ARTICLE IN PRESS

Carbohydrate Research xxx (2008) xxx-xxx

Contents lists available at ScienceDirect

Carbohydrate Research

journal homepage: www.elsevier.com/locate/carres



Synthesis of anomeric sulfonamides and their behaviour under radical-mediated bromination conditions

Katalin Czifrák, László Somsák*

Department of Organic Chemistry, University of Debrecen, PO Box 20, H-4010 Debrecen, Hungary

ARTICLE INFO

Article history: Received 10 October 2008 Received in revised form 5 November 2008 Accepted 5 November 2008 Available online xxxx

Keywords: Sulfonamide Glycopyranosyl Bromination Radical

ABSTRACT

Q-Peracetylated methyl 3-(p-glycopyranosylthio)propanoates of β -p-gluco, and α - and β -p-galacto configurations were oxidized to the corresponding S-dioxides (sulfones) by Oxone® or MCPBA. Oxidation of the β -p-gluco derivative with H₂O₂/Na₂WO₄ gave the corresponding S-oxide (sulfoxide). DBU-induced elimination of methyl acrylate from the β -p-gluco and β -p-galacto configured S-dioxides (sulfones) gave Q-peracetylated β -p-glycopyranosyl-1-C-sulfinates which, on treatment with H₂NOSO₃H, furnished the corresponding β -p-glycopyranosyl-1-C-sulfinates whick, on treatment of the protected methyl 3-(β -p-glycopyranosyl-1-C-sulfonamides. Radical-mediated bromination of the protected methyl 3-(β -p-glycopyranosyl bromides of the Q-peracetylated β -p-glycopyranosyl-1-C-sulfonamides. Radical-mediated bromination of the protected in the formation of α -p-glycopyranosyl bromides and 1-C- and 5-C-bromoglycosyl compounds. Similar brominations of the Q-peracetylated β -p-glucopyranosyl-1-C-sulfonamides. A rationale for these observations was proposed. Methyl 3-(β -p-glucopyranosyl sulfonamides. A ratio-glucopyranosyl-1-C-sulfonamide proved inefficient when tested as inhibitors of rabbit muscle glycogen phosphorylase b.

© 2008 Elsevier Ltd. All rights reserved.

1. Introduction

As part of an ongoing project to design and synthesize glycoenzyme inhibitors, we have prepared several compounds¹⁻⁴ with glycogen phosphorylase (GP) inhibitory activity^{5.6} from (3,4,5,7-tetra-*O*-acyl- α -*D*-glyco-hept-2-ulopyranosyl bromide)onamides (**A**, Chart 1). Among them, compound **C** was shown to have moderate inhibition against rabbit muscle GPb (RMGPb).⁴ Since GP is a validated target for the treatment of type 2 diabetes mellitus many efforts

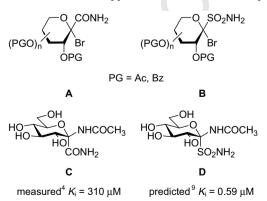


Chart 1.

* Corresponding author. Tel.: +36 52 512 900/22348; fax: +36 52 453 836. *E-mail address:* somsak@tigris.unideb.hu (L. Somsák).

0008-6215/\$ - see front matter © 2008 Elsevier Ltd. All rights reserved. doi:10.1016/j.carres.2008.11.002

are devoted to finding new inhibitors (GPIs).^{7.8} To this end, among others, computational chemistry is in the forefront of GPI design.⁸ By using 4D-QSAR methods, it was predicted that compound **D**, a sulfonyl analogue of **C**, might be a very efficient GPI with an inhibition constant in the nanomolar range.⁹ Therefore, we envisaged to prepare **D** from **B** in a way analogous to the chemistry² applied to obtain **A** and its further transformations to **C**.

To the best of our knowledge, the necessary starting materials to get **A**, that is, N-unsubstituted anomeric sulfonamides, have not yet been described. The only analogous compound is the *N*,*N*-diethylsulfonamide in the α -anomeric position of *N*-acetyl-pglucosamine obtained by substituting Et₂NH in the corresponding glycosylsulfenyl chloride or bromide and subsequent oxidation.¹⁰

In this paper we disclose our experiences in the preparation of glycopyranosyl sulfonamides and their reactivity under radicalmediated bromination conditions.

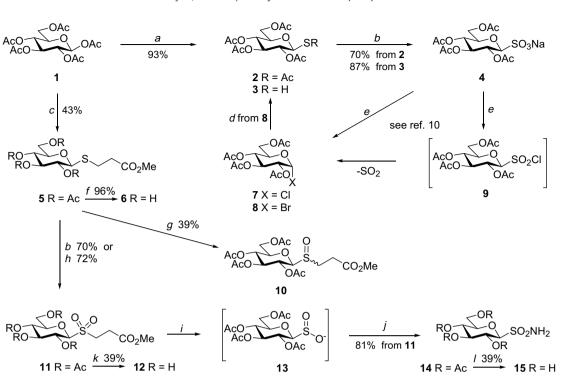
2. Results and discussion

A generally used method for the preparation of sulfonamides is the ammonolysis of the corresponding sulfonylchlorides. At the commencement of our work, glycosyl sulfonyl chlorides (e.g., 9) were not known, therefore chlorination of sugar sulfonate salt 4 was envisaged. Several sulfonic acids and their salts attached to a sugar skeleton were described.^{11–13,†} The general method for

[†] For a more detailed list of citations see Ref. 23 in a paper by Knapp et al.¹⁰

90

100



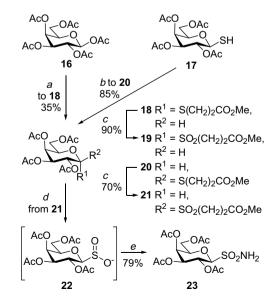
Scheme 1. Reagents and conditions: (a) AcSH, BF₃·OEt₂, dry CH₂Cl₂, 0–20 °C; (b) Oxone[®], NaOAc, AcOH, rt; (c) HSCH₂CH₂CO₂Me, BF₃·OEt₂, CH₂Cl₂, 0–20 °C; (d) (1) H₂NCSNH₂, dry acetone, reflux; (2) NaHSO₃, CHCl₃–H₂O, reflux; (e) SOCl₂, or PCl₅, dry THF or dry Et₂O, reflux; (f) NaOMe, dry MeOH, rt; (g) Na₂WO₄, H₂O₂, MeOH, rt; (h) *m*CPBA, NaHCO₃, CH₂Cl₂, 0–20 °C; (i) DBU, CHCl₃, rt; (j) H₂NOSO₃H, water, rt; (k) AcCl, dry MeOH rt; (l) NH₄OH, MeOH, rt.

obtaining such compounds was a nucleophilic displacement by AcSK in the corresponding sugar derivative followed by oxidation,¹³ and it was also extended to the anomeric position.¹⁴ Thus, to get sodium sulfonate **4** (Scheme 1), either O-peracetylated 1-S-acetyl-1-thio-β-**D-glucopyranose**¹⁵ (**2**) prepared from 1,2,3,4,6-penta-O-acetyl- β -Dglucopyranose (1) and thiolacetic acid¹⁶ or O-peracetylated 1-thio- β -p-glucopyranose (3) obtained by the conventional literature method^{17,18} from **8** was oxidized by Oxone[®].¹⁴ The corresponding triethylammonium salt¹⁰ was made from **2** by DMDO oxidation followed by chromatography in the presence of Et₃N. Reactions of **4** under usual chlorination conditions (neat SOCl₂, reflux; THF, SOCl₂; THF-Et₂O, PCl₅) gave several compounds which were not separated. Experiments to add nucleophiles to the crude chlorination mixtures with the hope of trapping any sulfonyl chloride formed were unsuccessful. Knapp et al. isolated the corresponding glycosyl chloride 7 (70%) from the chlorination mixture of their triethylammonium sulfonate,¹⁰ and accounted for the formation of **7** by SO₂ extrusion from the unstable intermediate sulfonyl chloride 9. For these reasons other possibilities were sought for the preparation of the target sulfonamides.

Various sulfonamides were prepared by reactions of sulfinate salts with electrophilic nitrogen sources such as hydroxylamine-O-sulfonic acid^{19,20} or bis(2,2,2-trichloroethyl)azodicarboxylate.²¹ The preparation of the required anomeric sulfinate **13** was effected by base-induced β -elimination from sulfon **11** which was obtained by oxidation of thioglucoside **5**^{22,23} by either Oxone[®] or *mCPBA*. Application of H₂O₂/Na₂WO4^{20,21} to oxidize **5** gave sulfoxides **10** (two TLC spots of very similar mobility) from which one isomer could be isolated in pure state in moderate yield. As an alternative route to sulfinates, the reductive cleavage of benzothiazolyl sulfones was proposed;²¹ however, practically no change could be observed in the reaction of O-peracetylated β -p-glucopyranosyl benzothiazol-2-yl sulfone²⁴ with NaBH₄ in THF/MeOH for ~one week even at elevated temperature. Sulfinate **13**, without being purified, was treated with H_2NOSO_3H in water to give the desired sulfonamide **14** after chromatographic purification in <u>81%</u> yield for the two steps.

For the preparation of the p-galacto configurated derivative **23** (Scheme 2), the corresponding thioglycoside had to be prepared first. BF₃ catalyzed reaction of 1,2,3,4,6-penta-O-acetyl- β -D-galactopyranose (**16**) with methyl 3-sulfanylpropanoate gave the α -thiogalactoside **18** in moderate yield. To get the β -anomer, 2,3,4,6-tetra-O-acetyl-1-thio- β -D-galactopyranose^{25,26} (**17**) was

110



Scheme 2. Reagents and conditions: (a) $HSCH_2CH_2CO_2Me$, $BF_3 \cdot OEt_2$, CH_2CI_2 , 0-20 °C; (b) $BrCH_2CH_2CO_2Me$, Et_3N , CH_2CI_2 , rt; (c) $Oxone^{\circledast}$, NaOAc, AcOH, rt; (d) DBU, $CHCI_3$, rt; (e) H_2NSO_3H , water, rt.

ARTICLE IN PRESS

3

K. Czifrák, L. Somsák/Carbohydrate Research xxx (2008) xxx-xxx

Table 1

Radical-mediated bromination of sulfonyl derivatives

12 November 2008 Disk Used

	R^{1} Q Q R^{3} Q Q R^{3} Q Q R^{3}	bromination conditions <i>a-c</i>	R ¹ OAc AcO AcO R ³	+	R^{1} OAc R^{3} R^{3} R^{3} R^{3} R^{3}	$+ \begin{array}{c} R^2 \\ ACO \\ ACO \\ ACO \\ Br \end{array} $	
		R ¹	R ²			Isolated yields (product ratios) (%)	
11	$R^3 = SO_2(CH_2)_2CO_2Me$	OAc	Н	a^1	24 21	25 30	_
21		Н	OAc	a ²	26 43	27 27	_
14	$R^3 = SO_2NH_2$	OAc	Н	b^3	28 11 (22)	29 12 (17)	8 24 (26)
				b^4	(45)	(36)	(6)
				c ⁵	28 12	29 11	8 19
23		Н	OAc	b^6	30 (12)	31 (6)	32 (59)
Description							

Bromination conditions:

(a) Br₂, CCl₄, K₂CO₃, hv, reflux.²⁷

(b) Br_2 , CHCl₃, K_2CO_3 , hv, reflux.²

(*c*) Br₂, PhCF₃, K₂CO₃, *hv*, reflux.³⁰

¹ Reaction time 3 h.

² Reaction time 2 h.

120

130

140

³ Conversion 65% after 8 h with 0.6 g starting material.

⁴ Conversion 87% after 4 h with 0.1 g starting material. Product ratio calculated from the ¹H NMR spectrum of the crude mixture.

⁵ Conversion 81% after 4 h.

⁶ Conversion 77% after 4 h. Product ratio calculated from the ¹H NMR spectrum of the crude mixture,

reacted with methyl 3-bromopropanoate in the presence of Et₃N, and the required compound **20** was obtained in good yield. The anomeric configuration of these derivatives was clearly indicated by the vicinal coupling constants: ${}^{3}J_{1,2} = 5.3$ Hz for **18** and ${}^{3}J_{1,2} = 9.2$ Hz for **20**. Oxidation of thiogalactosides **18** and **20** by Oxone[®] afforded the corresponding sulfones **19** and **21**, respectively, in good yields (${}^{3}J_{1,2} = 2.6$ Hz for **19** and ${}^{3}J_{1,2} = 9.2$ Hz for **21**). Treatment of the β -anomeric sulfone **21** with DBU followed by reaction of the non-isolated sulfinate **22** with H₂NOSO₃H gave the expected sulfonamide **23** in 79% overall yield for the two steps. Similar base-induced reaction of the α -anomer **19** resulted in a mixture from which no discrete product could be isolated.

Radical-mediated bromination of various carbohydrate derivatives proved a useful tool to produce versatile compounds for further functionalization.²⁷ Several pyranoid monosaccharide substrates showed excellent regio- and stereoselectivities under the bromination conditions to give either 1-*C*-bromo- or 5-*C*-bromo derivatives of pyranoid rings with the bromine substituents in axial positions. On the other hand, a study on bromination of phenyl 2,3,4,6-tetra-*O*-acetyl-β-*D*-glucopyranosyl sulfone demonstrated similar reactivities for the 1- and 5-positions (C-1 bromide 48%, C-5 bromide 38%).²⁸

With these préliminaries in mind, it was to be expected that bromination (Table 1) of the *gluco* (11) and *galacto* (21) sulfones would give isomeric axial bromides 24 and 25, as well as 26 and 27, respectively. Bromination of sulfonamides 14 and 23 under different conditions gave varying proportions of the corresponding <u>o-</u> -glycopyranosyl bromides[‡] 8 and 32, respectively, the ratio also being dependent on the scale of the reaction. The ring brominated derivatives 28 and 29, as well as 30 and 31, respectively, were formed as minor products when the reaction was performed on a larger scale. Compounds 28 and 29 were isolated by chromatography; however, the very similar chromatographic behaviour of 30 and 31 did not allow these derivatives to be prepared in pure state. The position of the bromine substitution in the sugar ring followed clearly from the coupling pattern of the ¹H NMR spectra of these compounds.

These observations can be correlated with the relative radical stabilization (RRS) factors of substituents (Table 2) as in earlier explanations for the regioselectivity of radical-mediated bromination of sugar derivatives.²⁷ As shown in Figure 1a, the radical stabilization is highest in position 1 of the sugar ring (RRS = 8.3; for positions 2-4 the OAc substituent is characterized by an RRS value of 0.9³¹). Although the radical in position next to the CO₂Me group seems to be better stabilized than C-5 (RRS 7.9 vs 6.5), the observed higher reactivity of position 5 might be attributed to its tertiary character (as compared to the secondary carbon in the aglycon) which is not taken into consideration in the calculation of the RRS value. However, bromination in the methylene groups of the aglycon cannot be excluded, as very minor bromine-containing spots were observed by TLC in the reaction mixtures. Compounds with a carbon substituent at position one (for the present particular comparison a carboxamido group, but also other anhydro-aldonic acid derivatives) showed an exclusive selectivity for that position^{2,32,33} as it can be concluded from the RRS values²⁷ (Fig. 1b). In the bromination of sulfonamides the relative reactivity of positions 1 and 5 was similar to those of the sulfones (Fig. 1a). However, the formation of glycosyl bromides needs further considerations. We propose that in the bromination of sulfonamides (Fig. 1c) the bromine radical may also abstract a hydrogen from the SO_2NH_2 moiety as illustrated in I. Further β -fragmentation of the intermediate sulfonamidyl radical II might give glycosyl radical³⁴ III which, on bromine abstraction from the reagent, could give the glycosyl bromide IV with high stereoselectivity. The preponderance of this pathway may be attributed to the relatively free

Table 2

Selected relative radical stabilization $factors^{31}$ of substituents (RRS_X) and their combined values $(RRS_{0,X})$ at positions 1 and 5 of the pyranoid ring

Х	RRS _X	RRS _{O,X}
CH_3 (= CH_2OAc)	2.3	6.5
OMe (= O-ring)	4.5	
SO ₂ Ph	4.5	8.3
СООМе	7.9	11.3
CN	8.6	11.9
СОМе	10.2	13.2

X represents the substituent in position 1 or 5 of the pyranoid ring while O stands for the ring oxygen (its numerical value equated with that of OMe). The combined effects were calculated by using the expression:³¹ $0.03 \times \text{RRS}_{O,X} = 1 - (1 - 0.03 \times \text{RRS}_{OME})(1 - 0.03 \times \text{RRS}_X)$.

170

160

[‡] In the literature there are some examples with a similar outcome to the present one: the corresponding α -D-glycosyl bromides were formed on radical mediated bromination of O-peracetylated phenyl β -D-glucopyranosyl sulfoxide,²⁸ cellobiosyl pyperidine²⁹ and β -D-glucopyranosyl isothiocyanate,²⁹

ARTICLE IN PRESS

K. Czifrák, L. Somsák/Carbohydrate Research xxx (2008) xxx-xxx

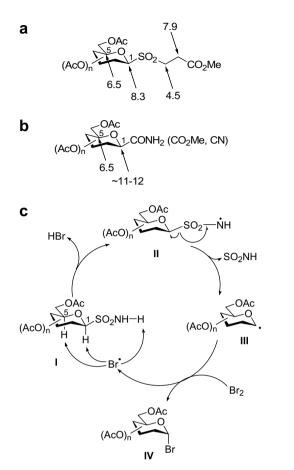


Figure 1. (a) Relative radical stabilization factors taken from Table 2 for glucopyranosyl sulfones and (b) anhydro-aldonic acid derivatives; (c) proposed reactivity pattern for the radical-mediated bromination of anomeric sulfonamides.

access to the NH₂ as compared to the ring positions, but radical stabilization factors may also play a role.

In 1-bromoglucopyranosyl sulfonamide **28** as well as sulfone **24** some nucleophilic substitutions, reactions analogous to those carried out easily^{1-3,35,36} with *O*-peracyl derivatives of (*p-glyco*-hept-2-ulopyranosyl bromide)onamides (Chart 1, **A**) were attempted. Contrary to these transformations, reactions either with azide or thiocyanate ions failed: either no change (**28** and AgSCN or NH₄SCN, CH₃NO₂, 80 °C, Ar atm; **28** and NaN₃, DMF, rt; **24** and NaN₃, acetone, LiN₃, DMF, or acetone, each at rt) or decomposition (**24** and NaN₃, DMF, rt; AgOTF, TMSN₃, CH₂Cl₂, rt) of the starting material could be observed.

190

200

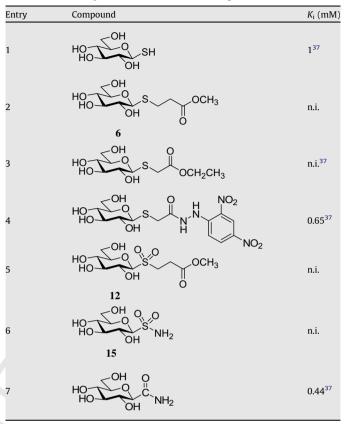
180

The prepared 1-thioglucoside **5** and glucosyl sulfonyl derivatives **11** and **14** were deprotected as shown in Scheme 1 to give compounds **6**, **12** and **15**, respectively. These compounds were tested as inhibitors of RMGP*b* (Table 3) in the way described earlier.³ While 1-thio-β-D-glucopyranose (entry 1) is a moderate inhibitor, its glycosides are either non-inhibitory (entries 2 and 3) or only slightly more efficient (entry 4). The latter effect may be due to interactions of the bulky aglycon in the so called β-channel of the enzyme.^{7,8} Sulfon **12** (entry 5) and sulfonamide **15** (entry 6) were inactive. Comparing the latter result to the effect of the 2,6-anhydro-aldonamide in entry 7 shows that introduction of the tetrahedral sulfonyl moiety in the β-anomeric position of Dglucose is not a useful modification with respect to inhibition of RMGP*b*.

In conclusion, a synthetic sequence has been elaborated for the preparation of N-unsubstituted β -D-glycopyranosyl-1-C-sulfonamides, which circumvents the unavailability of anomeric sulfonyl

Table 3

Inhibition of RMGPb by selected 1-thio derivatives of D-glucose



chlorides. Further functionalization at C-1 of these sulfonamides by radical-mediated bromination, although possible, has proven limited because of formation of the corresponding glycosyl bromides and 5-<u>C</u>-bromides as accompanying products. 1-Bromoglycosyl sulfones and sulfonamides seem to be unreactive in nucleophilic substitutions. Appending the tetrahedral sulfonyl moiety to the anomeric centre of <u>p</u>-glucose results in a loss of activity against RMGPb.

3. Experimental

3.1. General methods

Melting points were measured in open capillary tubes or on a Kofler hot-stage and are uncorrected. Optical rotations were determined on a Perkin-Elmer 241 polarimeter at room temperature. NMR spectra were recorded with Bruker WP 360 SY (360/90 MHz for ${}^{1}H/{}^{13C}$) and Varian UNITYINOVA 400 WB (400/100 MHz for ¹H/¹³C) spectrometers. Chemical shifts are referenced to Me₄Si as the internal reference (1H) or to the residual solvent signal (^{13}C) . ESIMS were recorded with a Bruker micrOTOF-Q instrument. Thin-layer chromatography (TLC) was carried out on aluminium sheets coated with Silica Gel 60 F_{254} (Merck). TLC plates were inspected by UV light ($\lambda = 254$ nm) and after gentle heating. Brominated compounds were visualized on TLC plates by spraying first a 0.1% w/v fluorescein soln in absolute MeOH, then a 1:1 mixture of H₂O₂ (30% in water) and AcOH. Upon gentle heating, bromocompounds gave pink-coloured spots. Silica gel column chromatography was performed with Silica Gel Si 60 (40-63 µm) purchased from Merck (Darmstadt, Germany). Organic solutions were dried over anhydrous MgSO₄, and concentrated at diminished pressure at 40-50 °C (water bath).

220

230

210

270

12 November 2008 Disk Used

300

310

K. Czifrák, L. Somsák/Carbohydrate Research xxx (2008) xxx-xxx

3.2. 2,3,4,6-Tetra-O-acetyl-1-*S*_,acetyl-1-thio-β-D-glucopyranose (adapted from¹⁴) (2)

1,2,3,4,6-Penta- $O_{\overline{a}}$ acetyl- β -D-glucopyranose (1, 1.00 g, 2.56) mmol) was dissolved in dry CH₂Cl₂ (10 mL). To the soln, thioacetic acid (0.28 mL, 3.84 mmol) and BF₃;OEt₂ (0.54 mL, 5.12 mmol) were added at 0 °C. The mixture was then stirred at rt for 1 d, and after completion of the reaction (TLC, 95:5 CH₂Cl_{2⁻}acetone) it was diluted with CH₂Cl₂ (10 mL), washed with satd NaHCO₃ $(2 \times 10 \text{ mL})$ and water $(1 \times 10 \text{ mL})$. After drying the solvent was removed under diminished pressure. The crude product was crystallized from EtOH to give 0.97 g (93%) of **2** as a white crystalline product. Mp: 119–120 °C, lit.¹⁵ mp: 118 °C; $[\alpha]_D \pm 29$ (*c* 0.38, CHCl₃), (lit.¹⁵ $[\alpha]_D \pm 11$ (*c* 1.49, CHCl₃)); ¹H NMR (CDCl₃, 360 MHz): δ (ppm) 5.27, 5.12, 5.10 (pseudo t, 3H, $J \sim 9.2$ Hz in each, H-2, H-3, H-4), 5.26 (d, 1H, J_{1,2} 9.2 Hz, H-1), 4.24 (dd, 1H, J_{&6'} 11.9, J_{5,6} 4.0 Hz, H-6), 4.10 (dd, 1H, J_{&6'} 11.9, J_{×6'} 2.6 Hz, H-6'), 3.86 (ddd, 1H, *J*_{4,5} 9.2, *J*_{5,6} 4.0, *J*_{5,6} 2.6 Hz, H-5), 2.39 (s, 3H, SCOCH₃), 2.08, 2.03, 2.02, 2.01 (12H, $\frac{4}{5}$, $4 \times \text{OCOCH}_3$); ¹³C NMR (CDCl₃, 90 MHz): δ (ppm) 191.8 (SCOCH₃), 170.5, 169.2, 169.3 (2) (CO), 80.1 (C-1), 76.3, 73.8, 68.9, 67.8 (C-2 to C-5), 61.6 (C-6), 30.7 (SCOCH₃) 20.6, 20.5 (3) (CH₃).

3.3. Sodium 2,3,4,6-tetra-O-acetyl- β -D-glucopyranosyl-1-C-sulfonate (adapted from¹⁴) (4)

(a) Compound **2** (0.50 g, 1.23 mmol) was suspended in glacial acetic acid (10 mL), and Oxone® (1.89 g, 3.07 mmol) and NaOAc (1.34 g, 12.3 mmol) were added. The mixture was stirred at rt until TLC (4:1 CH₂Cl₂-MeOH) showed complete transformation of the starting material. After filtration the solvent was removed under diminished pressure. The obtained crude product was purified by column chromatography (eluent, 4:1 CHCl₃, MeOH) to give 0.42 g (70%) of **4** as a white crystalline product. Mp: 218–220 °C; $[\alpha]_D$ -59 (*c* 0.23, H₂O); ¹H NMR (D₂O, 360 MHz): δ (ppm) 5.43, 5.37, 5.17 (pseudo t, 3H, J ~ 9.4 Hz in each, H-2, H-3, H-4), 4.57 (d, 1H, J_{1,2} 9.4 Hz, H-1), 4.43 (dd, 1H, J_{6,6} 11.7, J_{5,6} 2.1 Hz, H-6), 4.27 (dd, 1H, $J_{5,6'}$ 11.7, $J_{5,6'}$ 1.0 Hz, H-6'), 4.14 (ddd, 1H, $J_{4,5}$ 9.4, $J_{5,6}$ 2.1, $J_{5,6'}$ 1.0 Hz, H-5), 2.15, 2.12, 2.11, 2.09 (12H, 4s, 4 × OCOCH₃); ¹³C NMR (D₂O, 90 MHz): δ (ppm) 174.2, 173.6, 173.2, 172.8 (CO), 85.5 (C-1), 75.7, 74.7, 69.5, 68.5 (C-2 to C-5), 62.5 (C-6), 21.9, 20.8, 20.7, 20.6 (CH₃); ESI MS calcd for C₁₄H₁₉O₁₂SNa 434.354, found 457.018 (M+Na).

(b) Also prepared from 2,3,4,6-tetra-0-acetyl-1-thio- β -D-gluco-pyranose¹⁷ (**3**, 0.50 g, 1.31 mmol) by using the same procedure. Yield: 0.52 g (87%).

3.4. Methyl 3-(2,3,4,6-tetra-O-acetyl-β-Dglucopyranosylthio)propanoate (5)

Compound 1 (5.00 g, 12.8 mmol) was dissolved in dry CH₂Cl₂ 280 (30 mL), then methyl 3-sulfanylpropanoate (2.1 mL, 19.2 mmol) and BF₃·OEt₂ (2.7 mL, 25.6 mmol) were added at 0 °C. The soln was stirred overnight (~16 h) at rt, (TLC, 1:2 EtOAc-hexane). After complete transformation of the starting material, the mixture was diluted with CH_2Cl_2 and washed with satd NaHCO₃ (2 × 15 mL) and water $(1 \times 15 \text{ mL})$, and then dried. The solvent was removed under diminished pressure and the crude product crystallized from EtOH to give 2.50 g (43%) of **5** as white needles. Mp: $86-87 \circ C$ (lit.²² mp: 88–89 °C); $[\alpha]_D^{\wedge}$ –64 (c 0.26, CHCl₃ (lit.²² $[\alpha]_D$ –32 (c 1.00, CHCl₃)); ¹H NMR (CDCl₃, 360 MHz): δ (ppm) 5.23, 5.03, 5.01 (pseudo t, 3H, J ~ 9.2 Hz in each, H-2, H-3, H-4), 4.56 (d, 1H, J_{1,2} 10.6 Hz, 290 H-1), 4.23 (dd, 1H, *J*_{6,6'} 11.9, *J*_{5,6} 5.3 Hz, H-6), 4.14 (dd, 1H, *J*_{6,6'} 11.9, J_{5,6} 2.6 Hz, H-6'), 3.74 (ddd, 1H, J_{4,5} 9.2, J_{5,6} 5.3, J_{5,6} 2.6 Hz, H-5), 3.69 (s, 3H, OCH₃), 2. 99 (dt, 1H, J 7.9, J 6.6 Hz, CH₂), 2.88 (dt, 1H, J 7.9, J 6.6 Hz, CH₂), 2.69 (t, 2H, J 7.9, J 6.6 Hz, CH₂), 2.09, 2.06, 2.03, 2.01 (12H, 4s, $4 \times OCOCH_3$); ¹³C NMR (CDCl₃, 90 MHz): δ (ppm) 171.9 (COOCH₃), 170.5, 169.9, 169.2 (2) (CO), 83.7 (C-1), 75.7, 73.6, 69.5, 68.1 (C-2 to C-5), 61.9 (C-6), 51.6 (OCH₃), 35.1, 25.2 (CH₂), 20.5, 20.4 (2), 20.3 (CH₃). Anal. Calcd for C₁₈H₂₆O₁₁S₁ (450.47): C, 47.99; H, 5.82. Found: C, 47.86; H, 5.66.

3.5. Methyl 3-(β-D-glucopyranosylthio)propanoate (6)

Thioglucoside 5 (0.18 g, 0.41 mmol) was dissolved in dry MeOH (5 mL), and NaOMe (\sim 1 M in MeOH) was added. The reaction mixture was kept at *t* until completion of the transformation (TLC, 1:1 CHCl₃₇MeOH). Amberlyst 15 (H⁺ form) was then added to remove sodium ions, the resin was filtered off, and the solvent was evaporated to give 0.11 g (96%) of **6** as a colourless oil. $R_{\rm f}$ = 0.73 (1:1 CHCl_{3 $\overline{\}}MeOH$); [α]_D $\overline{\}$ 74 (c 0.19, H₂O); ¹H NMR (D₂O, 360 MHz):</sub> δ (ppm) 4.51 (dd, 1H, $J_{\beta,6}$ 13.2, $J_{5,6}$ 4.0 Hz, H-6), 3.85 (dd, 1H, $J_{\beta,6}$ 13.2, *J*_{5.6'} 1.2 Hz, H-6'), 3.68 (s, 3H, OCH₃), 3.64 (d, 1H, *J*_{1,2} 9.2 Hz, H-1), 3.44-3.23 (m, 3H, H-2, H-3, H-4), 3.26 (ddd, 1H, J_{4,5} 9.2, J_{5,6} 4.0, J₅₆ 1.2 Hz, H-5), 2.99 (quintet, 1H, J 7.9, J 6.6 Hz, CH₂), 2.93 (quintet, 1H, J 7.9, J 6.6 Hz, CH₂), 2.76–2.74 (m, 2H, CH₂); ¹³C NMR (D₂O, 90 MHz). δ (ppm) 175.7 (COOCH₃), 86.5 (C-1), 80.4, 77.4, 72.7, 70.0, (C-2 to C-5), 61.5 (C-6), 53.5 (OCH₃), 35.5, 25.7 (CH₂). Anal. <u>Calcd</u> for C₁₀H₁₈O₇S₁ (282.31): <u>C</u>, 42.55; H, 6.43. Found: **Ç**, 42.50; H, 6.48.

3.6. Methyl 3-(2,3,4,6-tetra-O-acetyl-β-Dglucopyranosylthio)propanoate S-oxide (10)

Thioglucoside 5 (0.1 g, 0.22 mmol) was dissolved in MeOH (3 mL). To this soln Na_2WO_4 (6.5 mg, 0.02 mmol) and H_2O_2 (0.1 mL, 3.30 mmol) were added in small portions. The mixture was stirred at rt until completion of the transformation (TLC, 1:1 EtOAc-hexane). After filtration the solvent was removed under diminished pressure. The obtained oil was purified by column chromatography (2:1 EtOAc-hexane) to give 0.04 g (39%) of 10 as a white powder. Mp: 115–117 °C; $[\alpha]_D = -128 (c 0.22), CHCl_3); {}^{1}H$ NMR (CDCl₃, 360 MHz): δ (ppm) 5.34, 5.26, 5.11 (pseudo t, 3H, $J \sim 9.2$ Hz in each, H-2, H-3, H-4), 4.37 (d, 1H, $J_{1,2}$ 9.2 Hz, H-1), 4.29 (dd, 1H, J_{6,6'} 13.2, J_{5,6} 5.3 Hz, H-6), 4.21 (dd, 1H, J_{6,6'} 13.2, J_{5,6} 2.6 Hz, H-6'), 3.82 (ddd, 1H, J_{4,5} 9.2, J_{5,6} 5.3, J_{5,6'} 2.6 Hz, H-5), 3.73 (s, 3H, OCH₃), 3.22-3.20 (m, 2H, CH₂), 2.85 (m, 2H, CH₂), 2.09, 2.07, 2.05, 2.03 (12H, $\frac{4s}{4s}$, $4 \times \text{OCOCH}_3$); ¹³C NMR (CDCl₃, 90 MHz): δ (ppm) 171.5 (COOCH₃), 170.4, 169.9, 169.5, 169.2 (CO), 90.1 (C-1), 76.8, 73.0, 68.3, 67.5 (C-2 to C-5), 61.3 (C-6), 52.2 (OCH₃), 42.0, 26.1 (CH₂), 20.5 (2), 20.4 (2) (CH₃). Anal. Calcd for C₁₈H₂₆O₁₂S₁ (466.46): C, 46.35; H, 5.62. Found: C, 46.15; H, 5.55.

3.7. Methyl 3-(2,3,4,6-tetra-O-acetyl-β-Dglucopyranosylthio)propanoate <u>S</u>,S-dioxide (11)

(a) Thioglucoside **5** (0.1 g, 0.22 mmol) was dissolved in glacial acetic acid (3 mL). To this soln, Oxone[®] (0.19 g, 0.33 mmol) and NaOAc (0.14 g, 2.20 mmol) were added. The mixture was stirred at rt until complete transformation of the starting material (TLC, 1:1 EtOAc-hexane). After filtration it was diluted with water (10 mL) and washed with CH₂Cl₂ (3 × 5 mL). The organic phase was washed with satd NaHCO₃ (2 × 5 mL) and water (1 × 5 mL), dried, and the solvent removed under diminished pressure. The crude product was recrystallized from Et₂O to give 0.07 g (70%) of **11**.

(b) Thioglucoside **5** (0.1 g, 0.22 mmol) was suspended in CH_2CI_2 (3 mL), and NaHCO₃ (0.19 g, 2.20 mmol) and *mCPBA* (0.22 g, 15.8 mmol, ~70%) were added in five portions at 0 °C. The mixture was stirred at rt until complete transformation of the starting material (TLC, \therefore 1 EtOAc-hexane). After filtration it was diluted

340

350

12 November 2008 Disk Used

6

370

390

410

with water (10 mL) and washed with CH_2Cl_2 (2 × 5 mL). The organic phase was washed with NaCl $(1 \times 5 \text{ mL})$, satd NaHCO₃ $(2 \times 5 \text{ mL})$ and water $(1 \times 5 \text{ mL})$, dried, and the solvent removed under diminished pressure. The crude product was recrystallized from Et_2O to yield 0.08 g (72%) of **11** as white crystalline product. Mp: 146–147 °C; $[\alpha]_D$ $\overline{\ }^{36}$ (*c* 0.22, CHCl₃); ¹H NMR (CDCl₃, 360 MHz): δ (ppm) 5.52, 5.33, 5.14 (pseudo t, 3H, $J \sim$ 9.3 Hz in each, H-2, H-3, H-4), 4.56 (d, 1H, J_{1,2} 9.2 Hz, H-1), 4.28 (dd, 1H, J_{6,6} 11.9, J_{5,6} 5.3 Hz, H-6), 4.21 (dd, 1H, J_{6,6} 11.9, J_{5,6} 2.6 Hz, H-6'), 3.88 (ddd, 1H, J_{4,5} 9.3, J_{5,6} 5.3, J_{5,6'} 2.6 Hz, H-5), 3.75 (s, 3H, OCH₃), 3.60 (dt, 1H, J 7.9, 6.6 Hz, CH₂), 3.41 (dt, 1H, J 7.9, 6.6 Hz, CH₂), 2.89 (m, 2H, CH₂), 2.10, 2.05 (2), 2.03 (12H, 4s, $4 \times 0COCH_3$); ¹³C NMR (CDCl₃, 90 MHz): δ (ppm) 170.5 (COOCH₃), 170.3, 169.9, 169.2, 169.1 (CO), 87.9 (C-1), 76.6, 72.9, 67.2, 66.2 (C-2 to C-5), 61.2 (C-6), 52.3 (OCH₃), 45.3, 26.2 (CH₂), 20.4 (2), 20.3 (2) (CH₃). Anal. <u>Calcd</u> for C₁₈H₂₆O₁₃S₁ (482.46): <u>C</u>, 44.81; H, 5.43. Found: <u>C</u>, 44.78; H, 5.40.

3.8. Methyl 3-(β-D-glucopyranosylthio)propanoate S.S-dioxide (12)

Sulfone **11** (0.1 g, 0.21 mmol) was suspended in a mixture of dry MeOH (3 mL) and dry CHCl₃ (2 mL). To this soln two drops of AcCl were added. The mixture was kept at rt until completion of the transformation (~one week, TLC, 4:1 CHCl₃–MeOH). It was then neutralized with solid NaHCO₃ and the solvent evaporated after filtration. The crude oil was purified by column chromatography (4:1 CHCl₃–MeOH) to give 0.05 g (75%) of **12** as a colourless oil. $R_f = 0.37$ (4:1 CHCl₃–MeOH) to give 0.05 g (75%) of **12** as a colourless oil. $R_f = 0.37$ (4:1 CHCl₃–MeOH); [α]_D –53 (*c* 0.26, H₂O); ¹H NMR (D₂O, 360 MHz): δ (ppm) 4.68 (d, 1H, $J_{1,2}$ 9.2 Hz, H-1), 3.90 (dd, 1H, $J_{\beta,6'}$ 13.2, $J_{5,6}$ 5.3 Hz, H-6), 3.85 (s, 3H, OCH₃), 3.83 (dd, 1H, $J_{\beta,6'}$ 13.2, $J_{5,6'}$ 2.6 Hz, H-6'), 3.78–3.36 (m, 6H, H-2, H-3, H-4, H-5, CH₂), 2.98–2.92 (m, 2H, CH₂); ¹³C NMR (D₂O, 90 MHz): δ (ppm) 173.8 (COOCH₃), 90.0 (C-1), 81.3, 77.2, 69.2, 69.1, (C-2 to C-5), 61.0 (C-6), 53.4 (OCH₃), 47.1, 27.1 (CH₂). Anal. Calcd for C₁₀H₁₈O₉S₁ (314.31): C, 38.21; H, 5.77. Found: C, 38.25; H, 5.80.

3.9. Mixed salt of 2,3,4,6-tetra-O-acetyl-β-D-glucopyranosyl-1-Csulfinic acid (13)

Sulfone 11 (0.1 g, 0.21 mmol) was dissolved in freshly distilled dry CHCl₃ (10 mL), and DBU (30 µL, 0.21 mmol) was added. The mixture was stirred at rt until complete transformation of the starting material (TLC, 7:3 CHCl₃-MeOH). The solvent was evaporated and the crude oil was purified by column chromatography (7:3 CHCl₃–MeOH) to give 0.06 g of **13** as a white crystalline product. ¹H NMR (D₂O, 360 MHz): δ (ppm) 5.36, 5.29, 5.08 (pseudo t, 3H, $J \sim 9.2$ Hz in each, H-2, H-3, H-4), 4.38 (dd, 1H, $J_{6,6}$ 11.9, $J_{5,6}$ 2.6 Hz, H-6), 4.27 (dd, 1H, $J_{6,6'}$ 11.9, $J_{5,6'}$ 1.0 Hz, H-6'), 4.00 (ddd, 1H, $J_{4,5}$ 9.2, $J_{5,6}$ 2.6, $J_{5,6'}$ 1.0 Hz, H-5), 3.66 (d, 1H, $J_{1,2}$ 9.2 Hz, H-1), 2.10, 2.05, 2.04 (2) (12H, 3s, $4 \times OCOCH_3$); ¹³C NMR (D₂O, 90 MHz): δ (ppm) 174.3, 173.9, 173.3 (2) (CO), 92.8 (C-1), 75.8, 75.2, 68.7, 68.5 (C-2 to C-5), 62.5 (C-6), 20.9, 20.7, 20.6 (2) (CH₃); Ms: [M+Na]⁺ 419.059, [M+K]⁺ 435.035. Repeated from 1.5 g sulfone. Yield: 1.35 g (~100%). For further preparative purposes, the crude product obtained after solvent removal was used without purification.

3.10. 2,3,4,6-Tetra-O-acetyl-β-D-glucopyranosyl-1-C-sulfonamide (14)

Sulfinate **13** (0.1 g) was suspended in water (5 mL). To this soln, H₂NOSO₃H (0.08 g, 0.71 mmol) and NaOAc (0.06 g, 0.73 mmol) were added and the mixture was stirred at rt for 2 h. After extraction with EtOAc, (5 × 5 mL) the organic phase was washed with satd NaHCO₃ (2 × 5 mL) and water (5 mL), dried and evaporated. **J**t was crystallized slowly to give 0.07 g (\$1%, from the sulfone, 0.1 g) of **14** as a white crystalline product. Mp: 201–204 °C; $[\chi]_D$ –20 (*c* 0.2, CHCl₃); ¹H NMR (CDCl₃, 360 MHz): δ (ppm) 5.39, 5.31, 5.14 (pseudo t, 3H, $J \sim 9.3$ Hz in each, H-2, H-3, H-4), 4.56 (d, 1H, $J_{1,2}$ 9.2 Hz, H-1), 4.36 (dd, 1H, $J_{𝔅𝔅'}$ 13.2, $J_{5.6}$ 5.3 Hz, H-6), 4.46 (dd, 1H, $J_{𝔅𝔅'}$ 13.2, $J_{𝔅𝔅'}$ 1.0 Hz, 𝓜-6'), 3.93–3.86 (m, 3H, H-5, NH₂), 2.11, 2.08, 2.06, 2.04 (12H, 𝔄𝔅𝔅i 4 × OCOCH₃); ¹³C NMR (CDCl₃, 90 MHz): δ (ppm) 170.9, 170.4, 170.1, 169.5 (CO), 87.0 (C-1), 75.9, 72.9, 67.8, 67.5 (C-2 to C-5), 61.3 (C-6), 20.2 (2), 20.1 (2) (CH₃). Anal. Calcd for C₁₄H₂₁O₁₁NS (411.39): C, 40.88; H, 5.15. Found: C, 40.82; H, 5.05. Ms: [M+Na]⁺ 434.081. Repeated from 1.5 sulfone. Yield: 1.03 g (\$1%).

3.11. β-D-Glucopyranosyl-1-C-sulfonamide (15)

Sulfonamide **14** (0.1 g, 0.24 mmol) was suspended in dry MeOH (3 mL). A few drops of NH₄OH were added and the mixture was kept at rt until completion of the transformation (TLC, **4**:1 CHCl₃, MeOH). Amberlyst 15 (H⁺ form) was then added to remove sodium ions, the resin was filtered off and the solvent removed under diminished pressure. The crude oil was purified by column chromatography (**4**:1 CHCl₃, MeOH) to give 0.05 g (**85**%) of **15** as a colourless oil. $R_f = 0.59$ (**1**:1 CHCl₃, MeOH); $[\alpha]_D$, **34** (*c* 0.24, H₂O); ¹H NMR (D₂O, 360 MHz): δ (ppm) 4.46 (d, 1H, $J_{1,2}$ 9.2 Hz, H-1), 3.91 (dd, 1H, $J_{6,6}$ '11.9, $J_{5,6}$ 5.3 Hz, H-6), 3.74–3.54 (m, 4H, H-2, H-3 or H-4, H-5, H-6') 3.43 (t, 1H, J 9.2, J 9.2 Hz, H-4 or H-3); ¹³C NMR (D₂O, 90 MHz): δ (ppm) 90.5 (C-1), 80.9, 77.1, 70.8, 69.5, (C-2 to C-5), 61.2 (C-6); Anal. Calcd for C₆H₁₃O₇S₁ (243.24): **C**, **29.60**; H, 5.39. Found: **C**, **29.65**; H, 5.36.

3.12. Methyl 3-(2,3,4,6-tetra-O-acetyl-α-Dgalactopyranosylthio)propanoate (18)

1,2,3,4,6-Penta- $O_{acetyl-\beta-D-galactopyranose}$ (**16**, 5.0 g, 12.8 mmol) was dissolved in dry CH₂Cl₂ (30 mL), and methyl 3-sulfanylpropanoate (2.1 mL, 19.2 mmol) and BF₃·OEt₂ (2.7 mL, 25.6 mmol) were added at 0 °C. The soln was stirred overnight (~ 16 h) at rt (TLC, 1:2 EtOAc-hexane). After complete transformation of the starting material the mixture was diluted with CH₂Cl₂ and washed with satd NaHCO₃ (2×15 mL) and water (1×15 mL), dried, and the solvent removed under diminished pressure. The crude product was purified by silica gel column chromatography (1:2 EtOAc-hexane) EtOH to give 2.00 g (35%) of 18 as a colourless oil. $R_{f} = 0.16$ (1:2 EtOAc-hexane), $[\alpha]_{D}$ +496 (c 0.34, CHCl₃); ¹H NMR (CDCl₃, 360 MHz): δ (ppm) 5.76 (d, 1H, $J_{1,2}$ 5.3 Hz, H-1), 5.45 (dd, 1H, J_{3.4} 2.6, J_{4.5} 1.0 Hz, H-4), 5.26 (dd, 1H, J_{2.3} 10.6, J_{1.2} 5.3 Hz, H-2), 5.18 (dd, 1H, J_{2,3} 10.6, J_{3,4} 2.6 Hz, H-3), 4.58 (t, 1H, J_{5,6} 6.6, J_{5,6'} 5.8 Hz, H-5), 4.15–4.09 (m, 2H, H-6, H-6'), 3.71 (s, 3H, OCH₃), 2.88 (dt, 1H, J 7.9, J 6.6 Hz, CH₂), 2.78 (dt, 1H, J 7.9, J 6.6 Hz, CH₂), 2.68–2.64 (m, 2H, CH₂), 2.15, 2.06 (2), 1.96 (12H, 3s, $4 \times \text{OCOCH}_3$); ¹³C NMR (CDCl₃, 90 MHz): δ (ppm) 171.8 (COOCH₃), 170.3, 170.1 (2), 169.7 (CO), 89.9 (C-1), 67.9, 67.8, 67.7, 66.6 (C-2 to C-5), 61.8 (C-6), 51.7 (OCH₃), 34.4, 25.1 (CH₂), 20.6, 20.5 (3) (CH₃); Anal. Calcd for C₁₈H₂₆O₁₁S₁ (450.47): C, 47.99; H, 5.82. Found: C, 47.89; H, 5.76.

3.13. Methyl 3-(2,3,4,6-tetra-O-acetyl-α-Dgalactopyranosylthio)propanoate S,S-dioxide (19)

Thiogalactoside **18** (0.5 g, 1.10 mmol) was dissolved in glacial acetic acid (15 mL). To this soln, Oxone[®] (0.95 g, 1.65 mmol) and NaOAc (0.60 g, 11.0 mmol) were added. The mixture was stirred at rt until complete transformation of the starting material (TLC, 1:1 EtOAc-hexane). After filtration it was diluted with water (15 mL) and extracted with CH₂Cl₂ (3 × 15 mL). The organic phase was washed with satd NaHCO₃ (2 × 15 mL) and water (1 × 15 mL),

470

510

530

12 November 2008 Disk Used

7

540

550

K. Czifrák, L. Somsák/Carbohydrate Research xxx (2008) xxx-xxx

dried, and the solvent removed under diminished pressure. The crude product was purified by column chromatography (1:1 EtOAc-hexane) to give 0.48 g (90%) of **19** as a colourless oil. $R_f = 0.36$ (1:1 EtOAc-hexane), [α]_D +33 (c 0.25, CHCl₃); ¹H NMR (CDCl₃, 360 MHz): δ (ppm) 5.79 (dd, 1H, $J_{2,3}$ 10.6, $J_{3,4}$ 2.6 Hz, H-3), 5.56 (dd, 1H, $J_{3,4}$ 2.6, $J_{4,5}$ 1.0 Hz, H-4), 5.47 (dd, 1H, $J_{2,3}$ 10.6, $J_{1,2}$ 2.6 Hz, H-2), 5.36 (d, 1H, $J_{1,2}$ 2.6 Hz, H-1), 4.81 (t, 1H, $J_{5,6}$ 6.6, $J_{4,6}$ (5.8 Hz, H-5), 4.19–4.06 (m, 2H, H-6, H-6'), 3.62 (s, 3H, OCH₃), 3.49–3.44 (m, 2H, CH₂), 2.93–2.85 (m, 2H, CH₂), 2.16, 2.13, 2.09, 2.01 (12H, 4s, 4 × OCOCH₃); ¹³C NMR (CDCl₃, 90 MHz): δ (ppm) 170.6 (COOCH₃), 170.3, 170.0, 169.5, 169.2 (CO), 84.8 (C-1), 71.3, 66.9, 66.7, 65.7 (C-2 to C-5), 62.2 (C-6), 52.3 (OCH₃), 46.5, 25.8 (CH₂), 20.4, 20.2 (3) (CH₃); Anal. Calcd for C₁₈H₂₆O₁₃S₁ (482.46): C, 44.81; H, 5.43. Found: C, 44.86; H, 5.38.

3.14. Methyl 3-(2,3,4,6-tetra-O<u>acetyl-β-D</u>-galactopyranosylthio)propanoate (20)

A 2,3,4,6-tetra- $O_{acetyl-1-thio-\beta-D-galactopyranose^{25,26}}$ (17, 3.0 g, 7.86 mmol) was dissolved in dry CH₂Cl₂ (50 mL). To this soln methyl 3-bromopropanoate (1.11 mL, 9.43 mmol) and Et₃N (1.05 mL, 9.43 mmol) were added. The mixture was stirred at rt for 3 days (TLC, 1:2 EtOAc-hexane). It was then diluted with CH_2Cl_2 and washed with satd NaHCO₃ (2 × 50 mL) and water $(1 \times 50 \text{ mL})$, dried, and the solvent evaporated. The crude product was purified by column chromatography (1:2 EtOAc-hexane) to give 2.10 g (85%) of 20 (conversion is 65) as a yellowish oil. $R_{\rm f} = 0.18 \ (1:2 \ \text{EtOAc-hexane}); \ [\alpha]_{\rm D} + 169 \ (c \ 0.42, \ \text{CHCl}_3); \ ^1\text{H NMR}$ (CDCl₃, 360 MHz): δ (ppm) 5.40 (dd, 1H, $J_{3,4}$ 2.6, $J_{4,5}$ 1.0 Hz, H-4) 5.23 (pseudo t, 1H, $J_{2,3}$ 10.6, $J_{1,2}$ 9.2 Hz, H-2), 5.05 (dd, 1H, $J_{2,3}$ 10.6, J_{3,4} 2.6 Hz, H-3) 4.54 (d, 1H, J_{1,2} 9.2 Hz, H-1), 4.16 (dd, 1H, J_{§,6'} 11.9, J_{5,6} 5.3 Hz, H-6), 4.12 (dd, 1H, J_{§,6'} 11.9, J_{5,6'} 2.6 Hz, H-6'), 3.95 (t, 1H, J_{5.6} 6.6, J_{5.6} 5.8 Hz, H-5), 3.70 (s, 3H, OCH₃), 3.02 (dt, 1H, J 7.9, J 6.6 Hz, CH₂), 2.93 (dt, 1H, J 7.9, J 6.6 Hz, CH₂), 2.72-2.68 (m, 2H, CH₂), 2.16, 2.05 (2), 1.98 (12H, 3s, 4 OCOCH₃); ¹³C NMR (CDCl₃, 90 MHz): δ (ppm) 172.0 (COOCH₃), 170.3, 170.1, 169.9, 169.7 (CO), 84.5 (C-1), 74.4, 71.7, 67.1, 67.0 (C-2 to C-5), 61.4 (C-6), 51.7 (OCH₃), 35.1, 25.4 (CH₂), 20.6, 20.5 (2), 20.4 (CH₃); Anal. Calcd for C₁₈H₂₆O₁₁S₁ (450.47): C, 47.99; H, 5.82. Found: C, 47.85; H, 5.92.

3.15. Methyl 3-(2,3,4,6-tetra-O-acetyl-β-Dgalactopyranosylthio)propanoate *S*,*S*-dioxide (21)

Thiogalactoside 20 (1.0 g, 2.20 mmol) was dissolved in glacial acetic acid (30 mL). To this soln, Oxone[®] (1.90 g, 3.30 mmol) and NaOAc (1.20 g, 22.0 mmol) were added. The mixture was stirred at rt until complete transformation of the starting material (TLC, 1:1 EtOAc-hexane). After filtration it was diluted with water (30 mL) and extracted with CH₂Cl₂ (3 × 30 mL). The organic phase was washed with satd NaHCO₃ (2×30 mL) and water (1×30 mL), dried, and the solvent was removed under diminished pressure. The crude product was recrystallized from Et₂O to give 0.74 g (70%) of **21** as a white crystalline product. Mp: 120–122 °C; $[\alpha]_D$ +6 (c 0.24, CHCl₃); ¹H NMR (CDCl₃, 360 MHz): δ (ppm) 5.68 (1H, pséudo t, J_{2,3} 10.6, J_{1,2} 9.2 Hz, H-2), 5.47 (dd, 1H, J_{3,4} 4.0, J_{4,5} 2.6 Hz, H-4) 5.16 (dd, 1H, J_{2,3} 10.6, J_{3,4} 4.0 Hz, H-3), 4.52 (d, 1H, $J_{1,2}$ 9.2 Hz, H-1), 4.23–4.16 (m, 2H, H-5, H-6), 4.11 (dd, 1H, $J_{6,6'}$ 11.9, J_{5,6'} 5.3 Hz, H-6'), 3.74 (s, 3H, OCH₃), 3.55 (dt, 1H, J 7.9, J 6.6 Hz, CH₂), 3.42 (dt, 1H, J 7.9, J 6.6 Hz, CH₂), 2.92–2.87 (m, 2H, CH₂), 2.20, 2.07, 2.06, 1.98 (12H, 4s, 4 × OCOCH₃); ¹³C NMR (CDCl₃, 90 MHz): δ (ppm) 170.5 (COOCH₃), 170.2, 169.9, 169.8, 169.2 (CO), 88.6 (C-1), 75.4, 71.1, 66.6, 63.3 (C-2 to C-5), 61.0 (C-6), 52.3 (OCH₃), 45.1, 26.1 (CH₂), 20.6, 20.5, 20.4, 20.3 (CH₃); Anal. Calcd for C₁₈H₂₆O₁₃S₁ (482.46): C, 44.81; H, 5.43. Found: C, 44.74; H, 5.46.

3.16. Mixed salt of 2,3,4,6-tetra-O₋acetyl-β-D-galactopyranosyl-1-C-sulfinic acid (22)

Sulfone **21** (0.5 g, 1.05 mmol) was dissolved in freshly distilled dry CHCl₃ (15 mL) and DBU (150 µL, 1.05 mmol) was added. The mixture was stirred at rt until complete transformation of the starting material (TLC, **7**:3 CHCl₃, MeOH). The solvent was evaporated and the crude oil was purified by column chromatography (**7**:3 CHCl₃, MeOH) to give 0.35 g of **22** as a yellowish oil. ¹H NMR (D₂O, 360 MHz): δ (ppm) 5.47 (dd, 1H, $J_{3,4}$ 4.0, $J_{4,5}$ 1.0 Hz, H-4), 5.43 (t, 1H, $J_{1,2}$ 10.6, $J_{2,3}$ 10.6 Hz, H-2), 5.26 (dd, 1H, $J_{2,3}$ 10.6, $J_{3,4}$ 4.0 Hz, H-3), 3.64 (d, 1H, $J_{1,2}$ 10.6 Hz, H-1), 3.55–3.45 (m, 2H, H-6, H-6'), 3.29 (pseudo t, 1H, $J_{5,6}$ 6.6 Hz, H-5), 2.21, 2.06 (2), 2.00 (12H, **3**s, $4 \times \text{OCOCH}_3$); ¹³C NMR (D₂O, 90 MHz): δ (ppm) 173.6, 173.4, 173.0, 172.9 (CO), 92.8 (C-1), 74.4, 72.4, 68.3, 65.4 (C-2 to C-5), 62.1 (C-6), 20.4, 20.2, 20.1, 20.0 (CH₃). For further preparative purposes, the crude product obtained after solvent removal was used without purification.

3.17. 2,3,4,6-Tetra-O-acetyl-β-D-galactopyranosyl-1-C-sulfonamide (23)

Sulfinate salt **22** (\sim 0.35 g) was suspended in water (5 mL). H₂NOSO₃H (0.28 g, 2.5 mmol) and NaOAc (0.24 g, 2.92 mmol) were added and the mixture was stirred at rt for 2 h. After extraction with EtOAc (5 \times 10 mL), the organic phase was washed with satd NaHCO₃ $(2 \times 10 \text{ mL})$ and water (10 mL), dried, and the solvent evaporated to give 0.27 g (79%, from the sulfone, 0.4 g) of 23 as a yellowish oil. $R_f = 0.23$ (1:1 EtOAc-hexane); $[\alpha]_D + 32$ (c 0.22, Q1 CHCl₃); ¹H NMR (CDCl₃, 360 MHz): δ (ppm) 5.51 (s, 1H, NH₂), 5.48 (1H, dd, J_{3,4} 4.0, J_{4,5} 1.0 Hz, H-4), 5.29 (s, 1H, NH₂), 5.26 (t, 1H, J_{1,2} 10.6, J_{2,3} 10.6 Hz, H-2), 5.17 (dd, 1H, J_{2,3} 10.6, J_{3,4} 2.3 Hz, H-3), 4.41 (d, 1H, J_{1,2} 10.6 Hz, H-1), 4.22 (t, 1H, J_{5,6} 6.6, J_{5,6} 6.6 Hz, H-5), 4.16–4.11 (m, 2H, H-6, H-6'), 3.93–3.86 (m, 3H, H-5, NH₂), 2.17, 2.08, 2.05, 1.99 (12H, 4s, 4 × OCOCH₃); ¹³C NMR (CDCl₃, 90 MHz): δ (ppm) 171.1, 170.5, 170.0, 169.8 (CO), 87.8 (C-1), 75.2, 70.7, 66.9, 65.3 (C-2 to C-5), 60.9 (C-6), 20.8, 20.6 (2), 20.5 (CH₃). Anal. Calcd for C₁₄H₂₁O₁₁NS (411.39): C, 40.88; H, 5.15. Found: C, 40.80; H, 5.21.

3.18. Bromination methods

(a) A methyl 3-(2,3,4,6-tetra-O-acetyl- β -D-glycopyranosylthio)propanoate *S*,*S*-dioxide (**11** or **21**, 0.1 g, 0.21 mmol) was dissolved in dry CCl₄ (10 mL), and bromine (30 µL, 0.73 mmol) and some K₂CO₃ were added. The mixture was placed in an Erlenmeyer flask above a heat lamp (375 W, distance from the lamp ~2–3 cm, height of the soln 1–2 cm), and refluxed. It was monitored by (TLC, 1:1 EtOAc-hexane). After 3 h CHCl₃ (10 mL) was added, and the mixture washed with 1 M aq Na₂S₂O₃ (10 mL), satd NaHCO₃ (2 × 10 mL) and water (10 mL). After drying, the solvent was removed and the residue purified by column chromatography (1:1 EtOAc-hexane).

(b) A 2,3,4,6-tetra-O-acetyl- β -D-glycopyranosyl-1-C-sulfonamide (**14** or **23**, 0.1 g, 0.24 mmol) was dissolved in dry CHCl₃ (10 mL), and bromine (30 µL, 0.73 mmol) and some K₂CO₃ were added. The mixture was placed in an Erlenmeyer flask above a heat lamp (375 W, distance from the lamp ~2–3 cm, height of the soln 1–2 cm), and refluxed. If the mixture decolourized, bromine (10 µL) was added again. It was monitored by (TLC, **1**:1 EtOAc-hexane). After 8 h CHCl₃ (10 mL) was added, and the mixture washed with 1 M aq Na₂S₂O₃ (10 mL), satd NaHCO₃ (2 × 10 mL) and water (10 mL). After drying, the solvent was removed and the residue purified by column chromatography (**1**:1 EtOAc-hexane).

(c) 2,3,4,6-Tetra-O-acetyl- β -D-glycopyranosyl-1-C-sulfonamide (14, 0.1 g, 0.24 mmol) was dissolved in dry PhCF₃ (10 mL), and

Please cite this article in press as: Czifrák, K. Somsák, L., Carbohydr. Res. (2008), doi:10.1016/j.carres.2008.11.002

590

570

12 November 2008 Disk Used

660

670

680

690

8

610

630

K. Czifrák, L. Somsák/Carbohydrate Research xxx (2008) xxx-xxx

bromine (30 μ L, 0.73 mmol) and some K₂CO₃ were added. The mixture was placed in an Erlenmeyer flask above a heat lamp (375 W, distance from the lamp ~2–3 cm, height of the soln 1–2 cm), and refluxed. It was monitored by (TLC, 1:1 EtOAc-hexane). After 4 h, the mixture was washed with 1 M aq Na₂S₂O₃ (10 mL), satd NaH-CO₃ (2 × 10 mL) and water (10 mL). After drying, the solvent was removed under diminished pressure, and the residue purified by column chromatography (1:1 EtOAc-hexane).

3.19. Methyl (2,3,4,6-tetra-O-acetyl-1-C_pbromo-β-Dglucopyranosylthio)propanoate <u>S</u>S-dioxide (24)

Isolated from the bromination mixture of **11** (1.0 g, 2.07 mmol) obtained by method (a) as a white crystalline product (0.25 g, 21%) from hexane. Mp: 92–93 °C; $[\alpha]_D$ +176 (*c* 0.28, CHCl₃); ¹H NMR (CDCl₃, 360 MHz.): δ (ppm) 5.66 (d, 1H, $J_{1,2}$ 9.2 Hz, H-2), 5.49, 5.25 (pseudo t, 2H, $J \sim$ 9.2 Hz in each, H-3, H-4), 4.36–4.18 (m, 3H, H-5, H-6, H-6'), 3.75 (s, 3H, OCH₃), 3.72–3.63 (m, 2H, CH₂), 2.89–2.84 (m, 2H, CH₂), 2.12, 2.09, 2.06, 2.02 (12H, 4s, 4 × OCOCH₃); ¹³C NMR (CDCl₃, 90 MHz): δ (ppm) 170.3 (COOCH₃), 170.2, 169.6, 168.9, 168.6 (CO), 106.8 (C-1), 75.4, 71.3, 66.9, 66.2 (C-2 to C-5), 60.2 (C-6), 52.4 (OCH₃), 43.1, 26.3 (CH₂), 20.4 (3) 20.3 (CH₃). Anal. Calcd for C₁₈H₂₅O₁₃S₁ (561.36): C, 38.50; H, 4.49; S, 5.71. Found: C, 38.55; H, 4.42; S, 5.77.

3.20. Methyl (2,3,4,6-tetra-O-acetyl-5-C₂bromo-β-Dglucopyranosylthio)propanoate <u>S</u>,S-dioxide (25)

Isolated from the bromination mixture of **11** (1.0 g, 2.07 mmol) by method (a) as a colourless oil (0.35 g, **3**0%). $R_{\rm f}$ = 0.66 (1:1 EtOAchexane); [α]_D -252 (*c* 0.20, CHCl₃); ¹H NMR (CDCl₃, 360 MHz): δ (ppm) 5.66, 5.58 (pseudo t, 2H, $J \sim 9.2$ Hz in each, H-2, H-3), 5.26 (d, 1H, $J_{1,2}$ 10.6 Hz. H-1), 4.96 (d, 1H, $J_{3,4}$ 9.2 Hz, H-4), 4.58 (d, 1H, $J_{5,6'}$ 11.9 Hz, H-6), 4.46 (d, 1H, $J_{6,6'}$ 11.9 Hz, H-6'), 3.76 (s, 3H, OCH₃), 3.82–3.54 (m, 2H, CH₂), 2.88–2.84 (m, 2H, CH₂), 2.13, 2.10, 2.07, 2.03 (12H, 4s, 4 × OCOCH₃); ¹³C NMR (CDCl₃, 90 MHz): δ (ppm) 170.1 (COOCH₃), 169.6, 169.5, 168.9, 168.7 (CO), 98.3 (C-5), 85.5 (C-1), 71.2, 67.9, 66.9, (C-2 to C-4), 60.2 (C-6), 52.4 (OCH₃), 45.9, 26.1 (CH₂), 20.4 (2) 20.3 (2) (CH₃). Anal. Calcd for C₁₈H₂₅O₁₃S₁ (561.36): C, 38.50; H, 4.49. Found: C, 38.48; H, 4.45.

3.21. Methyl (2,3,4,6-tetra-*O*-acetyl-1-*C*-bromo-β-pgalactopyranosylthio)propanoate *S*,*S*-dioxide (26)

Isolated from the bromination mixture of **21** (0.20 g, 0.41 mmol) obtained by method (a) as a colourless oil (0.1 g, **43%**). $R_{\rm f} = 0.55$ (1:1 EtOAc–hexane); $[\alpha]_{\rm D}$ +154 (*c* 0.18, CHCl₃); ¹H NMR (CDCl₃, 360 MHz): δ (ppm) 5.82 (d, 1H, $J_{1,2}$ 10.6 Hz, H-2), 5.55 (dd,1H, $J_{3,4}$ 2.6, $J_{4,5}$ 1.0 Hz, H-4), 5.32 (dd, 1H, $J_{2,3}$ 10.6, $J_{3,4}$ 2.6 Hz, H-3), 4.52 (t, 1H, $J_{5,6}$ 6.6, $J_{5,6}$ 6.6 Hz, H-5), 4.26–4.22 (m, 2H, H-6, H-6'), 3.75 (s, 3H, OCH₃), 3.72–3.66 (m, 2Ĥ, CH₂), 2.96–2.92 (m, 2H, CH₂), 2.12, 2.07, 2.05, 2.00 (12H, 4s, 4 × OCOCH₃); ¹³C NMR (CDCl₃) δ (ppm): 170.2 (COOCH₃), 172.0, 170.1, 169.7, 169.6 (CO), 108.1 (C-1), 74.8, 69.6, 66.1, 63.8 (C-2 to C-5), 60.3 (C-6), 52.4 (OCH₃), 43.0, 26.3 (CH₂), 20.9 20.4 (3) (CH₃). Anal. Calcd for C₁₈H₂₅O₁₃S₁ (561.36): C, 38.50; H, 4.49; S, 5.71. Found: C, 38.53; H, 4.41; S, 5.77.

⁶⁵⁰ **3.22.** Methyl (2,3,4,6-tetra-O-acetyl-5-*C*-bromo-β-Dgalactopyranosylthio)propanoate *S,S*-dioxide (27)

Isolated from the bromination mixture of **21** (0.20 g, 0.41 mmol) obtained by method (a) as a colourless oil (0.06 g, 27%). $R_{\rm f} = 0.50$ (1:1 EtOAc-hexane); $[\alpha]_{\rm D} - 131$ (*c* 0.18, CHCl₃); ¹H NMR (CDCl₃, 360 MHz): δ (ppm) 5.80–5.70 (m, 2H, H-2, H-3),

5.63 (dd, 1H, $J_{3,4}$ 2.6, $J_{4,5}$ 1.0 Hz, H-4), 4.92 (d, 1H, $J_{1,2}$ 9.2 Hz. H-1), 4.67 (d, 1H, $J_{\beta,6'}$ 11.9 Hz, H-6), 4.40 (d, 1H, $J_{\beta,6'}$ 11.9 Hz, H-6'), 3.73 (3H, s, OCH₃), 3.46–3.40 (2H, m, CH₂), 2.89–2.82 (2H, m, CH₂), 2.15, 2.06 (2), 1.97 (12H, 3s, 4 × OCOCH₃); ¹³C NMR (CDCl₃, 90 MHz): δ (ppm) 170.2 (COOCH₃), 169.5 (2), 169.0 (2) (CO), 95.7 (C-5), 85.9 (C-1), 69.8, 68.5, 65.9, (C-2 to C-4), 60.3 (C-6), 52.5 (OCH₃), 45.8, 26.0 (CH₂), 20.9, 20.5, 20.4, 20.3 (CH₃). Anal. Calcd for C₁₈H₂₅O₁₃S₁ (561.36): C, 38.50; H, 4.49. Found: C, 38.59; H, 4.53.

3.23. 2,3,4,6-Tetra-O-acetyl-1-C-bromo-β-D-glucopyranosyl-1-C-sulfonamide (28)

Obtained by bromination of **14** (0.60 g, 1.45 mmol) by method (b) 0.05 g (11%) and method (c) 0.058 g (12%) as a white crystalline product from hexane. Mp: 185–187 °C; $[\alpha]_D$ +235 (c 0.26, CHCl₃); ¹H NMR (CDCl₃, 360 MHz): δ (ppm) 5.56–5.45 (m, 2H, H-2, H-3 or H-4), 5.38–5.34 (m, 2H, NH₂), 5.26 (t, 1H, *J* 9.2, *J* 9.2 Hz, H-4 or H-3), 4.33–4.26 (m, 3H, H-5, H-6, H-6'), 2.10 (2), 2.05, 2.01 (12H, 4s, 4 × OCOCH₃); ¹³C NMR (CDCl₃, 90 MHz): δ (ppm) 170.9, 169.8, 169.7, 169.3 (CO), 105.4 (C-1), 75.2, 71.5, 68.7, 66.6 (C-2 to C-5), 60.4 (C-6), 20.7 (2), 20.5, 20.4 (CH₃). Anal. Calcd for C₁₄H₂₀O₁₁BrNS (490.28): C, 34.30; H, 4.11. Found: C, 34.35; H, 4.01.

3.24. 2,3,4,6-Tetra-O-acetyl-5-C-bromo-β-D-glucopyranosyl-1-C-sulfonamide (29)

Obtained by bromination of **14** (0.60 g, 1.45 mmol) by method (b) 0.054 g (12%) and method (c) 0.056 g (12%) as a yellowish oil. $R_f = 0.59$ (1:1 EtOAc-hexane); $[\alpha]_D -284$ (c 0.24, CHCl₃); ¹H NMR (CDCl₃, 360 MHz): δ (ppm) 5.58, 5.51 (pseudo t, 2H, $J \sim 9.2$ Hz in each, H-2, H-3), 5.38–5.32 (m, 2H, NH₂), 5.23 (d, 1H, $J_{1,2}$ 9.2, Hz, H-1), 4.84 (d, 1H, $J_{3,4}$ 9.2 Hz, H-4), 4.90–4.56 (m, 2H, H-6, H-6'), 2.11, 2.08 (2), 2.01 (12H, 3s, $4 \times \text{OCOCH}_3$); ¹³C NMR (CDCl₃, 90 MHz): δ (ppm) 170.2 (2), 169.6, 169.0 (CO), 98.4 (C-5), 85.6 (C-1), 71.1, 68.2, 67.0 (C-2 to C-4), 65.5 (C-6), 20.6 (2), 20.4 (2) (CH₃). Anal. Calcd for C₁₄H₂₀O₁₁BrNS (490.28): C, 34.30; H, 4.11. Found: C, 34.28; H, 4.08.

Acknowledgements

Financial support for this work was provided by the Hungarian Scientific Research Fund (Grants: OTKA 46081 and 61336). The authors thank P. Gergely and T. Docsa for the glycogen phosphorylase assays.

References

- 1. Somsák, L.; Nagy, V.; Docsa, T.; Tóth, B.; Gergely, P. Tetrahedron: Asymmetry 2000, 11, 405–408.
- Somsák, L.; Nagy, V. Tetrahedron: Asymmetry **2000**, 11, 1719–1727. Corrigendum 2247.
- Somsák, L.; Kovács, L.; Tóth, M.; Ősz, E.; Szilágyi, L.; Györgydeák, Z.; Dinya, Z.; Docsa, T.; Tóth, B.; Gergely, P. J. Med. Chem. 2001, 44, 2843–2848.
- Czifrák, K.; Kovács, L.; Kövér, K. E.; Somsák, L. Carbohydr. Res. 2005, 340, 2328– 2334.
- Somsák, L; Nagy, V.; Hadady, Z.; Docsa, T.; Gergely, P. Curr. Pharm. Des. 2003, 9, 1177–1189.
- Somsák, L.; Nagy, V.; Hadady, Z.; Felföldi, N.; Docsa, T.; Gergely, P. In Frontiers in Medicinal Chemistry; Reitz, A. B., Kordik, C. P., Choudhary, M. I., Rahman, A. u., Eds.; Bentham, 2005; pp 253–272.
- 7. Oikonomakos, N. G.; Somsák, L. Curr. Opin. Invest. Drugs 2008, 9, 379–395.
- Somsák, L.; Czifrák, K.; Tóth, M.; Bokor, É.; Chrysina, E. D.; Alexacou, K. M.; Q2₇₁₀ Hayes, J. M.; Tiraidis, C.; Lazoura, E.; Leonidas, D. D.; Zographos, S. E.; Oikonomakos, N. G. *Curr. Med. Chem.*, in press.
- Hopfinger, A. J.; Reaka, A.; Venkatarangan, P.; Duca, J. S.; Wang, S. J. Chem. Inf. Comp. Sci. 1999, 39, 1151–1160.
- 10. Knapp, S.; Darout, E.; Amorelli, B. J. Org. Chem. 2006, 71, 1380-1389.
- 11. Lipták, A. In *Progress in Biological Chirality*; Pályi, G., Zucchi, C., Cagliotti, L., Eds.; Elsevier, 2004; pp 181–202.
- 12. Lipták, A.; Borbás, A. Magy. Kém. Foly. 2004, 109-110, 60-63.

12 November 2008 Disk Used

K. Czifrák, L. Somsák/Carbohydrate Research xxx (2008) xxx-xxx

9

740

- 13. Lipták, A.; Balla, E.; Jánossy, L.; Sajtos, F.; Szilágyi, L. Tetrahedron Lett. 2004, 45, 26
- 839–842. 14. Lipták, A.; Csávás, M.; Borbás, A. 2008, unpublished results.
- Elptak, A., Csavas, M., Borbas, A. 2008, unpublished result
 Bonner, W. A. J. Am. Chem. Soc. 1951, 73, 2659–2666.
- Johner, W. A. J. Ant. Chem. Soc. 1951, 75, 2039–2000.
 Sanhueza, C. A.; Dorta, R. L.; Vazquez, J. T. Tetrahedron: Asymmetry 2008, 19, 258–264.
- 17. Černý, M.; Vrkoč, J.; Staněk, J. Collect. Czech. Chem. Commun. 1959, 24, 64-69.
- 18. Horton, D. Meth. Carbohydr. Chem. **1963**, *2*, 433–437.
- 19. Graham, S. L.; Scholz, T. H. Synthesis 1986, 1031-1032.
- 20. Baskin, J. M.; Wang, Z. Tetrahedron Lett. 2002, 43, 8479-8483.
- 21. Chan, W. Y.; Berthelette, C. Tetrahedron Lett. 2002, 43, 4537–4540.
- 22. Sokolov, S. D.; Savochkina, L. P.; Kochetkov, N. K. Zh. Obsh. Khim. **1964**, 34, 4099–4103.
- Sridhar, P. R.; Prabhu, K. R.; Chandrasekaran, S. Eur. J. Org. Chem. 2004, 4809– 4815.
- Micskei, K.; Juhász, Z.; Ratkovic, Z.; Somsák, L. Tetrahedron Lett. 2006, 47, 6117– 6120.
- 25. Černý, M.; Staněk, J.; Pacák, J. Monatsh. Chem. 1963, 94, 290-294.

- Frgala, J.; Černý, M.; Staněk, J. Collect. Czech. Chem. Commun. 1975, 40, 1411– 1425.
- 27. Somsák, L.; Ferrier, R. J. Adv. Carbohydr. Chem. Biochem. 1991, 49, 37–92.
- 28. Praly, J. P.; Descotes, G. Tetrahedron Lett. 1990, 31, 1133-1136.
- Praly, J.-P.; Somsák, L.; Mahmoud, S. H.; El Kharraf, Z.; Descotes, G.; Farkas, I. J. Carbohydr. Chem. **1992**, *11*, 201–216.
- 30. Czifrák, K.; Somsák, L. Tetrahedron Lett. 2002, 43, 8849–8852.
- Merényi, R.; Janousek, Z.; Viehe, H. G. In Substituent Effects in Radical Chemistry; Viehe, H. G., Janousek, Z., Merényi, R., Eds.; Reidel, D.: Dordrecht, 1986; pp 301– 324.
- 32. Kiss, L.; Somsák, L. Carbohydr. Res. 1996, 291, 43-52.
- 33. Czifrák, K.; Szilágyi, P.; Somsák, L. Tetrahedron: Asymmetry 2005, 16, 127-141.
- 34. Praly, J.-P. Adv. Carbohydr. Chem. Biochem. 2001, 56, 65–151.
- Ősz, E.; Sós, E.; Somsák, L.; Szilágyi, L.; Dinya, Z. Tetrahedron 1997, 53, 5813– 5824.
- 36. Ősz, E.; Szilágyi, L.; Somsák, L.; Bényei, A. Tetrahedron **1999**, 55, 2419–2430.
- Watson, K. A.; Mitchell, E. P.; Johnson, L. N.; Son, J. C.; Bichard, C. J. F.; Orchard, M. G.; Fleet, G. W. J.; Oikonomakos, N. G.; Leonidas, D. D.; Kontou, M.; Papageorgiou, A. *Biochemistry* 1994, 33, 5745–5758.