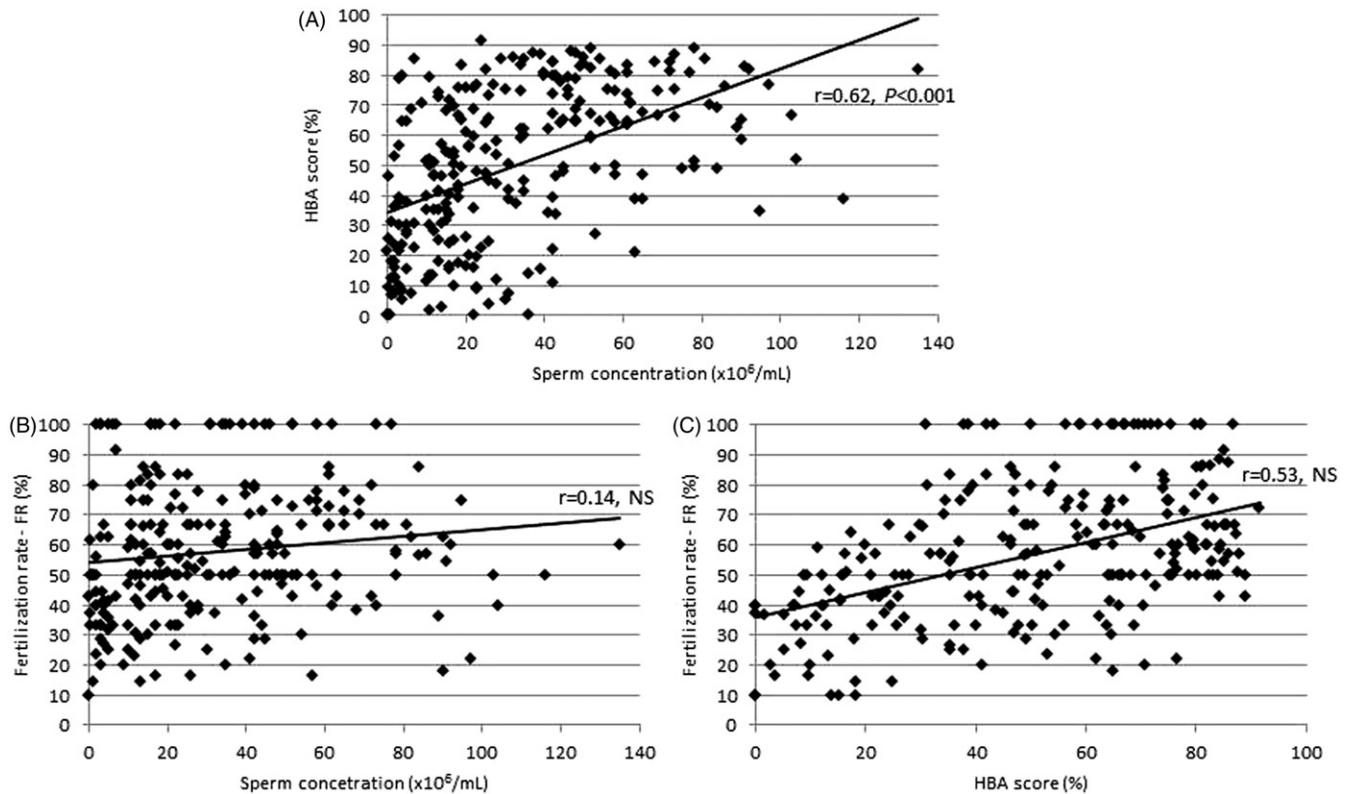


Figure 3. Correlation analysis between the sperm concentration and HBA score (A), between the sperm concentration and FR (B) and between the HBA score and FR (C). A statistically significant correlation was found between the sperm concentration and the HA-binding capacity. Higher positive correlation was found between HBA score and FR than between sperm concentration and FR. HBA: hyaluronic acid (HA) binding ability; FR: fertilization rate; r : Pearson correlation coefficient; NS: non-significant.

capacity ($r=0.62, p<0.001$). We found a higher positive correlation between HBA score and FR ($r=0.53, \text{NS}$) than between sperm concentration and FR ($r=0.14, \text{NS}$), but the difference was not statistically significant. In the ICSI and in the PICSI groups a higher positive correlation between HBA score and FR ($r=0.51$ and $r=0.49, \text{NS}$) than between sperm concentration and FR ($r=0.22$ and $r=0.19, \text{NS}$) was observed. This association was not statistically significant.

Discussion

We compared conventional ICSI ($n=140$) to ICSI in which the spermatozoa were selected for their capacity to bind to HA (PICSI, $n=110$). We observed a significantly higher FR in the PICSI group with $>60\%$ initial HBA; IR of the PICSI group with $\leq 60\%$ HBA, and CPR in every PICSI group compared to the ICSI groups ($p<0.01$). We also observed a significantly higher LBR in the PICSI group with $\leq 60\%$ HBA compared to ICSI of patients with $\leq 60\%$ HBA ($p<0.001$). PLR was significantly lower in PICSI patients compared to the same parameter in the ICSI group ($p<0.0001$). When the outcome was assessed as a function of the HBA score, the FR, IR, CPR, and LBR of the PICSI group with $<50\%$ HBA were significantly higher and the PLR was significantly lower than in the ICSI group with $<50\%$ HBA ($p<0.01$). A statistically significant positive correlation was found between the sperm concentration and the HA-binding capacity ($r=0.62, p<0.001$). We found a closer relationship between HBA score and FR ($r=0.53, \text{NS}$) than between sperm concentration and FR ($r=0.14, \text{NS}$).

Previous studies regarding the development and function of biochemical and molecular markers of human sperm are supported by the above clinical results. A relationship between HA selected sperm and increased levels of developmental maturity [Cayli et al. 2004; Huszar et al. 1994, 2003], as well as nuclear [Kovanci et al. 2001; Jakab et al. 2005], and cytoplasmic integrity [Huszar et al. 1997; Sakkas et al. 1999] have been demonstrated.

A similar increase in IR, CPR, and lower PLR values was found by Worrilow and colleagues [Worrilow et al. 2006; Worrilow et al. 2007; Worrilow et al. 2012]. Others compared conventional sperm selection and the use of sperm selected from a liquid source of HA and an increased IR was found [Parmegiani et al. 2010]. The same positive trend was observed comparing polyvinylpyrrolidone-ICSI ($n=110$) and PICSI ($n=92$) treatments [Ménézo and Nicollet 2004]. In a study of 50 couples, a higher FR was observed when HA-selected spermatozoa were injected into oocytes [Nasr-Esfahani et al. 2008]. These studies, in accordance with ours, did not demonstrate any negative effect on embryogenesis using HA sperm selection for ICSI, but they all was 'in-house' developed HA slides.

In two further reports, no association was found between HA binding and FR, fragmentation, and embryo quality though they used washed sperm [Choe et al. 2012; Tarozzi et al. 2009]. In another report the clinical outcome of sperm functional assays including HBA was studied [Nijs et al. 2009]. A correlation of HA-binding was found with morphology, but it did not predict FR and CPR. Another recent study did not find any differences in FR, IR, and CPR

481 between ICSI and PCSI patients. The only benefit of
482 injecting HA selected sperm was a lower PLR which
483 consequently translated to a higher LBR, both of
484 which were not statistically significant [Majumdar and
485 Majumdar 2013].

486 No visual integrity of the DNA in selected sperm can be
487 assessed which can basically determine the overall success of
488 ICSI. When natural and assisted reproduction fails defects in
489 sperm chromatin have been blamed [Bungum et al. 2007;
490 Carrell et al. 2007]. Sperm DNA damage was found to be
491 positively correlated with PLR when 11 studies involving
492 1,549 *in vitro* fertilization (IVF) and ICSI cycles was
493 systematically reviewed [Zini et al. 2008]. It is well known
494 that the proportion of immature sperm closely correlates with
495 chromosomal disomies [Kovanci et al. 2001]. The relation-
496 ship between the frequencies of chromosomal aneuploidies
497 and diminished sperm maturity is thought to reflect that
498 cytoplasmic retention and diminished maturity in sperm are
499 associated with a low expression of the HspA2 [Eddy 1999;
500 Huszar et al. 2000]. The relationship between sperm zona
501 pellucida binding competence and maturity has been
502 identified earlier. In the semen samples there were sperm
503 with various degrees of cytoplasmic retention, but all sperm
504 bound to the zona pellucida were mature as characterized with
505 the absence of any cytoplasmic retention. Diminished HspA2
506 chaperone activity found in developmentally immature sperm
507 is thought to be connected with a diminished presence of
508 DNA repair enzymes, causing DNA chain breaks and
509 fragmentation [Dix et al. 1996; Eddy 1999; Huszar et al.
510 2000]. There is a correlation between the decreased levels of
511 expression of the HspA2 chaperone and sperm cellular
512 development as well as IVF success [Ergur et al. 2002;
513 Huszar et al. 1992, 2000]. Van Steirteghem et al. [2002] found
514 increased rates of *de novo* numerical and cytogenetically
515 detectable structural chromosomal aberrations following
516 ICSI. The low concentration of HspA2 in the undeveloped
517 spermatozoa likely suggests numerical chromosomal aberrations
518 in sperm of oligozoospermic or severely oligozoosper-
519 mic men [Huszar et al. 2007].

520 An enhancement of DNA and chromosomal integrity was
521 demonstrated in HA-bound sperm by Yagci et al. [2010] when
522 they analyzed HA-bound sperm with acridine orange fluo-
523 rescence and they did not find DNA fragmentation. Selecting
524 individual mature sperm with low levels of chromosomal
525 disomy, diploidy, and sex chromosome disomy is facilitated
526 by HA-binding and might reduce the potential genetic
527 complications in male candidates for ICSI [Jakab et al.
528 2005]. It has been observed that almost all HA-bound
529 spermatozoa are devoid of persistent histones, which
530 correlated with DNA strand breakage [Sati et al. 2004].

531 After ICSI, no sperm function tests were well correlated
532 with FR. These results are in line with the data of several
533 studies [Bakos et al. 2008; Henkel et al. 2003; Nasr-Esfahani
534 et al. 2008] but contradictory to the data presented above,
535 where we found PCSI proved to be significantly more
536 effective than ICSI in respect of clinical success for patients
537 with a low initial HBA score ($\leq 50\%$). Based on our results
538 HA selection becomes an important factor in cases with low
539 binding scores, where the expected number of normal sperm
540 is much lower. It has been observed in a single study where a

correlation was found between sperm HA-binding capacity 541
and FR after IVF [Pregl Breznik et al. 2013]. Our results 542
indicate that sperm selection by HA binding is promising and 543
significantly improves the success of the result in patients 544
with a low HBA score. We conclude that HBA screening prior 545
to ICSI may be useful to increase clinical success. It has been 546
demonstrated that injection of spermatozoa recovered from 547
HA-containing products had no negative effects on post- 548
injection zygote development [Balaban et al. 2003; Barak 549
et al. 2001]. A statistically significant reduction in PLR was 550
observed in patients with a low HBA score. The use of HA 551
sperm selection may be considered in patients with an initial 552
HBA score of $\leq 50\%$. To determine the use of HA-bound 553
sperm in ICSI, the use of HBA score would be beneficial 554
since it could offer a balance to unnecessary treatment. 555

556 Materials and Methods 557

558 Patients 559

A total of 250 couples referred to the Assisted Reproduction 560
Center, Kaali Institute, Medical and Health Science Center, 561
University of Debrecen for ICSI were studied. The study was 562
done between January 2012 and March 2013. In this period, 563
140 ICSI and 110 PCSI were carried out on the basis of the 564
sperm HA-binding ability of the male partner (HBA score): 565
when initial HBA score was $>60\%$ ICSI was carried out 566
($n=98$), in cases with HBA score $\leq 60\%$ PCSI was 567
performed ($n=102$). The ICSI group included 42 couples 568
where the sperm number and the HBA score was low and 569
unsuitable for PCSI. We carried out eight control PCSI 570
where the HBA score was $>60\%$. 571

572 Women under the age of 40 (mean: 33.18, range: 22–40) 573
with regular (21–35 days) menstrual cycles, with normal 574
baseline follicle stimulating hormone (FSH) level (≤ 12 IU/L) 575
were eligible. Within the overall studied population the 576
average male age was 35.8 years (range: 23–45). Patients 577
excluded from the study were as follows: those from whom 578
testicular sperm were taken, who got donor or cryopreserved 579
gametes, received preimplantation genetic diagnosis, under- 580
went sperm sorting procedures, patients whose maternal age 581
was >40 years, and those who demonstrated a sperm count 582
 $<10,000$ motile sperm/mL.

583 Prior to the study, all patients were given detailed 584
information about the aim and method of investigation and 585
their consents were obtained. All protocols had to be 586
approved by the author's respective Institutional Review 587
Board (IRB) for human subjects (IRB reference number: 588
2976/2012-EHR). 589

590 Stimulation protocols 591

Standard stimulation protocols, gonadotropin releasing hor- 592
mone (GnRH) agonist long ($n=75$), short ($n=120$), and 593
GnRH antagonist ($n=55$), were used. The stimulation 594
protocol and dose of gonadotropins were not standardized 595
for the study; the decision was made by the physician. 596

597 For the long protocol, GnRH agonist was started in the 598
midluteal phase. During suppression the dosage was reduced 599
to half and stimulation with either recombinant FSH or human 600
menopausal gonadotropin, or the combination of the two were

used. For the short protocol, the GnRH agonist was started on cycle day 2 and gonadotropin stimulation was initiated on day 3. In the case of the antagonist protocol, stimulation was started on day 2 of the cycle and the GnRH antagonist was started when the largest follicles had reached 13–14 mm in size. When at least two follicles reached 17 mm in diameter, recombinant human chorionic gonadotropin (hCG) was used to trigger ovulation. Transvaginal oocyte retrieval was performed 35–36 h later.

610

611 Semen analysis and hyaluronic acid binding assay

612 Semen specimens were collected after a requested abstinence
613 of two to three days on the day of the oocyte retrieval. The
614 sperm sample was maintained at room temperature (18–
615 28 °C) for 30 to 60 min to allow it to liquefy. Semen analysis
616 was performed manually according to WHO guidelines and
617 morphology was examined using strict criteria [WHO 2010].
618 The HBA-test (hyaluronic acid binding assay) (MidAtlantic
619 Diagnostics, Marlton, NJ, USA) was carried out at room
620 temperature: the sample was mixed and a pipette of 7–10 µl
621 was placed near the center of the chamber. The CELL-VU
622 gridded cover slip was located over the chamber to avoid air
623 bubble formation. The chamber was incubated at room
624 temperature for at least 10 min, but not more than 20 min:
625 this period proved to be necessary for sperm to bind to HA
626 (according to the HBA-test protocol). The number of bound,
627 motile sperm and the totality of motile sperm was scored. At
628 least 200 spermatozoa in the same square or the entire 100
629 squares were counted. The ratio of hyaluronic binding motile
630 sperm was calculated as follows:

$$632 \quad \% \text{Bound} = 100 \times \text{Bound Motile} / \text{Total Motile}.$$

633

634

635 Fertilization

636 Gradient centrifugation (600 g for 10 min) was used to
637 separate the cellular components of semen (PureCeption™
638 Sperm Washing Solution, SAGE, Pasadena, CA, USA).
639 Following centrifugation the supernatant was removed and
640 the sediment was washed twice (Quinn's Advantage® Sperm
641 Washing Medium, SAGE, Pasadena, CA, USA; 600 g for
642 10 min). The supernatant was removed again and the sediment
643 was diluted.

644 In order to select the morphologically 'best' spermatozoon,
645 sperm were placed into standard ICSI dishes which were later
646 injected into oocytes. We placed the final sperm suspension of
647 PICSU patients upon microdots of hyaluronic acid in the
648 PICSU® Sperm Selection Device (Biocoat, Inc., Horsham, PA,
649 USA) and then overlaid it with oil (SAGE, Pasadena, CA,
650 USA). After an incubation period of 5 to 10 min, HBA sperm
651 were selected as per the manufacturer's instructions. We
652 selected spermatozoa bound to HA in the junction zone of the
653 two droplets and it was easy to detach then by an injecting
654 pipette (ICSI Micropipette; ORIGIO, Charlottesville, VA,
655 USA) and subsequently injected into oocytes.

656

657 Embryo culture

658

659 In the presence of two pronuclei fertilization was confirmed.
660 The embryos were transferred to Quinn's Advantage® Protein

Plus Cleavage Medium at this stage (SAGE) and in 661
microdroplets of 20–25 µL under Washed Oil for Tissue 662
Culture, groups of 3–5 were cultured until the 6–8 cell stage 663
(SAGE). After this, embryos were put to Quinn's Advantage® 664
Protein Plus Blastocyst Medium (SAGE). 665

One, two, or three embryos were transferred following 3 or 666
5 d of fertilization. It was the couple's decision of how many 667
embryos to be transferred after consulting with their physician. 668
The morphology of the embryos was the basis for the transfer. 669

670

671 Statistical analysis

672 Statistical analyses were performed with commercial software
673 SigmaStat and SPSS. Sample normality was assessed using
674 Shapiro-Wilk test, sample homogeneity using Barlett test.
675 Differences in the sperm concentration, HA-binding ability,
676 FR, IR, CPR, LBR, and PLR between the ICSI and PICSU
677 groups were analyzed using Mann-Whitney/Wilcoxon Two-
678 Sample Test, Kruskal-Wallis test (when normality does
679 not exist), and Two-sample t-probe (when normality exists).
680 A value of $p < 0.05$ was considered a significant difference.
681 Correlation analyses between the sperm concentration,
682 HA-binding capacity, and FR using all samples in the two
683 groups were examined with Pearson correlation test.

684 It was the number of eggs fertilized with the given method
685 that determined the fertilization rate (FR) for each patient.
686 Implantation rate (IR) was calculated from the number of
687 intrauterine sacs/the number of embryos transferred in each
688 patient. There is an agreement that clinical pregnancy means
689 that fetal cardiac activity is present within an intrauterine
690 gestational sac. Vaginal ultrasound was used to assess preg-
691 nancy loss rate (PLR) and it means the proportion of patients
692 demonstrating an intrauterine sac at 5–7 w of gestation and
693 those where no fetal cardiac activity was present at 8–10 w of
694 gestation. 695

696

697 Declaration of interest

698 The authors report no declarations of interest. This research
699 was supported by the European Union and the State of
700 Hungary, co-financed by the European Social Fund in the
701 framework of TÁMOP-4.2.4.A/2-11/1-2012-0001 National
702 Excellence Program. 703

704

705 Author contributions

706 Collected the data, conceived and carried out the statistical
707 analyses, evaluated the results, and wrote the first draft of the
708 manuscript: AM; Collected the data, conceived and designed
709 the analyses: EVT; Involved in clinical examination and follow
710 up of the patients: BB, ZT; Conceived and designed the
711 analyses: ZM, AJ, AU; Made substantial contribution to
712 the design and interpretation of data, critically revised the
713 manuscript, and approved the final version to be published: EO. 714

715

716 References

- 716 Bakos, H.W., Thompson, J.G., Feil, D. and Lane, M. (2008) Sperm DNA
717 damage is associated with assisted reproductive technology preg-
718 nancy. *Int J Androl* **31**:518–26. 719
718 Balaban, B., Lundin, K., Morrell, J.M., Tjellström, H., Urman, B. and
719 Holmes, P.V. (2003) An alternative to PVP for slowing sperm prior to
720 ICSI. *Hum Reprod* **18**:1887–9. 720

- 721 Barak, Y., Menezo, Y., Veiga, A. and Elder, K. (2001) A physiological
722 replacement for polyvinylpyrrolidone (PVP) in assisted reproductive
723 technology. *Hum Fertil (Camb)* **4**:99–103.
- 724 Bonduelle, M., Van Assche, E., Joris, H., Keymolen, K., Devroey, P.,
725 Van Steirteghem, A., et al. (2002) Prenatal testing in ICSI
726 pregnancies: incidence of chromosomal anomalies in 1586 karyotypes
727 and relation to sperm parameters. *Hum Reprod* **17**:2600–14.
- 728 Bungum, M., Humaidan, P., Axmon, A., Spano, M., Bungum, L.,
729 Erenpreiss, J., et al. (2007) Sperm DNA integrity assessment in
730 prediction of assisted reproduction technology outcome. *Hum Reprod*
731 **22**:174–9.
- 732 Carrell, D.T., Emery, B.R. and Hammoud, S. (2007) Altered protamine
733 expression and diminished spermatogenesis: what is the link? *Hum*
734 *Reprod Update* **13**:313–27.
- 735 Cayli, S., Sakkas, D., Vigue, L., Demir, R. and Huszar, G. (2004)
736 Cellular maturity and apoptosis in human sperm: creatine kinase,
737 caspase-3 and Bcl-XL levels in mature and diminished maturity
738 sperm. *Mol Hum Reprod* **10**:365–72.
- 739 Celik-Ozenci, C., Jakab, A., Kovacs, T., Catalanotti, J., Demir, R., Bray-
740 Ward, P., et al. (2004) Sperm selection for ICSI: shape properties do
741 not predict the absence or presence of numerical chromosomal
742 aberrations. *Hum Reprod* **19**:2052–9.
- 743 Choe, S.A., Tae, J.C., Shin, M.Y., Kim, H.J., Kim, C.H., Lee, J.Y., et al.
744 (2012) Application of sperm selection using hyaluronic acid binding
745 in intracytoplasmic sperm injection cycles: a sibling oocyte study.
746 *J Korean Med Sci* **27**:1569–73.
- 747 Dix, D.J., Allen, J.W., Collins, B.W., Mori, C., Nakamura, N., Poorman-
748 Allen, P., et al. (1996) Targeted gene disruption of Hsp70-2 results in
749 failed meiosis, germ cell apoptosis, and male infertility. *Proc Natl*
750 *Acad Sci USA* **93**:3264–8.
- 751 Eddy, E.M. (1999) Role of heat shock protein HSP70-2 in spermatog-
752 nesis. *Rev Reprod* **4**:23–30.
- 753 Ergur, A., Dokras, A., Giraldo, J., Habana, A., Kovanci, E. and Huszar,
754 G. (2002) Sperm maturity and treatment choice of in vitro fertilization
755 (IVF) or intracytoplasmic sperm injection: diminished sperm HspA2
756 chaperone levels predict IVF failure. *Fertil Steril* **77**:910–18.
- 757 Henkel, R., Kierspel, E., Hajimohammad, M., Staf, T., Hoogendijk, C.,
758 Mehnert, C., et al. (2003) DNA fragmentation of spermatozoa and
759 assisted reproduction technology. *Reprod Biomed Online* **7**:477–84.
- 760 Huszar, G., Vigue, L. and Morphed, M. (1992) Sperm creatine
761 phosphokinase M-isoform ratios and fertilizing potential of men: a
762 blinded study of 84 couples treated with in vitro fertilization. *Fertil*
763 *Steril* **57**:882–8.
- 764 Huszar, G. and Vigue, L. (1993) Incomplete development of human
765 spermatozoa is associated with increased creatine phosphokinase
766 concentration and abnormal head morphology. *Mol Reprod Dev* **34**:
767 292–8.
- 768 Huszar, G., Vigue, L. and Oehninger, S. (1994) Creatine kinase
769 immunocytochemistry of human sperm-hemizona complexes: select-
770 ive binding of sperm with mature creatine kinase-staining pattern.
771 *Fertil Steril* **61**:136–42.
- 772 Huszar, G., Sbracia, M., Vigue, L., Miller, D.J. and Shur, B.D. (1997)
773 Sperm plasma membrane remodeling during spermiogenetic matu-
774 ration in men: relationship among plasma membrane beta 1,4-
775 galactosyltransferase, cytoplasmic creatine phosphokinase, and creat-
776 ine phosphokinase isoform ratios. *Biol Reprod* **56**:1020–4.
- 777 Huszar, G., Stone, K., Dix, D. and Vigue, L. (2000) Putative creatine
778 kinase M-isoform in human sperm is identified as the 70-kilodalton
779 heat shock protein HspA2. *Biol Reprod* **63**:925–32.
- 780 Huszar, G., Ozenci, C.C., Cayli, S., Zavaczki, Z., Hansch, E. and Vigue,
781 L. (2003) Hyaluronic acid binding by human sperm indicates cellular
782 maturity, viability, and unreacted acrosomal status. *Fertil Steril* **79**:
783 1616–24.
- 784 Huszar, G., Jakab, A., Sakkas, D., Ozenci, C.C., Cayli, S., Delpiano, E.,
785 et al. (2007) Fertility testing and ICSI sperm selection by hyaluronic
786 acid binding: clinical and genetic aspects. *Reprod Biomed Online* **14**:
787 650–63.
- 788 Jakab, A., Sakkas, D., Delpiano, E., Cayli, S., Kovanci, E., Ward, D.,
789 et al. (2005) Intracytoplasmic sperm injection: a novel selection
790 method for sperm with normal frequency of chromosomal aneuploid-
791 ies. *Fertil Steril* **84**:1665–73.
- 792 Kovanci, E., Kovacs, T., Moretti, E., Vigue, L., Bray-Ward, P., Ward,
793 D.C., et al. (2001) FISH assessment of aneuploidy frequencies in
794 mature and immature human spermatozoa classified by the absence or
795 presence of cytoplasmic retention. *Hum Reprod* **16**:1209–17.
- 796 Majumdar, G. and Majumdar, A. (2013) A prospective randomized study
797 to evaluate the effect of hyaluronic acid sperm selection on the
798 intracytoplasmic sperm injection outcome of patients with unex-
799 plained infertility having normal semen parameters. *J Assist Reprod*
800 *Genet* **30**:1471–5.
- 801 Ménéz, Y. and Nicollet, B. (2004) Replacement of PVP by hyaluronate
802 (SpermSlow) in ICSI - impact on outcome. Eighteenth World
803 Congress on Fertility and Sterility, International Federation of
804 Fertility Societies.
- 805 Nasr-Esfahani, M.H., Razavi, S., Vahdati, A.A., Fathi, F. and Tavalae,
806 M. (2008) Evaluation of sperm selection procedure based on
807 hyaluronic acid binding ability on ICSI outcome. *J Assist Reprod*
808 *Genet* **25**:197–203.
- 809 Nijs, M., Creemers, E., Cox, A., Franssen, K., Janssen, M., Vanheusden,
810 E., et al. (2009) Chromomycin A3 staining, sperm chromatin structure
811 assay and hyaluronic acid binding assay as predictors for assisted
812 reproductive outcome. *Reprod Biomed Online* **19**:671–84.
- 813 Palermo, G.D., Neri, Q.V., Hariprashad, J.J., Davis, O.K., Veeck, L.L.
814 and Rosenwaks, Z. (2000) ICSI and its outcome. *Semin Reprod Med*
815 **18**:161–9.
- 816 Parmegiani, L., Cognigni, G.E., Bernardi, S., Troilo, E., Ciampaglia, W.
817 and Filicori, M. (2010) ‘‘Physiologic ICSI’’: hyaluronic acid (HA)
818 favors selection of spermatozoa without DNA fragmentation and with
819 normal nucleus, resulting in improvement of embryo quality. *Fertil*
820 *Steril* **93**:598–604.
- 821 Pregl Breznik, B., Kovac c, B. and Vlaisavljevi c, V. (2013) Are sperm
822 DNA fragmentation, hyperactivation, and hyaluronan-binding ability
823 predictive for fertilization and embryo development in in vitro
824 fertilization and intracytoplasmic sperm injection? *Fertil Steril* **99**:
825 1233–41.
- 826 Sakkas, D., Mariethoz, E. and St. John, J.C. (1999) Abnormal sperm
827 parameters in humans are indicative of an abortive apoptotic
828 mechanism linked to the Fas-mediated pathway. *Exp Cell Res* **251**:
829 350–5.
- 830 Sati, L.G., Ovari, L., Demir, R., Ward, D.C., Bray-Ward, P. and Huszar,
831 G. (2004) Persistent histones in immature spermatids are associated with
832 DNA fragmentation and affect paternal contribution of sperm: a study
833 of aniline blue staining, fluorescence in situ hybridization (FISH) and
834 DNA nick translation. *Fertil Steril* **82**:S52.
- 835 Simpson, J.L. and Lamb, D.J. (2001) Genetic effects of intracytoplasmic
836 sperm injection. *Semin Reprod Med* **19**:239–49.
- 837 Tarozzi, N., Nadalini, M., Bizzaro, D., Serrao, L., Fava, L., Scaravelli,
838 G., et al. (2009) Sperm-hyaluronan-binding assay: clinical value in
839 conventional IVF under Italian law. *Reprod Biomed Online* **19**:35–43.
- 840 Van Steirteghem, A., Bonduelle, M., Devroey, P. and Liebaers, I. (2002)
841 Follow-up of children born after ICSI. *Hum Reprod Update* **8**:111–16.
- 842 WHO (2010) WHO laboratory manual for the examination and
843 processing of human semen. World Health Organization, 5th edition.
844 p. 271.
- 845 Worrirow, K.C., Huynh, H.T., Bower, J., Peters, A.J. and Johnston, J.B.
846 (2006) The clinical impact associated with the use of PICSI-derived
847 embryos. *Fertil Steril* **86**:S62.
- 848 Worrirow, K.C., Huynh, H.T., Bowers, J.B., Anderson, A., Schillings, W.
849 and Crain, J. (2007) PICSI versus ICSI: Statistically significant
850 improvement in clinical outcomes in 240 in vitro fertilization (IVF)
851 patients. *Fertil Steril* **88**:S37.
- 852 Worrirow, K.C., Eid, S., Woodhouse, D., Perloe, M., Smith, S., Witmyer,
853 J., et al. (2012) Use of hyaluronan in the selection of sperm for
854 intracytoplasmic sperm injection (ICSI): significant improvement in
855 clinical outcomes—multicenter, double-blinded and randomized con-
856 trolled trial. *Hum Reprod* **28**:306–14.
- 857 Yagci, A., Murk, W., Stronk, J. and Huszar, G. (2010) Spermatozoa
858 bound to solid state hyaluronic acid show chromatin structure with
859 high DNA chain integrity: an acridine orange fluorescence study.
860 *J Androl* **31**:566–72.
- 861 Zavaczki, Z., Celik-Ozenci, C., Ovari, L., Jakab, A., Sati, G.L., Ward,
862 D.C., et al. (2006) Dimensional assessment of X-bearing and
863 Y-bearing haploid and disomic human sperm with the use of
864 fluorescence in situ hybridization and objective morphometry. *Fertil*
865 *Steril* **85**:121–7.
- 866 Zini, A., Borman, J.M., Belzile, E. and Ciampi, A. (2008) Sperm DNA
867 damage is associated with an increased risk of pregnancy loss after
868 IVF and ICSI: Systematic review and meta-analysis. *Hum Reprod* **23**:
869 2663–8.