

Department of Restorative Dentistry, Faculty of Dentistry,  
Medical and Health Science Center, University of Debrecen

## The influence of chronic apical periodontitis on oral and general health

DR. ILDIKO J. MARTON

In healthy condition, the periodontal space between the root surface and the alveolar bone is relatively poor in cells. In case of root canal infection however, large number of immune-inflammatory cells infiltrate the periapical region of the affected teeth. A major issue is if and to what extent microbial and host cells contribute to lesion formation and whether the local inflammation may impair general health. The question is important as it may fundamentally influence the therapeutic strategy in patients with apical periodontitis. The aim of this paper is to review the results of recent experimental and clinical observations that investigate the importance of cellular interactions in exerting protective and destructive effects in periapical inflammatory lesions. The majority of studies indicate that the lesion would not develop in the absence of permanent release of bacteria and their by-products from the infected root canal. On the other hand, the formation of the classical granulation tissue is dependent on the presence and proper function of host cells and regulatory molecules. The dynamic encounter of root canal microbiota and the local immune system prevents overwhelming bacterial infiltration of the periradicular space but it is also connected with degenerative changes, most importantly bone resorption, resulting ultimately in tooth loss. However, by the use of proper endodontic methods, the lesion can be successfully treated in the majority of cases. Remineralization of the lost hard tissue will occur or the lesion will transform into an inert periapical scar.

Key words: apical periodontitis, inflammation, microbiology, immunity

### Introduction

Apical periodontitis is an inflammation of periapical tissues in response to irritative root canal stimuli. Although the initiating event is most commonly the infection of the root canal, lesion progression – and repair – is largely determined by local interactions of host cells. The periapical inflammatory infiltrate consists of the cellular and soluble components of the specific and nonspecific branches of the immune system [4, 22].

Depending on the intensity and duration of the pro-inflammatory stimuli, the periapical reaction may take either an acute or a chronic course. The histomorphology of the lesion correlates with the clinical stage of apical periodontitis [4]. Acute apical periodontitis is characterized by vasodilation, vascular congestion and edema. Neutrophil leukocytes and macrophages infiltrate the periapical ligament. The first line of host defense can effectively limit microbial invasion from the infected root canal. Virulent root canal microbes may establish a film on the external root surfaces or may form nests within the previously healthy periapex. In response, an abundant number of neutrophil leukocytes accumulate and the incipient acute lesion develops into primary periapical abscess [22]. Acute exacerbation of a pre-existing chronic lesion results in the development of a secondary abscess with similar mor-

phology. In the incipient lesion, lamina dura disruption and bone resorption of the neighbouring spongiosa can be demonstrated by imaging approaches. Both primary and secondary abscess formation are accompanied by further significant loss of hard tissues giving space for the formation of a chronic lesion [42].

In chronic periapical lesions, which develop either as periapical granulomas or radicular cysts, exogenous irritative agents, usually root canal microbiota and their by-products, and host defense factors are in a dynamic equilibrium. In these cases, the immune mechanisms are not able to destroy and completely eliminate the pathogenic agents but, by forming a histologic as well as functional barrier, effectively prevent their further invasion [72]. The histological appearance of periapical granulomas represents every momentum of the periradicular inflammatory process from the acute incipient response to the burned-out end-stage lesion. Neighbouring histological structures, called necrotic, exudative, granulomatous and fibrous zones, penetrate into the surrounding alveolar bone in an onion leaf-like fashion starting from the apical foramen (Fig. 1). According to the predominant zone(s), lesions can be classified as exudative, granulomatous, granulofibrotic and fibrotic granulomas [67, 105].

Over 50% of chronic periapical lesions contain epithelial strands originating from the epithelial rests of



Malassez [71, 92]. Coalescing epithelial strands form radicular cysts with a frequency varying from 6 to 55% of the lesions. The cavity of pocket or bay cyst communicates with the root canal. In contrast, the true cyst is independent of the root canal system. The epithelial lining of cysts is embedded into the inflammatory granulation tissue [22, 82, 84].

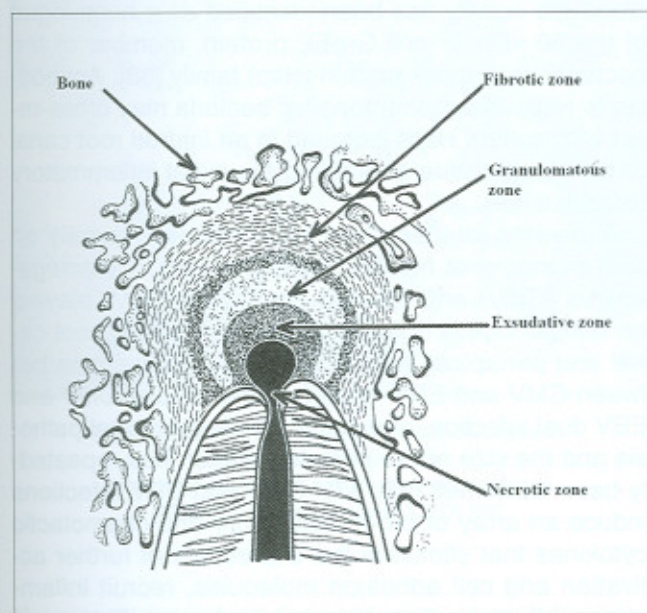


Figure 1.  
Schematic structure of periapical granuloma

Apical periodontitis is a significant oral disease, as it may cause local symptoms, including pain and as it results in alveolar bone resorption and ultimately, tooth loss [72, 84]. The lesion has the potential to compromise general health, as well. Using contemporary non-surgical endodontic treatment, periapical lesions can be cured in about 85–90% of patients. Radiologic signs of recovery can be seen in 89% of cases after 1 year. However, complete disappearance of the lesion may take more than 2 years, indicating that active functioning of host repair mechanisms is as much important as the successful eradication of root canal infection. True cysts are characterized by a much higher endodontic treatment failure rate, therefore some investigators consider these lesions to be autonomous once initiated [14, 37, 74, 76].

In this article the delicate interplay of microbial as well as host cell interactions, which play a role in the initiation and in the development of different phases of the lesion including the healing process, will be reviewed. A brief overview will also be provided on the different mechanisms how apical periodontitis may impair general health of patients.

### Microbial pathogenic factors in apical periodontitis

There is overwhelming evidence, including our own investigation, that the vast majority of the root canal of teeth with periapical lesions is infected [57]. Endodontopathic bacteria represent only a small minority of the over 500 species that colonize the healthy human mouth [98]. Early pulpal infections are characterized by aerobic and facultative anaerobic species. In parallel with local oxygen consumption, obligate anaerob bacteria become predominant [49].

A recent, carefully planned and executed histological study, investigating 50 human periapical lesions demonstrated, that bacteria were always present in the root canal of the affected teeth. Interestingly, in 18 of the 50 roots there was inflamed but vital pulp tissue in the apical part of the root canals [82]. This study has confirmed earlier investigations reporting that chronic periapical lesions were rarely contaminated except for actinomycotic infections. The presence of bacteria was repeatedly demonstrated only in the immediate vicinity of the apical foramen and in resorptive lacunae neighbouring the main apical foramen. Even in periapical abscesses and exudative-type of chronic lesions, bacteria only colonized in necrotic areas – surrounded by abundant, actively phagocytosing neutrophil leukocytes [45, 50, 73, 88].

Bacteria however, do not need to emigrate from the infected root canal to stimulate a periapical inflammatory response. There is a number of soluble bacterial products with a powerful pro-inflammatory potential. These compounds may attract phagocytes, representing the first line of defense, directly or by inducing the expression of pro-inflammatory cytokines and cell adhesion molecules by resident host cells. Other bacterial molecules can stimulate host regulatory molecules (Table I). Lipopolysaccharides (LPS), present in and released from cell walls of Gram-negative bacteria are probably the best characterized components of endodontopathic bacteria to induce cytokines and other inflammatory compounds. LPS isolated from *A. actinomycetemcomitans*, *F. nucleatum*, *P. gingivalis* and *Prev. intermedia* have been shown to induce the production of an array of pro-inflammatory and chemotactic cytokines including granulocyte-macrophage colony-stimulating factor (GM-CSF), interferon (IFN)- $\gamma$ , interleukin (IL)-1 $\alpha$  and - $\beta$ , IL-6, IL-8, macrophage chemotactic peptide (MCP)-1 and tumor necrosis factor (TNF)- $\alpha$  [104]. Local IL-1, IL-6 and GM-CSF production was repeatedly detected in human dental pulp and periapical lesions [2, 47, 79, 102]. Interestingly, LPS preparations extracted from endodontopathic bacteria were shown to induce cytokine secretion in oral cells but not in skin-derived epithelial cells [19]. In contrast to most LPS preparations, *P. gingivalis*-derived LPS was shown to downregulate IL-8 and major histocompatibility complex (MHC) Class II antigens in oral



epithelial cell lines. This immunosuppressive effect of *P. gingivalis* LPS may contribute to the successful bacterial invasion of periapical tissues [11, 19, 29].

There are other cell components of endodontopathic bacteria that are able to initiate inflammation [104]. Fimbrial protein extracts from *P. gingivalis* were demonstrated to increase the production of pro-inflammatory cytokines in stimulated host cells [20, 21]. A 12-mer amino acid sequence between residues 69 and 80, connected with the biological activities of the native fimbrial protein has been identified, opening a pathway for developing targeted therapy [75].

Table I.

Host regulatory factors induced by endodontopathogenic microorganisms

Cell adhesion molecules and ligands
ICAM-1
CD11/18
LFA-1
VLA-5, VLA-6
CD29, CD49
Chemokines and receptors
IL-8
KC
Macrophage inflammatory protein-1 and 2
Rantes
Pro-inflammatory cytokines, receptors and antagonists
GM-CSF
IL-1
IL-1R
IL-1Ra
IL-6
IFN- $\gamma$
TNF
Reactive oxygen and nitrogen intermediates and arachidonic acid metabolites
Superoxid anion
Hydrogenperoxid
Nitrogen-monoxid
Leukotrienes
Prostaglandines
Transcription factors
NF- $\kappa$ B
IE2-protein

The biological activity of surface-associated material (SAM) of certain Gram-negative bacteria may ex-

ceed that of corresponding LPS preparations. SAM prepared from *A. actinomycetemcomitans*, *E. corrodens* and *P. gingivalis* was shown to directly stimulate IL-6 synthesis of human gingival fibroblasts without affecting transcription of IL-1 and TNF genes that induce the expression of IL-6 by positive loops of stimulation upon challenge with LPS [80]. A 64 kDa protein component of *A. actinomycetemcomitans* SAM with potent osteolytic activity has been identified as a homologue of the 60 kDa *E. coli* GroEL protein, member of the bacterial heat shock protein (Hsp) family [38]. Antibodies to Hsps of endodontopathic bacteria may cross-react with human Hsps exposed in an injured root canal or periapical tissue, promoting the initial inflammatory response [89].

A growing attention has been focused recently on two members of human herpes viruses, *Cytomegalovirus* (CMV) and *Ebstein-Barr virus* (EBV) playing an etiopathogenic role in the development of root canal and periapical lesions. A positive correlation between CMV and EBV infection, in particular CMV and EBV dual infection, and symptomatic periapical pathosis and the size of the periapical lesion has repeatedly been confirmed [86, 87]. CMV and EBV infections induce an array of pro-inflammatory and chemotactic cytokines that stimulate the expression of further activation and cell adhesion molecules, recruit inflammatory effector cells to the site of infection. They were shown to dysregulate the receptor activator of nuclear factor- $\kappa$ B ligand (RANKL)/osteoprotegerin system, thereby contributing to alveolar bone resorption. These effects may become especially powerful in synergy with LPS [93, 107].

#### The role of host cells in the initiation and progression of the apical periodontitis lesion

Cells within the periodontal ligament surrounding the healthy root apex constitutively express cell adhesion molecules and chemokines in a low level providing a weak stimulus for attracting and activating polymorphonuclear leukocytes and macrophages [28, 48]. This activity of innate immunity has been suggested to play a key role in maintaining clinically healthy periodontal and periapical tissues [11, 19]. In acute incipient apical periodontitis root canal bacteria reach the tooth apex resulting in the inflammation of the apical ligament and the neighbouring spongiosa accompanied by the well-known symptoms, such as pain, tenderness to percussion. Soluble microbial components upregulate the expression of first line pro-inflammatory cytokines, chemokines, cell adhesion molecules and low molecular weight inflammatory mediators, such as NO, inducing a robust influx of inflammatory cells into the periodontal ligament (Fig. 2) [17, 18, 62]. Subsequently, a second wave of inflammatory mediators, produced by the



stimulated resident and newly recruited cells, may amplify, propagate and prolong the expression of cell adhesion molecules and chemokine genes [66, 78].

In cases of noninfectious pulp necrosis, root canal and periapical inflammation can be elicited by sensory neuropeptides that are released from nociceptive nerve endings by a variety of noxious stimuli [5, 24].

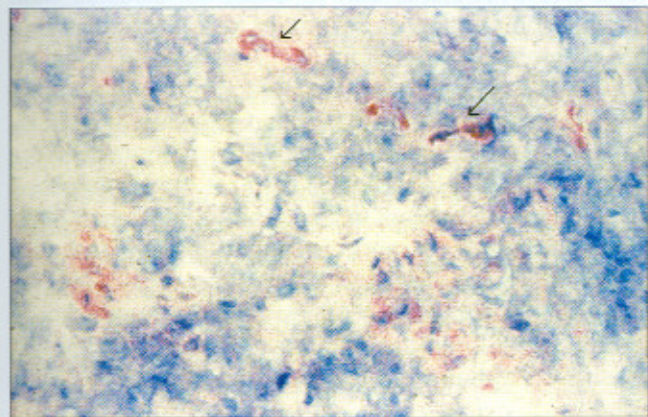


Figure 2. MCP-1 expression in endothelial cells of the granulomatous zone  
Alkaline phosphatase anti-alkaline phosphatase (APAAP) labeling (arrows; X350) [62]

Neuropeptides induce the production of pro-inflammatory cytokines [51]. Both in human and experimental ratine periapical granuloma samples neuropeptide CGRP and substance P (SP) immunoreactivity was confined to nerve fibers in close proximity to blood vessels and inflammatory cells, including antigen-presenting cells (APC) and mast cells [32, 106].

The initial interaction between root canal microbes and resident host cells mount a robust and rapid protective response reaction by recruiting a large number of phagocytic cells, monocyte/macrophages and neutrophil leukocytes, to the sites of bacterial invasion, mainly to the immediate vicinity of the apical foramen [4, 82]. A variety of partially overlapping signals is involved in this process therefore, the exact contribution of individual cytokines and cell-to-cell interactions has yet to be determined [39, 42]. The attraction of infiltrating leukocytes requires them to transiently attach to endothelial cells, migrate through them and then to interact with cells and substrates in the extravascular tissue. This cascade is regulated by a set of cell adhesion molecules and the receptors for them, whereas the chemotactic signal can be provided by complement components of the alternative pathway of complement activation and by chemokines [7, 53, 69]. Expression of chemokines IL-8, MCP-1 and RANTES as well as vascular endothelial growth factor (VEGF)/vascular permeability factor (VPF) was detected in epithelial cells and in inflammatory cells in chronic periapical lesions [46, 62]. Pleiotropic (IL-1, TNF, stem cell factor), lineage-restricted (GM-CSF) and lineage-

specific (granulocyte colony-stimulating factor, G-CSF and monocyte colony-stimulating factor, M-CSF) factors activate these infiltrating leukocytes to stimulate their effector functions [40, 41].

Neutrophil leukocytes, that are virtually absent in the healthy periodontal ligament, reach from the bone marrow via the circulating blood the infected root canal and inflamed periapical tissues. After exerting their protective role by phagocytosing invading bacteria, most neutrophil leukocytes die in human periapical lesions, indicating their short-lived effector cell nature [82, 101]. Newly recruited macrophages outnumber resident ones as demonstrated by immunohistochemistry, in situ hybridization and ultrastructural analysis. Actively phagocytosing macrophages of infiltrating phenotype and neutrophil leukocytes accumulate in great numbers just beneath the periapical abscess and always infiltrate epithelial strands, i.e. they are associated with bacterial and host cells that express chemoattractant molecules [33, 82]. Inhibition of leukocyte recruitment or activation either by disrupting regulatory signals mediated by IL-1, TNF, IL-6 and their receptors or by depressing bone marrow function resulted in a more widespread local bacterial infiltration, even in generalized infections [9, 30, 78].

If phagocytes fail to eliminate completely the invading microorganisms and the irritative effects of soluble bacterial by-products and viruses persist, the lesion becomes chronic and a characteristic, complex granulation tissue will be established as a protective host response to limit periapical tissue damage by localizing root canal microbiota and mitigating the inflammatory response. The initiation and formation of granulomas is largely dependent on the interaction of antigen-presenting cells with CD4<sup>+</sup> T-lymphocytes [91, 96]. Antigen-presenting cells activate CD4<sup>+</sup> T-lymphocytes through their antigen-specific receptors (TCR) in conjunction with MHC Class II molecules [81].

The sequence of events in periapical granuloma formation has been best studied in experimentally induced lesions in the rat [33]. In early lesions, established after 28 days of unsealed pulp exposure, MHC Class II-negative, actively phagocytosing macrophages accumulated in the vicinity of periapical abscess. In contrast, MHC Class II-positive macrophages were found evenly interspersed among other cell types within the cell-rich granulomatous zone. MHC Class II-positive dendritic cells (DC) were also noticed in great numbers, mostly located in the outer, fibrotic border of the lesion. Both MHC Class II-positive macrophages and DC established cell-to-cell contacts with MHC Class II-negative lymphocytes. These results indicate that local antigen presentation takes place in periapical lesions with DC being able to activate both naive and memory T-cells during primary and secondary immune responses, respectively and MHC Class II-positive macrophages, eliciting a secondary immune response [33]. A similar distribution and morphology of



macrophage and APC subpopulations were noticed in human periapical lesions by immunohistochemistry and flow cytometry (Fig. 3) [52, 58, 83].

According to their cytokine expression pattern, CD4<sup>+</sup> T-cells can be classified as T-helper 1 (Th1) cells that produce and secrete IL-2 and IFN- $\gamma$  and Th2 cells that produce and secrete IL-4, IL-5, IL-6, IL-10 and IL-13.

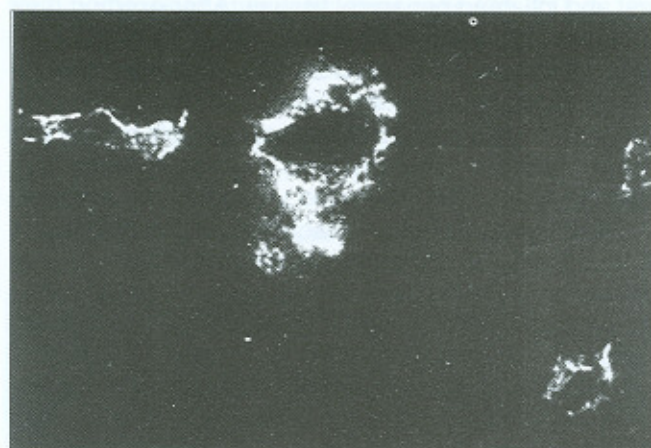


Figure 3  
Surface membrane CD14 positivity of macrophages  
in periapical granuloma sample  
Direct immunofluorescence reaction  
(anti-Leu M3-FITC monoclonal antibody;  
Becton-Dickinson, San Jose, CA, USA; X400)

Positive loops of stimulations exist between macrophages and Th1 cells as well as within the Th1 and Th2 subpopulations, respectively. In contrast, Th1 and Th2 cells exert a mutually inhibitory effect on each other.

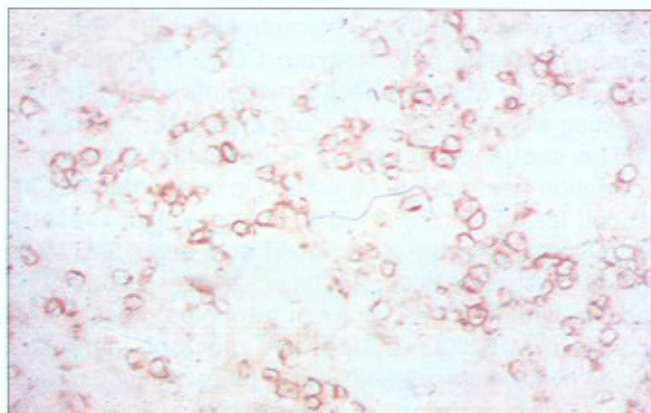


Figure 4  
Cell surface (ringed) expression of  
CD25 antigen (IL2 receptor  $\alpha$ ) on activated T-lymphocytes  
in the granulomatous zone  
Avidin-biotin-peroxidase complex  
(ABC) method (X250) [58]

er. Th1-type cytokines augment cytotoxic T-cell functions, stimulate the expression of pro-inflammatory and bone resorptive molecules in other cell types, whereas Th2-type cytokines participate in B-lymphocyte stimu-

lation to mount a humoral immune response and in dampening the inflammatory reactions [68]. Th1-lymphocytes prevail in early, expanding periapical lesions [31, 35]. As a result of positive loops of stimulation, the number of Th1 cells increases rapidly within the lesion.

In expanding periapical lesions, T-lymphocytes express activation markers, such as interleukin-2 receptor  $\alpha$ -chain (CD25) and MHC Class II molecules, suggesting their active participation in anti-infective and pro-inflammatory reactions (Fig. 4) [58, 99]. This hypothesis was confirmed by demonstrating that immunosuppressive treatment resulted in a decrease in lesion size accompanied by a significant reduction in the number of CD25-positive cells in rat molars exposed to the oral environment when compared with non-treated controls [34]. In more advanced lesions Th2 cells slowly outnumber Th1 lymphocytes. The resulting change in the cytokine milieu is accompanied by a steady increase in the number of B-lymphocytes and plasma cells present in the lesion [52, 56, 102].

#### Cellular interactions resulting in periapical tissue damage

The immune-inflammatory cells of apical periodontitis possess a Janus-faced character. Anti-infective effector mechanisms are not limited to eradicate pathogenic microbes, "innocent bystander" host tissue components also fall victim. The most significant tissue injury involves the resorption of apical dentine and cementum, the mineralized matrix of the neighbouring spongiosa resulting in the loss of supportive alveolar structures and, ultimately, the affected tooth itself. The rate of bone loss is most expressed during the acute exudative phase(s), but it starts as early as in incipient apical periostitis and the osteolytic lesion keeps growing in association with granuloma formation and cyst development.

In course of bone remodeling, bone formation and resorption occur in parallel. The net result of this dynamic process is largely determined by the interaction of osteoblasts and osteoclasts. These two cell types communicate by the surface membrane-bound receptor-ligand pair, receptor activator of NF- $\kappa$ B (RANK) and its ligand RANKL, members of the TNF receptor and ligand families [97]. Osteoblasts express RANKL that binds to RANK present on the surface of osteoclast precursor cells. As a result of RANKL-RANK binding, osteoclast precursors differentiate into mature osteoclasts that become activated and resorb bone. Osteoblasts also express and secrete osteoprotegerin (OPG), the decoy receptor for RANKL, which inhibits RANKL-RANK interaction and thus bone resorption. Reciprocal gene expression of RANKL and OPG has been shown to play a key role in the coupling of osteoblast-osteoclast interaction [70]. Humoral regulatory molecules of bone resorption and formation, such



as hormones and cytokines exert their effects largely by influencing RANKL-RANK interaction directly or by changing the ratio of RANKL/OPG reciprocal gene expression [15]. Prostaglandins appear to act even more indirectly as permissive cofactors of bone-resorptive cytokines [44, 95].

Results from the extensively studied rat periapical granuloma model indicated that both positive and negative regulators of bone resorption were induced at the same time, rapidly after pulp exposure to the oral microflora. Peak levels of RANKL and pro-inflammatory cytokines TNF $\alpha$ , IL-1 $\alpha$  and IL-1 $\beta$  preceded that of negative feed-back molecules OPG and IL-10. Therefore, early lesion was associated with alveolar bone loss and lesion expansion whereas severe progress of bone destruction was controlled in later phases of experimental apical periodontitis [36]. RANKL expression and local synthesis of bone-resorptive cytokines and arachidonic acid derivatives have been repeatedly described in human periapical lesions [10, 65, 85, 93, 103]. Bacterial compounds, such as LPS exert their bone resorptive effects rather indirectly, by inducing human molecules stimulating demineralization, then by direct bone destruction [61].

### Impact of periapical inflammation on general health

Recent epidemiologic studies clearly indicated a causal association between impaired oral health and an increased incidence and severity of several important systemic diseases, such as atherogenesis, atherothrombosis and cardiovascular diseases, adverse pregnancy outcome, lung diseases, diabetes and osteoporosis [90]. The impact of apical periodontitis on general health is difficult to evaluate. In the majority of cases, there are several co-existing oral conditions and risk factors that may contribute to the untoward remote effect(s) of periapical inflammation. There are only anecdotal case reports on patients with root canal infections and apical periodontitis as sole oral inflammatory conditions. A group of investigators from the Oita Medical University observed a 45-year-old Japanese woman with chronic urticaria not responding to antihistamine therapy but she became symptom free after having extracted decayed teeth and teeth with periapical abscesses and given root canal treatment to the other teeth with periapical radiolucencies [94].

The "classic" example for "dental focal infections" is represented by infective endocarditis, where pathogenic bacteria originate from the oral cavity. Application of advanced microbiologic methods provided solid evidence for endodontic origin of bacteremia in patients following root canal treatment of teeth with apical periodontitis [12]. Recent demonstration of the ability of *Porphyromonas gingivalis* to invade endothelial cells shed a new light on the possible connection be-

tween the formation of atherosclerotic lesions and oral bacterial infections [13]. DNA of periodontopathic/endodontopathic bacteria were recovered from atherosclerotic lesions [23].

A number of cross-sectional studies addressing a possible association between oral health and systemic diseases have investigated the presence or the absence of periapical lesions. However, the evidence of these investigations was not conclusive enough to establish or to discard a clear role of the periapical inflammatory lesion in deteriorating general health [54, 63, 64, 100]. Investigating an association between acute cerebrovascular ischemia and chronic and recurrent infection in a case-control study, a research group from the University of Heidelberg has suggested that periapical lesions were more severe in the patient group than in the control group [16]. A group from the Tokyo Dental College observed an association between immune responses to Hsp produced by oral bacteria, chronic marginal and periapical periodontitis, CMV infection, dental metal allergy, and their combinations [43].

In the early 1990s, our research group have performed an interventional follow-up study involving 36, otherwise healthy young adults with apical periodontitis. We have assessed serum and whole blood immune and inflammatory parameters on referral and following root canal treatment and apicoectomy. We have measured the serum concentration of two strong acute phase proteins, C-reactive protein (CRP) and  $\alpha_2$ -macroglobulin, two moderate acute phase proteins,  $\alpha_1$ -antitrypsin and haptoglobin and two weak acute phase proteins, complement component C3 and ceruloplasmin. The levels of the investigated acute phase proteins decreased significantly after treatment [55]. Pretreatment CRP level (mean $\pm$ SD: 6.6 $\pm$ 4.2 mg/L) was high enough to consider it as a cardiovascular risk factor as defined by the guidelines of the American Heart Association [77]. Similarly, we found an elevated whole blood chemiluminescence, which decreased significantly in parallel with the treatment, indicating an activated metabolic and functional state of the peripheral blood neutrophil granulocytes in patients with chronic apical periodontitis [60].

A novel, interesting way of possible association between periapical inflammation and general health is represented by polymorphisms of genes encoding for cytokines and enzymes influencing the function of immune-inflammatory cells. In the presence of certain polymorphic alleles both local and general inflammatory responses can be more intensive and the accompanying complications more severe than in the absence of the given allele [1, 8]. Peripheral blood monocytes of "hyperinflammatory type" patients with periodontal disease produce 3- to 10-fold greater amounts of IL-1 $\beta$  and TNF- $\alpha$  than those obtained from normal individuals [3, 25]. In murine monocytes, an impaired ability to respond to LPS from endodontopathic bacteria has



been attributed to a mutation in the gene that encodes toll-like receptor 4 [6, 27].

### Healing of the periapical lesion

The development of acute incipient apical periodontitis is a consequence of the persistent root canal infection. However, most periapical lesions very rarely heal spontaneously and are not self-limited despite the powerful local protective immune-inflammatory reactions. Effective disinfection of the root canal system combined with proper root canal treatment is essential for the healing of the periapical lesion. After successful elimination of the initiating root canal infection by proper endodontic and restorative treatment, repair mechanisms of the granulation tissue may prevail over tissue destructive events. In contrast to early, expanding periapical granulomas, CD8<sup>+</sup> cytotoxic/suppressor T-lymphocytes usually outnumber CD4<sup>+</sup> helper/inducer T-cells in advanced lesions contributing either to a more effective elimination of infected and destructed cells or to the suppression of the intensity of inflammatory reactions, or both [59, 96]. In parallel with this process, the Th1 nature of the early periapical inflammation changes to Th2-type lesion. Th2 lymphocytes stimulate B lymphocyte production and immunoglobu-

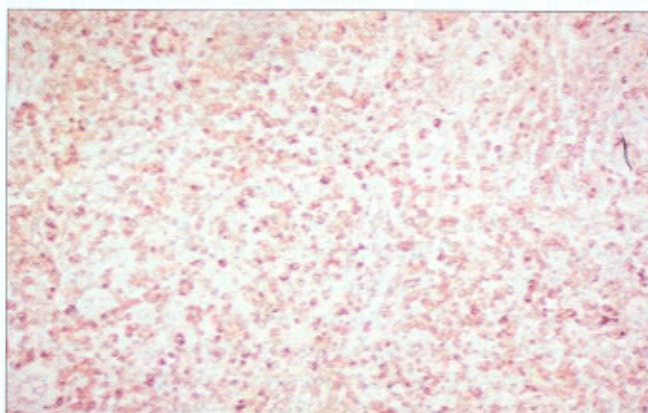


Figure 5  
Abundant number of B-lymphocytes and  
plasma cells in the granulomatous zone of periapical granuloma  
Immunohistochemical reaction  
(ABC method) using mixed polyclonal anti-heavy chain  
(anti-IgA+IgG+IgM) antibody (X160) [56]

lin synthesis, which also may contribute to the eradication of residual periradicular infection (Fig. 5). The functional significance of local humoral immunity in limiting advanced periapical lesions is emphasized by the observation that B-cell-deficient mice – but not T-cell- or complement-deficient mice – were at risk to develop disseminated anaerobic infections following experimental root canal infection [26].

With the exception of true radicular cysts, the proportion of which is not exactly characterized, the ma-

jority of periapical lesions improve significantly by conventional endodontics [37, 76]. The lesion may heal either completely by remineralization, as indicated by the disappearance of the former radiolucent area, or it may develop into fibrotic periapical granuloma, an inert periapical scar tissue.

### Concluding remarks

Recent advances in the understanding of the network of interactions of endodontopathic microbes, immune-inflammatory cells of the host and their soluble products have changed our concept on apical periodontitis. Periapical inflammation develops usually in response to root canal infection. A cascade of bacterial irritation, induction of pro-inflammatory cytokine production and cell adhesion molecule expression by a few resident cells of the narrow periradicular space recruit an extensive number of immune-inflammatory cells which form the highly organized structures of chronic periapical lesions. The granulation tissue exerts a protective function on the one hand by preventing the invasion of the periapical area by root canal microbes. However, it is the same granulation tissue that causes degenerative changes, most importantly the resorption of hard tissues. Extensive local inflammation may impair general health, as well. This chain of events, leading ultimately to the loss of the affected tooth, can be disrupted by disinfection of the root canal using proper endodontic treatment. Surgical intervention, such as apicoectomy and periapical curettage is necessary only in selected cases, since the granulation tissue has the capacity of healing, once constant irritation from the root canal has been eliminated. This dynamic equilibrium of protective function, degenerative and regenerative changes is regulated by the presence and proper function of host cells and regulatory molecules. Inhibiting leukocyte recruitment or activation result in a widespread local bacterial infiltration, even in generalized infections. Neither can remineralization of the lost hard tissue occur without the active functioning of the granulation tissue. Recent advances in the understanding of the pathomechanism of the periapical inflammatory processes resulted in a change in the treatment strategy of this disease. This paper may provide the practicing dentists a theoretical background to choose the proper treatment for their patients.

### Acknowledgement

Microphotographs were prepared in collaboration with Drs. Balázs Dezső, Ildiko J. Marton, Zoltan Nemes and Antal Rot. The paper was prepared by the support of the grant of the National Scientific Research Fund "OTKA" No T 046588.



## References

1. BARDI E, JENEI C, KISS C: Polymorphism of angiotensin converting enzyme is associated with severe circulatory compromise in febrile neutropenic children with cancer. *Pediatr Blood Cancer* 2005; 45: 217–221.
2. BARKHORDAR RA, HAYASHI C, HUSSAIN MZ: Detection of interleukin-6 in human dental pulp and periapical lesions. *Endodont Dental Traumatol* 1999; 15: 26–27.
3. BECK J, GARCIA R, HEISS G, VOKONAS PS, OFFENBACHER S: Periodontal disease and cardiovascular disease. *J Periodontol* 1996; 67: 1123–1137.
4. BEER R, BAUMANN MA, Kim S: Endodontology. In: RATEITSCHAK KH, WOLF HF (eds.): *Color Atlas of Dental Medicine*. Thieme, Stuttgart, 2000; 26–34.
5. BJURHOLM A, KREICBERGS A, BRODIN E, SCHULTZBERG M: Substance P- and CGRP-immunoreactive nerves in bone. *Peptides* 1988; 9: 165–171.
6. BRAMANTI TE, WONG GG, WEINTRAUB ST, HOLT SC: Chemical characterization and biological properties of lipopolysaccharide from *Bacteroides gingivalis* strains W50, W83 and ATCC 33277. *Oral Microbiol Immunol* 1989; 4: 183–192.
7. CARLOS TM, HARLAN JM: Leukocyte-endothelial adhesion molecules. *Blood* 1994; 84: 2068–2101.
8. CAVET J, DICKINSON AM, NORDEN J, TAYLOR PR, JACKSON GH, MIDDLETON PG: Interferon-gamma and interleukin-6 gene polymorphisms associate with graft-versus-host disease in HLA-matched sibling bone marrow transplantation. *Blood* 2001; 98: 1594–1600.
9. CHEN CP, HERTZBERG M, JIANG YL, GRAVES DT: Interleukin-1 and tumor necrosis factor receptor signaling is not required for bacteria-induced osteoclastogenesis and bone loss but is essential for protecting the host from a mixed anaerobic infection. *Am J Pathol* 1999; 155: 2145–2152.
10. COTTI E, TORABINEJAD M: Detection of leukotriene C<sub>4</sub> in human periradicular lesions. *Int Endod J* 1994; 27: 82–86.
11. DARVEAU RP, BELTON CM, REIFE RA, LAMONT RJ: Local chemokine paralysis, a novel pathogenic mechanism for *Porphyromonas gingivalis*. *Infect Immun* 1998; 66: 1660–1665.
12. DEBELIAN GJ, OLSEN I, TRONSTAD L: Electrophoresis of whole-cell soluble proteins of microorganisms isolated from bacteremias in endodontic therapy. *Eur J Oral Sci* 1996; 104: 540–546.
13. DESHPANDE RG, KHAN MB, GENCO CA: Invasion of aortic and heart endothelial cells by *Porphyromonas gingivalis*. *Infect Immun* 1998; 66: 5337–5343.
14. EL-SWIAH JM, WALKER RT: Reasons for apicectomies. A retrospective study. *Endod Dent Traumatol* 1996; 12: 185–191.
15. FUKAGAWA M, KAZAMA JJ, KUROKAWA K: Renal osteodystrophy and secondary hyperparathyroidism. *Nephrol Dial Transplant* 2002; 17: 2–5.
16. GRAU AJ, BUGGLE F, ZIEGLER C, SCHWARZ W, MEUSER J, TASMAN AJ ET AL: Association between acute cerebrovascular ischemia and chronic and recurrent infection. *Stroke* 1997; 28: 1724–1729.
17. HAMA S, TAKEICHI O, HAYASHI M, KOMIYAMA K, ITO K: Co-production of vascular endothelial cadherin and inducible nitric oxide synthase by endothelial cells in periapical granuloma. *Int Endod J* 2005; 39: 179–184.
18. HAMA S, TAKEICHI O, SAITO I, ITO K: Involvement of inducible nitric oxide synthase and receptor for advanced glycation end products in periapical granulomas. *J Endod* 2007; 33: 137–141.
19. HAN DC, HUANG GTJ, LIN LM, WARNER NA, GIM JS, JEWETT A: Expression of MHC class II, CD70, CD80, CD68 and pro-inflammatory cytokines is differentially regulated in oral epithelial cells following bacterial challenge. *Oral Microbiol Immunol* 2003; 18: 350–358.
20. HANAZAWA S, MURAKAMI Y, HIROSE K: *Bacteroides* (*Porphyromonas*) *gingivalis* fimbriae activate mouse peritoneal macrophages and induce gene expression and production of interleukin-1. *Infect Immun* 1991; 59: 1972–1977.
21. HANAZAWA S, MURAKAMI Y, TAKESHITA A, KITAMI H, OHTA K, AMANO S ET AL: *Porphyromonas gingivalis* fimbriae induce expression of the neutrophil chemotactic factor KC gene of mouse peritoneal macrophages; role of protein kinase C. *Infect Immun* 1992; 60: 1544–1549.
22. HAPPONEN RP, BERGENHOLTZ G: Apical periodontitis. In: BERGENHOLTZ G, HORSTED-BINDSLEV P, REIT C (ed.): *Textbook of Endodontology*. Blackwell, Oxford, 2003; 130–144.
23. HARASZTHY VI, ZAMBON JJ, TREVISAN M, SHAH R, ZEID M, GENCO RJ: Identification of pathogens in atheromatous plaques. *J Dent Res* 1997; 76: 666–666.
24. HARGREAVES KM, SWIFT JQ, ROSZKOWSKI MT, BOWLES W, GARRY MG, JACKSON DL: Pharmacology of peripheral neuropeptide and inflammatory mediator release. *Oral Surg Oral Med Oral Pathol Oral Radiol Endod* 1994; 78: 503–510.
25. HERNICHEL-GORBACH E, KORNMAN K, HOLE SC: Host responses in patients with generalized refractory periodontitis. *J Periodontol* 1994; 65: 8–16.
26. HOU L, SASAKI H, STASHENKO P: B-cell deficiency predisposes mice to disseminating anaerobic infections: Protection by passive antibody transfer. *Infect Immun* 2000; 68: 5645–5651.
27. HOU L, SASAKI H, STASHENKO P: Toll-like receptor 4-deficient mice have reduced bone destruction following mixed anaerobic infection. *Infect Immun* 2000; 68: 4681–4687.
28. HUANG GT, KIM D, LEE JK, KURAMITSU HK, HAAKE SK: Interleukin-8 and intercellular adhesion molecule 1 regulation in oral epithelial cells by selected periodontal bacteria: multiple effects of *Porphyromonas gingivalis* via antagonistic mechanisms. *Infect Immun* 2001; 69: 1364–1372.
29. HUANG GT, KINDER HAAKE S, KIM JW, PARK NH: Differential expression of interleukin-8 and intercellular adhesion molecule-1 by human gingival epithelial cells in response to *Actinobacillus actinomycetem-comitans* or *Porphyromonas gingivalis* infection. *Oral Microbiol Immunol* 1998; 13: 301–309.
30. HUANG GTJ, DO M, WINGARD M, PARK JS, CHUGAL N: Effect of interleukin-6 deficiency on the formation of periapical lesions after pulp exposure in mice. *Oral Surg Oral Med Oral Pathol Oral Radiol Endod* 2001; 92: 83–88.
31. KABASHIMA H, NAGATA K, MAEDA K, IJIMA T: Interferon- $\gamma$ -producing cells and inducible nitric oxide synthase-producing cells in periapical granulomas. *J Oral Pathol Med* 1998; 27: 95–100.
32. KABASHIMA H, NAGATA K, MAEDA K, IJIMA T: Involvement of substance P, mast cells, TNF- $\alpha$  and ICAM-1 in the infiltratory cells in human periapical granulomas. *J Oral Pathol Med* 2002; 31: 175–180.
33. KANEKO T, OKIJI T, KAN L, TAKAGI M, SUDA H: Ultrastructural analysis of MHC Class II molecule-expressing cells in experimentally induced periapical lesions in the rat. *J Endod* 2001; 27: 337–342.
34. KAWAHARA T, MURAKAMI S, NOIRI Y, EHARA A, TAKEMURA N, FURUKAWA S ET AL: Effects of cyclosporin-A-induced immunosuppression on periapical lesions in rat. *J Dent Res* 2004; 83: 683–687.
35. KAWASHIMA N, STASHENKO P: Expression of bone-resorptive and regulatory cytokines in murine periapical inflammation. *Arch Oral Biol* 1999; 44: 55–66.
36. KAWASHIMA N, SUZUKI N, YANG G, OHI C, OKUHARA S, NAKANO-KAWANISHI H ET AL: Kinetics of RANKL, RANK and OPG expressions in experimentally induced rat periapical lesions. *Oral Surg Oral Med Oral Pathol Oral Radiol Endod* 2007; 103: 707–711.
37. KERESKES K, TRONSTAD L: Long-term results of endodontic treatment performed with standardized techniques. *J Endod* 1979; 5: 83–90.
38. KIRBY AC, MEGHI S, NAIR S, WHITE P, REDDI K, NISHIHARA T ET AL: The potent bone resorbing mediator of *Actinobacillus actinomycetem-comitans* is homologous to the molecular chaperone GroEL. *J Clin Invest* 1996; 96: 1185–1194.
39. KISS C, BENKÖI, KOVÁCS P: Leukemic cells and the cytokine patchwork. *Pediatr Blood Cancer* 2004; 42: 113–121.
40. KISS C, CESANO A, ZSEBŐ KM, CLARK SC, SANTOLI D: Human stem cell factor (c-kit ligand) induces an autocrine loop of growth in a GM-CSF-dependent megakaryocytic leukemia cell line. *Leukemia* 1993; 7: 235–240.
41. KISS C, SURREY S, SCHREIBER AD, SCHWARTZ E, MCKENZIE SE: Human c-kit ligand (stem cell factor) induces platelet Fc receptor expression in megakaryoblastic cells. *Exp Hematol* 1996; 24: 1232–1237.
42. KISS C: Cell-to-cell interactions. *Endod Topics* 2004; 8: 88–103.



43. KOSUGI M, ISHIIHARA K, OKUDA K: Implication of responses to bacterial heat shock proteins, chronic microbial infections, and dental metal allergy in patients with pustulosis palmaris et plantaris. *Bull Tokyo Dent Coll* 2003; 44: 149–158.
44. KOTAKE S, UDAGAWA N, TAKAHASHI N, MATSUZAKI K, ITOH K, ISHIYAMA S ET AL: IL-17 in synovial fluids from patients with rheumatoid arthritis is a potent stimulator of osteoclastogenesis. *J Clin Invest* 1999; 103: 1345–1352.
45. KRONFELD R: *Histomorphology of the teeth and their surrounding structures*. 2<sup>nd</sup> ed. Lea and Febiger, Philadelphia 1939; 208.
46. LEONARDI L, CALTABIANO M, PAGANO M, PEZZUTO V, LORETO C, PALESTRO G: Detection of vascular endothelial growth factor/vascular permeability factor in periapical lesions. *J Endod* 2003; 29: 180–183.
47. LIM GC, TORABINEJAD M, KETTERING J, LINKHARDT TA, FINKELMAN RD: Interleukin 1-beta in symptomatic and asymptomatic human periapical lesions. *J Endod* 1994; 20: 225–227.
48. LIU F, ABIKO Y, NISHIMURA M, KUSANO K, SHI S, KAKU T: Expression of inflammatory cytokines and beta-defensin 1 mRNAs in porcine epithelial rests of Malassez *in vitro*. *Med Electron Microsc* 2001; 34: 174–178.
49. LOESCHE WJ, GUSBERTI F, METTRAUX G, HIGGINS T, SYED S: Relationship between oxygen tension and subgingival bacterial flora in untreated human periodontal pockets. *Infect Immunol* 1983; 42: 659–667.
50. LOMCALI G, SEN BH, CANKAYA H: Scanning electron microscopic observations of apical root surfaces of teeth with apical periodontitis. *Endod Dent Traumatol* 1996; 12: 70–76.
51. LOTZ M, VAUGHAN JH, CARSON DA: Effect of neuropeptides on production of inflammatory cytokines by human monocytes. *Science* 1988; 241: 1218–1221.
52. LUKIC A, VASILJIC S, MAJSTOROVIC I, VUCEVIC D, MOJISLOVIC S, GAZIVODA D ET AL: Characterization of antigen-presenting cells in human apical periodontitis lesion by flow cytometry and immunohistochemistry. *Int Endod J* 2006; 39: 626–636.
53. LUSTER AD: Chemokines – chemotactic cytokines that mediate inflammation. *N Engl J Med* 1998; 338: 436–445.
54. MACKENZIE RS, MILLARD HD: Interrelated effects of diabetes, arteriosclerosis and calculus on alveolar bone loss. *J Am Dent Assoc* 1963; 66: 192–198.
55. MARTON I, KISS C, BALLA G, SZABO T, KARMAZSIN L: Acute phase proteins in patients with chronic periapical granuloma before and after surgical treatment. *Oral Microbiol Immunol* 1988; 3: 95–96.
56. MARTON I, NEMES Z, HARMATI S: Quantitative Significance of IgE producing Plasma Cells and Tissue Distribution of Mast Cells in Apical Periodontitis. *Oral Microbiol Immunol* 1990; 5: 46–48.
57. MARTON I, RÉDAI I, KELENTÉY B, LÁZÁR SZ, ÖLVETI É: Granuloma periapikális fogak gyökércsatornabennékeinek mikrobiológiai vizsgálata. *Fogorv Szle* 1988; 81: 245–248.
58. MARTON IJ, DEZSŐ B, RADICS T, KISS C: Distribution of interleukin-2 receptor  $\alpha$ -chain and cells expressing major histocompatibility complex class II antigen in chronic human periapical lesions. *Oral Microbiol Immunol* 1998; 13: 259–262.
59. MARTON IJ, KISS C: Characterization of inflammatory cell infiltrate in dental periapical lesions. *Int Endod J* 1993; 26: 131–136.
60. MARTON IJ, KISS C: Influence of surgical treatment of periapical lesions on serum and blood levels of inflammatory mediators. *Int Endod J* 1992; 25: 229–233.
61. MARTON IJ, KISS C: Protective and destructive immune reactions in apical periodontitis. *Oral Microbiol Immunol* 2000; 15: 139–150.
62. MARTON IJ, ROT A, SCHWARZINGER E, SZAKÁLL S, RADICS T, VÁLYI-NAGY I ET AL: Differential *in situ* distribution of interleukin-8, monocyte chemoattractant protein-1 and Rantes in human chronic periapical granuloma. *Oral Microbiol Immunol* 2000; 15: 63–65.
63. MATTILA K, NIEMINEN M, VALTONEN V, RASI VP, KESANIEMI YA, SYRJALA SL ET AL: Association between dental health and acute myocardial infarction. *Br Med J* 1989; 298: 779–782.
64. MATTILA K, VALLE MS, NIEMINEN MS, VALTONEN VV, HIETANIEMI KL: Dental infections and coronary atherosclerosis. *Atherosclerosis* 1993; 103: 205–211.
65. MCNICHOLAS S, TORABINEJAD M, BLANKENSHIP J, BAKLAND L: The concentration of prostaglandin E<sub>2</sub> in human periradicular lesions. *J Endod* 1991; 17: 97–100.
66. MIZGERD JP: Molecular mechanisms of neutrophil recruitment elicited by bacteria in the lungs. *Semin Immunol* 2002; 14: 123–132.
67. MORSE DR: Immunologic aspects of pulpal-periapical diseases. A review. *Oral Surg Oral Med Oral Pathol Oral Radiol Endod* 1977; 43: 436–451.
68. MOSMANN TR, COFFMAN RL: TH1 and TH2 cells: different pattern of lymphokine secretion leads to different functional properties. *Annu Rev Immunol* 1989; 7: 145–173.
69. MULLER-EBERHARD HJ: Complement. *Annu Rev Biochem* 1975; 44: 697–724.
70. NAGAI M, SATO N: Reciprocal gene expression of osteogenesis inhibitory factor and osteoclast differentiation factor regulates osteoclast formation. *Biochem Biophys Res Commun* 1999; 257: 719–723.
71. NAIR PNR, PAJAROLA G, SCHROEDER HE: Types and incidence of human periapical lesions obtained with extracted teeth. *Oral Surg Oral Med Oral Pathol Oral Radiol Endod* 1996; 81: 93–102.
72. NAIR PNR: Apical periodontitis: a dynamic encounter between root canal infection and host response. *Periodontology* 2000 1997; 13: 121–148.
73. NAIR PNR: Light and electron microscopic studies of root canal flora and periapical lesions. *J Endod* 1987; 13: 29–39.
74. NAIR PNR: New perspectives on radicular cysts: Do they heal? *Int Endod J* 1998; 31: 155–160.
75. OGAWA T, KUSUMOTO Y, UCHIDA H, NAGASHIMA S, OGO H, HAMADA S: Immunobiological activities of synthetic peptide segments of fimbrial protein from *Porphyromonas gingivalis*. *Biochem Biophys Res Commun* 1991; 180: 1335–1341.
76. ORSTAVIK D: Time-course and risk analyses of the development and healing of chronic apical periodontitis in man. *Int Endod J* 1996; 29: 150–155.
77. PEARSON TA, MENSAH GA, ALEXANDER RW, ANDERSON JL, CANNON RO 3RD, CRICQUI M ET AL: Markers of inflammation and cardiovascular disease: application to clinical and public health practice: a statement from healthcare professionals from the Centers for Disease Control and Prevention and the American Heart Association. *Circulation* 2003; 107: 499–511.
78. PULITI M, VON HUNOLSTEIN C, BISTONI F, CASTRONARI R, OREFICI G, TISSI L: Role of macrophages in experimental group B streptococcal arthritis. *Cell Microbiol* 2002; 4: 691–699.
79. RADICS T, KISS C, TAR I, MARTON IJ: Interleukin-6 and granulocyte-macrophage colony-stimulating factor in apical periodontitis: correlation with clinical and histologic findings of the involved teeth. *Oral Microbiol Immunol* 2003; 18: 9–13.
80. REDDI K, WILSON M, NAIR S, POOLE S, HENDERSON B: Comparison of the pro-inflammatory cytokine-stimulating activity of the surface-associated proteins of periodontopathic bacteria. *J Periodontol Res* 1996; 31: 120–130.
81. REINHERZ EL, SCHLOSSMAN SF: The differentiation and function of human T lymphocytes. *Cell* 1980; 19: 821–827.
82. RICUCCI D, PASCON EA, PITT FORD TR, LANGELAND K: Epithelium and bacteria in periapical lesions. *Oral Surg Oral Med Oral Pathol Oral Radiol Endod* 2006; 101: 239–249.
83. RODINI CO, LARA VS: Study of the expression of CD68+ macrophages and CD8+ T cells in human granulomas and periapical cysts. *Oral Surg Oral Med Oral Pathol Oral Radiol Endod* 2001; 92: 221–227.
84. ROTSTEIN I, SIMON JHS: Diagnosis, prognosis and decision-making in the treatment of combined periodontal-endodontic lesions. *Periodontology* 2000 2004; 34: 165–203.
85. SABETI M, SIMON J, KERMANI V, VALLES Y, ROSTEIN I: Detection of receptor activator of NF-kappa beta ligand in apical periodontitis. *J Endod* 2005; 31: 17–18.
86. SABETI M, SIMON JH, SLOTS J: Cytomegalovirus and Epstein-Barr virus are associated with symptomatic periapical pathosis. *Oral Microbiol Immunol* 2003; 18: 327–328.
87. SABETI M, VALLES Y, NOWZARI H, SIMON JH, KERMANI-ARAB V, SLOTS J: Cytomegalovirus and Epstein-Barr virus DNA transcription in end-



- odontic symptomatic lesions. *Oral Microbiol Immunol* 2003; 18: 104–108.
88. SAKELLARIOU PL: Periapical actinomycosis: report of a case and review of the literature. *Endod Dent Traumatol* 1996; 12: 151–154.
89. SCANNAPIECO FA, GENCO RJ: Association of periodontal infections with atherosclerotic and pulmonary diseases. *J Periodont Res* 1999; 34: 340–345.
90. SCANNAPIECO FA: An update on periodontal medicine: associations between periodontal disease and atherosclerosis, lung disease and adverse pregnancy outcome. *Real Clin* 2003; 14: 303–316.
91. SCOTT P, PEARCE E, CHEEVER AW, COFFMAN RL, SHER A: Role of cytokines and CD4<sup>+</sup> T-cell subsets in the regulation of parasite immunity and disease. *Immunol Rev* 1989; 112: 161–182.
92. SELTZER S, SOLTANOFF W, BENDER IB: Epithelial proliferation in periapical lesions. *Oral Surg Oral Med Oral Pathol Oral Radiol Endod* 1969; 27: 110–121.
93. SLOTS J, SABETI M, SIMON JH: Herpesviruses in periapical pathosis: An etiopathogenic relationship? *Oral Surg Oral Med Oral Pathol Oral Radiol Endod* 2003; 96: 327–331.
94. SONODA T, ANAN T, ONO K, YANAGISAWA S: Chronic urticaria associated with dental infection. *Br J Dermatol* 2001; 145: 516–518.
95. STASHENKO P, WANG CY, TANI-ISHII N, YU SM: Pathogenesis of induced rat periapical lesions. *Oral Surg Oral Med Oral Pathol Oral Radiol Endod* 1994; 78: 494–502.
96. STASHENKO P, YU SM: T helper and T suppressor cell reversal during the development of induced rat periapical lesions. *J Dent Res* 1989; 68: 830–834.
97. SUDA T, TAKAHASHI N, UDAGAWA N, JIMI E, GILLESPIE MT, MARTIN TJ: Modulation of osteoclast differentiation and function by new members of the tumor necrosis factor receptor and ligand families. *Endocr Rev* 1999; 20: 345–357.
98. SUNDQUIST G: Taxonomy, ecology and pathogenicity of the root canal flora. *Oral Surg Oral Med Oral Pathol Oral Radiol Endod* 1994; 78: 522–530.
99. SUZUKI N, OKIJI T, SUDA H: Enhanced expression of activation-associated molecules on macrophages of heterogeneous populations in expanding periapical lesions in rat molars. *Arch Oral Biol* 1999; 44: 67–79.
100. SYRJANEN J, PELTOLA J, VALTONEN V, IIVANAINEN M, KASTE M, HUTTUNEN JK: Dental infections in association with cerebral infarctions in young and middle-aged men. *J Intern Med* 1989; 225: 179–184.
101. TAKAHASHI K, MACDONALD DG, MURAYAMA Y, KINANE DF: Cell synthesis, proliferation and apoptosis in human dental periapical lesions analysed by in situ hybridisation and immunohistochemistry. *Oral Dis* 1999; 5: 313–320.
102. WALKER KF, LAPPIN DF, TAKAHASHI K, HOPE J, MACDONALD DG, KINANE DF: Cytokine expression in periapical granulation tissue as assessed by immunohistochemistry. *Eur J Oral Sci* 2000; 108: 195–201.
103. WANG CY, STASHENKO P: Characterization of bone-resorbing activity in human periapical lesions. *J Endod* 1993; 19: 107–111.
104. WILSON M, REDDI K, HENDERSON B: Cytokine-inducing components of periodontopathogenic bacteria. *J Periodont Res* 1996; 31: 393–407.
105. YANAGISAWA S: Pathologic study of periapical lesions. I. Periapical granulomas: clinical, histopathologic and immunohistopathologic studies. *J Oral Pathol* 1980; 9: 288–300.
106. YANG G, KAWASHIMA N, KANEKO T, SUZUKI N, OKIJI T, SUDA H: Kinetic study of immunohistochemical colocalization of antigen-presenting cells and nerve fibers in rat periapical lesions. *J Endod* 2007; 33: 132–136.
107. YLDIRIM S, YAPAR M, KUBAR A, SLOTS H: Human cytomegalovirus, Epstein-Barr virus and bone resorption-inducing cytokines in periapical lesions of deciduous teeth. *Oral Microbiol Immunol* 2006; 21: 107–111.